Hazard assessment of *Staphylococcus* with positive coagulase in meat produced and distributed in the Northern regions of Cameroon

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Abstract

Aim: *Staphylococcus* with positive coagulase (SPC) is a major problem for beef consumers in the northern part of Cameroon. For this purpose, the SPC concentrations in beef produced and supplied in the northern regions were determined, as well as the resistance profile of these bacteria to antibiotics.

Materials and Methods: A total of 125 samples were obtained by the wet and dry swabbing method in traditional slaughterhouses and butcheries to evaluate the SPC concentration in meat, and then, 102 SPC isolations were collected to determine the antibiotic resistance profile.

Results: The distribution of concentration of the SPC indicated no significant differences of bacterial evidence in almost all the slaughterhouses except the one in Manwi (with 2.66 log CFU/cm²) and the density in SPC is higher than that one in Guider (1.99 log CFU/cm²). The assessment of density in SPC among the selected slaughterhouse highlighted a superiority of the SPC concentrations in the Ngaoundere butcheries (3.83 log CFU/cm²) in comparison with those of other towns. At the level of the slaughterhouses, a higher proportion of resistance to Penicillin G was recorded than Gentamicin. Some butcheries recorded that all SPC reacted to Kanamycin, whereas they were more resistant to Penicillin G.

Conclusion: These results reveal that the SPC found in meat poses a threat to meat consumers in the northern part of Cameroon.

Keywords: contamination, meat, Northern regions of Cameroon, resistance to antibiotics, *Staphylococcus* with positive coagulase.

Introduction

The food security of beef is a major concern in Sub-Saharan Africa, including Cameroon [1], wherein beef is the main source of animal proteins for human consumption. Beef is also considered as a high-risk food product because it harbors some pathogenic microorganisms [2]. In general, human microflora originates from various food intakes [3] with some bacteria which alone represent the twothirds of total microflora [4]. Among other infectious microbes, *Staphylococcus* are known as the main cause of toxi-infections among humans of all age groups [4]. Several *Staphylococcus* intoxication cases were reported in the world. A clear example is a case study carried out in La Loire between 1996 and 2013, wherein members of the 767 households were

Copyright: Afnabi, *et al.* Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/ publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated. infected, 211 cases are caused by *Staphylococcus* [4], and the other study carried out between 2011 and 2012 where the *Staphylococcus* play a great part with the bacteria responsible for food intoxication in Morocco with 31% [5]. *Staphylococcus* resistance to antibiotics is another emerging problem. Many cases of *Staphylococcus* resistance to antibiotic are reported throughout the world, namely, methicillin resistance found in 264 of examined meat samples, for example, 11.9% of the 2217 samples chosen most of which came from veal meat (15.2%) and beef (10.6%) [6].

In other studies carried out in the USA over 136 samples of meat and poultry, 96% of isolated *Staphylococcus aureus* were resistant to at least one type of antibiotic [7]. The dangers related to the consumption of meat contaminated with *Staphylococcus* justify the studies carried out on these bacteria. Studies were carried out on this topic in 2012 in the Cotonou-Porto-Novo slaughterhouses [8], the ones carried out the same year in the butcher's block of Kumbungu and Tolon towns [9] and the last results in the Ouagadougou slaughterhouses [10].

However, no study has explored the domain of beef contamination germs in Cameroon. The aim of this study was to assess the dangers associated with the *Staphylococcus* with positive coagulase (SPC) on raw beef produced and supplied in the northern regions of Cameroon. It is specifically based on the evaluation of the SPC densities of the meat produced and distributed in the northern part of Cameroon and the specification of the resistance profile of the isolated SPC to antibiotics.

Materials and Methods

Ethical approval

Samples were collected under the supervision of the Veterinary Services and microbiological analysis was carried out at the National Veterinary Laboratory.

Sampling

This study was carried out in three phases from March to June 2014. The samples were collected at the slaughterhouses and butcheries in the study areas Manwi, Guider, Pitoa, Garoua, and Maroua towns. Altogether, 125 samples were collected where 71 of them were from the slaughterhouses and 54 from the butcheries and their respective suppliers.

The samples were collected using previously published wet and dry swabbing method [11] on the surface of the carcasses (slaughterhouses) and the quarters (butcheries). A total of 982 swabs were collected in this study divided between 568 swabs from the slaughterhouses and 414 swabs from the butcheries. The surfaces corresponded to neck, thorax, flank, and rumsteck [11,12]. The sampling phases were just collected after a veterinary inspection in some slaughterhouses while in the butcheries, the time slot was 9:30 AM-10:30 AM right after the carving operations. The swabs were kept refrigerated $(0-4^{\circ}C)$ in an ice box containing some ice packs. The box was equipped with a thermometer to monitor the samples shipping temperature. Afterward, the sampling box was transported to the National Veterinary Laboratory in Boklé Garoua.

Microbiological analysis

All the samples were tested 24 h after the sampling phases proper. The discovery and the counting of the SPC were conducted according to the French norms (EN V08 057-1) [13]. To achieve the antibiogram, a total of 102 strains of SPC were used in this study. 56 strains came from slaughterhouses and 46 others came from butcheries. The methods used for the realization of the classical antibiogram and to test the sensibility of the SPC to methicillin were described by the dissemination method over the Mueller-Hinton oversalted agar [14]. In addition, antibiotics such as Penicillin G, Gentamicin, Kanamycin, Oxytetracycline, Trimethoprim-sulfonamide, and Nitrofuran were used for antibiogram experimentation.

Statistical analysis

The XL-Stat software was used for the statistical analysis. The Mann–Whitney U-test helped to compare the frequency of contaminations caused by the SPC between the slaughterhouses and the butcheries. Moreover, the relations existing between the antibiotic resistance profiles and the structures in charge of the sampling origins (slaughterhouses and butcheries) were examined by the χ^2 test (Σ^2).

Results

Meat microbiological contamination

Slaughterhouses

The densities of *Staphylococcus*, respectively, varied from 1.97±0.8 log (CFU/cm²) to 2.6±0.66 log (CFU/cm²) (Table-1). The comparison in the distribution of the loads between the various slaughterhouses disclosed a no significant difference of these densities in almost all the totality of those slaughterhouses; except the Manwi and Guider slaughterhouses which showed a significant difference (p=0.03 < α) in the distribution of their microbial loads (Table-2).

Butcheries

Staphylococcus concentration varied between $2.32\pm1.5 \log (CFU/cm^2)$ for butcheries receiving meat from the Garoua slaughterhouse and $3.82\pm1.01 \log (CFU/cm^2)$ for those which are supplied by those in Ngaoundere (Table-3). The difference in the distribution of the *Staphylococcus* among these butcheries revealed a significant difference of the SPC densities to Ngaoundere town, assessments of the concentration of *Staphylococcus* were rather not significant (Table-4).

Distribution of Staphylococcus contamination

Every bar chart represents the average of the standard deviation; p=0.05.

Antibiotic resistance to the SPC

Slaughterhouses

The evaluation of the resistance to the identified SPC in slaughterhouses showed a variation in resistant profiles. The resistance to Penicillin G was the highest proportions while the resistance to Gentamicin and Furan was the lowest proportions (Table-5). The χ^2 test analysis done revealed that the antibiotic resistance profiles varied according to slaughterhouses (p=0.025; α =0.05; p< α).

Butcheries

Irrespective of the butchery, all SPCs were sensitive to Kanamycin, but most were resistant to Penicillin G (Table-6). The χ^2 test revealed that the antibiotic resistance profiles varied according to the butcheries (p=0.003; α =0.05; p< α).

Discussion

Results showed that the SPC loads were almost closer in all the slaughterhouses studied with the exception of the Manwi municipal slaughterhouse where the load of SPC was much higher than those of Guider and K-djidéo. However, the environmental contamination could explain this point for the Manwi warehouse was not surrounded with used fence and its surface was damaged, badly, or not cleaned [15]. In addition, the water used to clean the carcasses and the equipment

Table-1: Mean	concentration o	f Stanhvlococc	us with nositive	o coaqulase in s	some slaughterhouses.
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Traditional slaughterhouse (log (CFU/cm ²)	MSK n=15	MSG n=15	MSM n=11	MSP n=15	MSGd n=15
Carcass	4.07±1.13	2.56±0.12	3.72±0.79	1.53±0.31	1.62±0.26
Carcass	2.1±0.17	2.27±0.07	2.28±0.13	2.47±0.97	2.87±0.61
Carcass	1.49 ± 0.61	1.84±0.39	1.68 ± 0.43	2.02±1.36	2.57±0.44
Carcass	3.57±0.81	2.37±0.04	3.19±0.45	0.8±2.33	2.04±0.1
Carcass	0.97±0.95	3.42±0.7	2.47±0.01	2.97±0.99	1.52±0.26
Carcass	1.1 ± 0.96	2.68±0.24	2.64±0.13	2.39±1.49	1.62±0.21
Carcass ₇	3.86±0.88	1.84±0.33	2.38±0.03	2.01±1.9	2.66±0.5
Carcass	2.1±0.25	3.49±0.8	3.15±0.51	2.01±2.12	0.97±0.63
Carcass	1.1 ± 0.99	1.57 ± 0.44	1.71±0.33	1.82±2.52	3.1±0.78
Carcass	2.75±0.01	1.81±0.35	3.16±0.52	1.44±3.14	2.51±0.49
Carcass	4.36±1.15	3.16±0.53	1.69 ± 0.00	2.12±3.19	1.77±0.07
Carcass	1.94±0.27	2.41±0.14		0.22±5.17	1.01±0.45
Carcass	2.82±0.26	2.8±0.46		1.97±5.22	2.7±0.6
Carcass	2.53±0.19	1.97±0.11		3.52±5.87	1.22±0.15
Carcass	2±0.00	1.67±0.00		2.32±9.65	1.65±0.00
Means±SD	2.45 ± 1.11	2.39±0.63	2.6±0.66	1.97±0.8	1.99 ± 0.7

MSK=Municipal slaughterhouse of K-djidéo, MSG=Municipal slaughterhouse of Garoua, MSM=Municipal slaughterhouse of Manwi, MSP=Municipal slaughterhouse of Pitoa, MSGd=Municipal slaughterhouse of Guider, SD=Standard deviation

Slaughterhouses	K-djidéo	Garoua	Pitoa-Garoua	Guider
Garoua	0.91			
Pitoa-Garoua	0.30	0.22		
Guider	0.29	0.12	0.98	
Manwi	0.65	0.38	0.05	0.03*

(*p<0.05; ANOVA de Mann-Whitney)

Table-3: Mean of *Staphylococcus* with positive coagulase in the butcheries.

Butcheries log (CFU/cm²)	B[MSK] n=12	B[MSG] n=9	B[MSM] n=12	B[MSP] n=9	B[MSGd] n=12
Carcass,	4.69±1.32	1.4±0.65	3.99±0.28	2.61±0.04	2.35±0.36
Carcass	3.18±0.37	0.57±1.32	4.03±0.34	2.52±0.1	2.67±0.17
Carcass	2.71±0.08	4.29±1.13	5.26±1.24	2.42±0.18	3.04±0.08
Carcass	2.65±0.04	2.49±0.04	3.97±0.47	1.48±0.88	3.9±0.69
Carcass	3.24±0.47	2.34±0.06	2.58±0.45	2.15±0.59	3.05±0.18
Carcass	2.05±0.31	4.05±1.13	5.03±1.21	2.48±0.5	3.18±0.3
Carcass	2.55±0.01	0±1.36	1.98±0.75	2.62±0.57	2.79±0.07
Carcass	3.8±0.87	2.11±0.54	3.94±0.49	3.46±0.26	1.64±0.73
Carcass	1.1±0.82	3.64±0.00	2.19±0.62	4.19±0.00	2.07±0.6
Carcass	2.24±0.29		2.99±0.27		4.66±1.03
Carcass	3.24±0.28		3.64±0.06		1.64±0.59
Carcass	2.46±0.00		3.47±0.00		3.31±0.00
Mean±SD	2.82±0.9	2.32±1.5	3.83±1.01	2.65±0.77	2.86±0.88

B[MSK]=Butchery associated with the Municipal slaughterhouse of K-djidéo, B[MSG]=Butchery associated with the Municipal slaughterhouse of Garoua, B[MSM] =Butchery associated with the Municipal slaughterhouse of Manwi, B[MSP]=Butchery associated with the Municipal slaughterhouse of Pitoa, B[MSGd]=Butchery associated with the Municipal slaughterhouse of Guider, SD=Standard deviation

Table-4: The SPC load comparisons among thebutcheries.

Butcheries	Maroua	Garoua	Pitoa-Garoua	Guider
Garoua	0.39			
Pitoa-Garoua	0.52	0.45		
Guider	0.93	0.39	0.48	
Ngaoundere	0.01*	0.04*	0.01*	0.02*

(*p<0.05, ANOVA de Mann–Whitney), SPC=*Staphylococcus* with positive coagulase

stemmed from a surrounding watercourse and of doubtful sanitary origin. This type of meat contamination by the environment was highlighted in the Cotonou-Porto-Novo slaughterhouse in Benin [16]; the SPC load was lower than the medium threshold value, however, showed a similarity with those reported at the traditional slaughterhouses in Tiaret (Algeria) (2.15 log CFU/cm², 2.93 log CFU/cm²) [17]. This could be justified by the least respect of the hygiene rules because the presence of *Staphylococcus* on the carcasses would be either through physical contact or indirect contact (water air, equipment, staff, etc.) [18,19]. Consequently, fresh meat obtained from SPC would probably be the cause of *Staphylococcus* toxi-infection among the consumers.

The SPC concentrations were higher in the Ngaoundere butcheries than in the other towns, but no significant difference was observed in the other butcheries of the other towns. In the different butcheries in Ngaoundere, the higher concentration of SPC would probably be due to the higher number of people manipulating meat at the level of the trading posts for the human being harbors different types of bacterial flora that could be a source of significant food contaminations [20]. The presence of SPC detected in the various slaughterhouses of the northern regions of Cameroon was of same magnitude like what was obtained in the Kumbungu and Tolon towns in Ghana; hence, justifying the similarity in the unhygienic practices observed in these areas where meat is sold [9,21]. The butchers were no more respecting the basic rules related to the cleaning up of hands before and after manipulating meat (during transport slicing processes), the non-conditioning of meat in packages and the regular disinfecting of the trading posts usage [22,23]. These non-conformities of hygienic rules with regard to the butchery norms lead to the increase in the growth and the proliferation of the SPC responsible for toxi-infection of *Staphylococcus* rate in the consumers.

The no significant difference found in almost all distribution systems compared to production systems (Figure-1). This could be explained by similar practices of manipulation of the raw meats. Moreover, the people who treated raw meats should have similar hygienic practices. Similar results were observed in the region of South Kivu in the Democratic Republic of Congo in 2015 [23].

The relation existing between antibiotic resistance profiles and the slaughterhouses could be due to the source of livestock in each slaughterhouse. Studies carried out by Lozano and Igbinosa revealed that the source of meat contamination by SPC in slaughterhouses would come from microbes on the external hides of slaughtered animals [24,25]. From there, the variety of resistant profiles would probably be the consequence of the germs which were

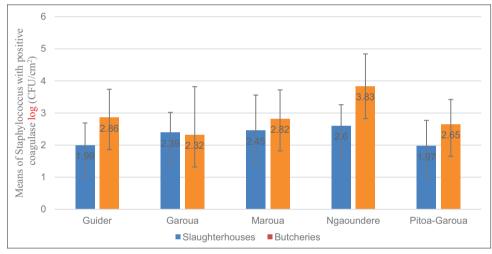


Figure-1: Distribution of the average loads of Staphylococcus with positive coagulase.

Slaughterhouses	K-Djidéo n (%)	Guider n (%)	Pitoa n (%)	Garoua n (%)	Manwi n (%)
Penicillin G	10 (71.42)	12 (66.67)	8 (80)	3 (42.85)	2 (28.57)
Gentamicin	0	0	0	0	0
Oxytetracycline	2 (14.28)	3 (16.67)	1(10)	0	4 (57.14)
Trimethoprim+Sulfonamide	1 (7.14)	1 (5.56)	0	0	0
Nitrofuran	1 (7.14)	0	0	0	1 (14.28)
Kanamycin	0	2 (11.12)	0	0	0
Methicillin	4 (28.57)	1 (5.56)	1 (10)	0	1 (14.28)

Table-6: Resistance to Staphylococcus with positive coagulase in the butcheries.

Butcheries	K-Djidéo n (%)	Guider n (%)	Pitoa n (%)	Garoua n (%)	Manwi n (%)
Penicillin G	7 (63.64)	10 (55.56)	3 (50)	3 (60)	4 (66.67)
Gentamicin	1 (9.1)	0	0	0	0
Oxytetracycline	2 (18.18)	8 (44.44)	6 (100)	2 (40)	2 (33.33)
Trimethoprim+Sulfonamide	1 (9.1)	1 (5.56)	0	0	0
Nitrofuran	Û	Û	2 (33.33)	0	0
Kanamycin	0	0	Ò Ó	0	0
Methicillin	2 (18.18)	2 (11.11)	2 (33.33)	1 (20)	2 (33.33)

subjected to different conditions of the acquisition of their resistance to antibiotics. Penicillin G and Oxytetracycline were the most representative resistances in all studied slaughterhouses. Similar results were found in Ethiopia [26]. As for the Oxacillin, in Ethiopia, the resistance rate was not superior to that one obtained [26]. This could not explain the important use of that molecule. However, the resistance to antibiotics due to *S. aureus* of meat in some slaughterhouses shows the rise of the danger of those bacteria; especially, the resistance of *S. aureus* to the Oxacillin which is also resistant to all the Penicillin group [27].

As in the slaughterhouses, the strong relationship between the SPC resistance profiles and the butchers would implicate the effect of meat contamination by the distribution chain workers. People involved in the selling of meat could in one way or the others contaminate the meat through their SPC, transmitting at the same time germs that are already resistant in man [28]. From there, the SPC-resistant profiles noticed in the butcheries of the greater North of Cameroon would simply reflect a mixing of Staphylococcus having predominantly acquired their resistance in man. On the other hand, the resistance profiles of the SPC samples were higher for the Penicillin G, the Oxytetracycline, and The Oxacillin in the butcheries. Some similar results were also found in the beef and commercialized avians in Ouebec [29] and Lumbumbashi [30]. In the slaughterhouses, the resistance to Staphylococcus might have originated from animals slaughtered, but in the distribution of meat, the importance of the probable transmissions of resisting germs between people and meat proves that the peak of resistance obtained could be the consequence of a frequent usage of antibiotic molecules for the treatment of patients at some hospitals of Cameroon [31].

Conclusion

Higher concentrations of SPC in beef in the slaughterhouses and butcheries of the northern regions of Cameroon were detected on the one hand, and on the other hand, high resistance to frequently used veterinary antibiotics such as Penicillin G, Oxytetracycline, and Oxacillin was also detected. The results highlight that the risk incurred by the beef consumers in the absence of strict hygiene rules and preventive methods in traditional production and distribution of that protein sources from animal origin.

Authors' Contributions

This study was designed, organized, and supervised by RBA, JJNS and RSPN performed sampling and laboratory analysis. RBA and JJNS analyzed and interpreted the data, while the preparation of the manuscript was done by MMMM, JJNS, and RBA. All authors have read and approved the final manuscript.

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Competing Interests

The authors declare that they have no competing interests.

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References

- Ilboudo, A.J., Savadogo, A., Samandoulougou, S., Abre, M., Seydi, M. and Traore, A. (2016) Qualite bactériologique des carcasses de viandes porcines et bovines produites à l'abattoir de Ouagadougou, Burkina Faso. *Rev. Microbiol. Ind. San Environn.*, 10(1): 33-35.
- 2. Fang, Z., Zhao, Y., Warner, R.D. and Johnson, S.K. (2017) Active and intelligent packaging in meat industry. *Trends Food Sci. Technol.*, 61(2): 60-71.
- Denayer, S., Delbrassinne, L., Nia, Y. and Botteldoorn, N. (2017) Food-borne outbreak investigation and molecular typing: High diversity of *Staphylococcus aureus* strains and importance of toxin detection. *Toxins (Basel)*, 9(12): 1-13.
- 4. Loury, P. and Hubert, B. (2015) Les toxi-infections alimentaires collectives dans les pays de la Loire. *Bull. Veill. Sanit.*, (27): 1-16.
- Khallaf, M., Benbakhta, B., Nasri, I., Sarhane, B., Senouci, S. and Ennaji, M.M. (2014) Prévalence du *Staphylococcus aureus* isolé à partir de la viande de poulet commercialisée au niveau de Rabat, Maroc [Prevalence of *Staphylococcus aureus* isolated from chicken meat marketed in Rabat, Morocco]. *Int. J. Innov. Appl. Stud.*, 7(4): 1665-1670.
- 6. De Boer, E., Zwartkruis, J.T.M., Wit, B., Huijsdens, X.W., De Neeling, A.J., Bosch, T., Van Oosteron, R.A.A., Vila, A. and Heuvelink, A.E. (2009) Prevalence of methicillin-resistant *Staphylococcus aureus* in meat. *Int. J. Food Microbiol.*, 134(1-2): 52-56.
- Waters, A.E., Contento-Cuomo, T., Buchhagen, J., Liu, C.M., Watson, L., Pearce, K., Foster, J., Bowers, J., Driebe, E.M., Engelthaler, D.M., Keim, P.S. and Price, L.B. (2011) Multidrug-resistant *Staphylococcus aureus* in US meat and poultry. *Clin. Infect. Dis.*, 52(10): 1227-1230.
- Salifou, C.F.A., Youssao, A.K.I., Salifou, S., Kpodekon, T.M., Tougan, P.U., Ahounou, G.S., Boco, C., Farougou, S., Mensah, G.A. and Clinquart, A. (2012) Evaluation du procédé d' abattage des bovins aux abattoirs de Cotonou-Porto-Novo au sud du Bénin. *Int. J. Biol. Chem. Sci.*, 6(6): 6049-6061.
- 9. Obeng, A.K., Johnson, F.S. and Appenteng, S.O. (2013) Microbial quality of fresh meat from retail outlets in Tolon and Kumbungu districts of the Northern region of Ghana. *Int. J. Sci. Technol.*, 2(6): 423-428.
- Kagambèga, A., Haukka, K., Siitonen, A., Traoré, A.S. and Barro, N. (2011) Prevalence of *Salmonella* Enterica and the hygienic indicator *Escherichia coli* in raw meat at markets in Ouagadougou, Burkina Faso. *J. Food Prot.*, 74(9):

1547-1551.

- Kunová, S., Kačániová, M., Haščík, P., Čuboň, J., Lopašovský, L. and Kluz, M. (2015) Evaluation of the disinfection effectiveness at meat processing by the two methods. J. Microbiol. Biotechnol. Food Sci., 4(6): 564-567.
- 12. Nouichi, M.H. and Taha, S. (2009) Superficial bacterial contamination of ovine and bovine carcasses at El-Harrach Slaughterhouse (Algeria). *Eur. J. Sci. Res.*, 38(3): 474-485.
- Hamiroune, M., Saidani, K., Naceur, R., Belarbi, H.S., Foughalia, A. and Berber, A. (2017) Microbiological quality of merguez in some retailing meat shops in the region of M'Sila (Algeria). *Afr. J. Microbiol. Res.*, 10(6): 211-217.
- CLSI/NCCLS. (2002) Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated from Animals; Approved Standard-Second Edition Serving the World's Medical Science Community through Voluntary Consensus. p22.
- Tanih, N.F., Sekwadi, E., Ndip, R.N. and Bessong, P.O. (2015) Detection of pathogenic *Escherichia coli* and *Staphylococcus aureus* from cattle and pigs slaughtered in abattoirs in Vhembe district, South Africa. *Sci. World J.*, 2015.
- 16. Salifou, C.F.A., Boko, K.C., Attakpa, Y.E., Agossa, R., Ogbankotan, I., Farougou, S., Mensah, G.A., Salifou, S., Clinquart, A. and Youssao, A.K.I. (2013) Evaluation de la qualité bactériologique de viande fraîche de bovins abattus aux abattoirs de Cotonou-Porto-Novo au cours de la chaîne de distribution. J. Anim. Plant Sci., 17(2): 2564-2579.
- Hamid, H.A., Fatma, B., Riad, B., Hebib, A. and Claude, S. (2013) Evaluation de la contamination bactérienne superficielle des carcasses bovines dans un abattoir Algérien. *J. Anim. Plant Sci.*, 19(2): 2901-2907.
- Benaissa, A., El Hadj Khelil, A.O., Adamou, A., Babelhadj, B., Hammoudi, M. and Riad, A. (2014) Qualité de la viande de dromadaire dans les abattoirs de Ouargla en Algérie.II. Contamination bactérienne superficielle des carcasses. *Rev. Elev. Med. Vet. Pays Trop.*, 67(4): 229-233.
- Ahouandjnou, H., Bonou, J., Dougnon, V., Adéoti, Z., Yedji, R. and Toukourou, F. (2015) Evaluation of the microbiological quality of cattle carcasses in some slaughterhouses at Benin, West Africa. *Int. J. Sci. Rep.*, 1(5): 228-234.
- Dibi, E.A.D., N²Goran-Aw, Z.E.B., Akmel, D.C., Kablan, T. and Assidjo, E.N. (2017) Risques microbiens liés à la consommation de la viande bovine braisée Choukouya en Côte d' Ivoire [Microbial hazards linked to the consumption of braised beef meat in Côte d' Ivoire]. *Int. J. Innov. Appl. Stud.*, 19(3): 496-507.
- FAO and IFIF. (2013) Bonnes Pratiques Pour l'industrie de l'alimentation Animale-Mise en oeuvre du Codex Alimentarius. 9th ed. Manuels FAO: Production et Santé

Animales, Rome, Italy.

- Akim, B.L., Gado, D., Innocent, D., Ahmed, A., Seydou, D.O.I. and Ahyi, V. (2017) Etude de la contamination de surface des carcasses de bovins dans la zone d' abattage de Kandi nord du Bénin. J. Appl. Biosci., 1(14): 11388-11392.
- 23. Babe, T., Munyuli, T., Ombeni, J., Kashosi, T. and Mwangi, T. (2018) Hygienic quality assessment of fresh beef meat in Bukavu urban Slaughterhouses, South Kivu Province of the long sale chain : Potential health risks for consumers Eastern D.R. *Bact. Emp.*, 1(1): 1-9.
- 24. Lozano, C., Gharsa, H., Slama, K.B., Zarazaga, M. and Torres, C. (2016) *Staphylococcus aureus* in animals and food : Methicillin resistance, prevalence and population structure. A review in the African continent. *Microorganisms*, 4(1): 1-19.
- 25. Igbinosa, E.O., Beshiru, A., Akporehe, L.U., Oviasogie, F.E. and Igbinosa, O.O. (2016) Prevalence of Methicillin-resistant *Staphylococcus aureus* and other *Staphylococcus* Species in raw meat samples intended for human consumption in Benin City, Nigeria : Implications for public health. *Int. J. Environ. Res. Public Health*, 13(10): 949.
- 26. Adugna, F., Pal, M. and Girmay, G. (2018) Prevalence and antibiogram assessment of *Staphylococcus aureus* in beef at the municipal abattoir and butcher shops in Addis Ababa, Ethiopia. *Biomed. Res. Int.*, 2018: 5017685.
- 27. Lee, J.H. (2003) Methicillin (oxacillin)-resistant *Staphylococcus aureus* strains isolated from major food animals and their potential transmission to humans. *Appl. Environ. Microbiol.*, 69(11): 6489-6494.
- Aubry-Damon, H., Grenet, K., Sall-Ndiaye, P., Che, D., Cordeiro, E., Bougnoux, M.E., Rigaud, E., Le Strat, Y., Lemanissier, V., Armand-Lefevre, L., Delzescaux, D., Desenclos, J.C., Lienard, M. and Andremont, A. (2004) Antimicrobial resistance in commensal flora of pig farmers. *Emerg. Infect. Dis.*, 10(5): 873-879.
- Bernier-Lachance, J. (2015) Prévalence et Caractérisation de *Staphylococcus aureus* Résistant à la Méthicilline d'Origine Aviaire au Québec par l'Université de Montréal. Thesis or Dissertation.
- Victor, O.E., Roger, L.W.L., Gael, N.M. and Celestin, P.S. (2016) Détection des résidus d'antibiotiques dans les denrées alimentaires d'origine bovine et aviaire commercialisées à Lubumbashi (RD Congo). *J. Appl. Biosci.*, 43(102): 9763-9770.
- Akoachere, J.F.T., Yvonne, S., Akum, N.H. and Seraphine, E.N. (2012) Etiologic profile and antimicrobial susceptibility of community-acquired urinary tract infection in two Cameroonian towns. *BMC Res. Notes*, 5(219): 1-8.
