Prevalence and fecal egg load of gastrointestinal parasites of Angora goats in four agro-ecological zones in Lesotho

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doi: www.doi.org/10.14202/vetworld.2021.339-346 **How to cite this article:** Matsepe LG, Molapo S, Phalatsi M, Phororo M (2021) Prevalence and fecal egg load of gastrointestinal parasites of Angora goats in four agro-ecological zones in Lesotho, *Veterinary World*, 14(2): 339-346.

Abstract

Background and Aim: Goats are reared for their meat, mohair and other socio-cultural needs in Lesotho. Helminth infections are some of the major setbacks in the goat production industry due to their negative impact on animals' health, resulting in significant losses on meat and mohair production and death. A cross-sectional study was conducted to determine the prevalence, fecal egg infestation, and morphological identification of gastrointestinal parasites in goats.

Materials and Methods: Fecal samples were collected from 765 goats and subjected to McMaster egg counting techniques using the flotation method. Statistical analyses were performed using the Statistical Package for the Social Sciences (SPSS v.26.0).

Results: The overall prevalence of gastrointestinal parasites was 94.7%, and the identified gastrointestinal parasites were nematodes (64.7%), coccidia (25.8%), and cestodes (4.2%). *Haemonchus contortus* was identified as the prevalent gastrointestinal nematode species found in goats. The prevalence and fecal egg count of gastrointestinal parasites were significantly higher (p<0.05) in goats located in the highlands and Senqu River Valley, while goats in the lowlands demonstrated a significantly (p<0.05) higher prevalence of *H. contortus*. Immature goats and kids were more significantly (p<0.05) prone to gastrointestinal parasites.

Conclusion: The nematodes and coccidia infestations were prevalent in goats located in the highlands and foothills, respectively, whereas nematode and coccidia fecal egg loads were higher in goats located in the foothills and Senqu River Valley, respectively.

Keywords: fecal egg counts, gastrointestinal parasites, goats, Haemonchus contortus, prevalence.

Introduction

Angora goat rearing is a traditional Basotho farmers' activity and plays a key role in their nutrition, social, and economic needs. Angora goat production serves as a cushion in the event of crop failure due to climatic vagaries, especially in arid and semi-arid environments [1,2]. Angora goats provide an essential income source to farmers by selling their mohair and meat or the animals themselves [3-5]. However, goat production faces multiple challenges, such as harsh climatic conditions, poor management and scarcity of fodder, and infectious and noninfectious diseases. In addition, Bath et al. [6] stated that despite Angora goats being renowned for their mohair production, they are susceptible to several diseases. Among the diseases, parasites are more problematic in the developing world, mainly where nutrition and sanitation standards are low. Moreover, Hoste et al. [7] indicated that parasitic nematodes of the gastrointestinal

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tract (GIT) are the main constraints to goat production worldwide and a significant health issue in goat rearing areas with poor sanitation and management. Gastrointestinal parasites remain a primary constraint to ruminant production since they can reduce in skeletal growth, live-weight gain, and milk yield [8].

Furthermore, Ademola *et al.* [9] reported gastrointestinal nematode (GIN) parasitism as the most serious constraint affecting ruminant production. Dappawar *et al.* [1] also stated the browsing habit of goats and their nomadic nature of husbandry as the most factors exposing them to parasitic infestation. Therefore, the GIT of animals harbors a wide variety of parasites, such as helminths, coccidia, and more, which cause clinical and subclinical parasitism. Zanzani *et al.* [10] also reported that GINs are important helminth groups that cause direct damage to livestock.

A cross-sectional study was conducted to determine the prevalence, fecal egg infestation, and morphological identification of gastrointestinal parasites in goats.

Materials and Methods

Ethical approval

The ethical approval was granted by the Department of Animal Science of the National University of Lesotho based on the recommended principles for the use of animals in conducting research. The animals were used with the consent of the farmers.

Study area and period

The fecal sample collection was conducted from December 2018 to October 2019 in four agro-ecological zones of Lesotho which are Sengu River Valley, Highlands (Mountains), Foothills and lowlands represented by Qacha's Nek, Thaba-Tseka, Leribe and Mafeteng districts, respectively (Figure-1). Sengu River Valley is a narrow strip of land lying between an altitude of 1400 m and 1800 m and situated at 30°02'46.03" S and 28°21'40.79" E. The Highlands are the largest ecological region occurring at an elevation of 2000 m to 3400 m with coordinates of 29°31'59.85" S and 28°16'7.04" E. The Foothills are a strip of land lying between 1800 m and 2000 m above sea level with the coordinates of 29°00'3.79" S and 28°12'36.53" E whereas the lowlands are a narrow belt of land where elevation ranges from 1400 m and 1800 m with coordinates of 29°36'2.32" S and 27°17'32.71" E [11,12].

Experimental animals

A total of 765 experimental animals kept under a semi-intensive management system were randomly selected. Animals <1 year (0-12 months) were considered kids, while those >1 year up to 2 years were immature (12-24 months), and those above 2 years were considered adults (>24 months).

Study design and sampling method

The simple random sampling technique was employed in the four agro-ecological zones of Lesotho. Four villages in each agro-ecological zone were selected with the assistance of shearing shed farmer associations. Three goat farmers were randomly selected in each agro-ecological zone, whereby each farmer provided 15 animals as experimental units composed of five kids, five immature, and five adult goats.

Fecal sample collection

Fecal samples were collected directly from each animal's rectum using sterile disposable plastic gloves and were placed in the labeled screw-capped plastic bottles and then kept in a cooler box with ice packs. The samples were then transported to the National University of Lesotho and refrigerated at 4°C, and the laboratory analyses were performed within 48 h.

Fecal sample examination

Two grams of crushed fecal samples were mixed with 58 ml of sodium chloride (flotation solution) and stirred. A homogenous solution was strained into the beaker, and few drops of amyl alcohol (3-5

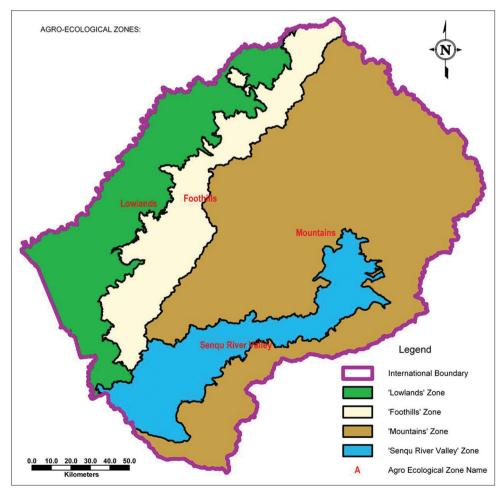


Figure-1: Agro-ecological Zones [Source: Lesotho Ministry of Public Works and Transport].

drops) were added to treat bubbles in the solution. A McMaster quantitative technique was used to determine the number of eggs present per gram of feces, and each number obtained was multiplied by a factor of 100 to give an approximate number of eggs/gram of feces. A sub-sample was drawn from each sample using disposable pipettes and filled both chambers of the McMaster slides, which were observed under a microscope at 100× following standard procedures outlined by several researchers [13-16].

Preparation of fecal cultures and harvesting of infective larvae

Copro-cultures comprising pooled samples from each agro-ecological zone were prepared using goat fecal pellets. The pellets were thoroughly crumbled before being mixed with sufficient vermiculite chips to yield a crumbly mixture, which was lightly compacted and moistened sufficiently. The crumbly mixture was ensured not to be water-logged and then incubated in jars in the dark (laboratory cupboards) at room temperature for 7 days. The inside of jars was sprayed lightly with water before being placed in bright light that stimulates the L₃ to migrate up the inner walls of culture jars and then harvested repeatedly by holding the jars at a slant position with mouth pointing downwards and spraving the inner walls with water to allow the larvae to drain into a suitable container (beaker) as recommended by van Wyk and Mayhew [17].

Larvae preservation

The larvae were preserved unchanged by adding formalin to the larval suspension to a final concentration of about 1-2% and then heated to 57°C in a water bath for about 1 min to kill larvae and straighten or uncurl it [17,18].

Larvae preparation for identification

Larvae were prepared for examination by adding a drop of diluted Lugol's iodine solution to a drop of larval suspension on a glass microscope slide and then mounted a coverslip as outlined by van Wyk *et al.* [18].

Larvae identification

Morphological identification of L_3 of most parasitic nematodes was based principally on examining the caudal (tail) and cranial (head) extremities. Conventional characteristics for identification (total length, sheath tail extension [STE] length, and filament length) of infective larvae GIN species were microscopically examined at $10 \times$ with ocular number ten and measured using a calibrated stage micrometer that was imputed in an ocular lens [17].

Statistical analysis

The data collected were manually inputted in Microsoft Excel spreadsheet and transferred into SPSS v.26.0 for analyses. General linear model was employed to determine the effect of agro-ecological zone and age on the prevalence and fecal egg load of gastrointestinal parasites in goats. Generalized estimating equation (GEE) was used to analyze the FEC, where negative binomial regression was involved in the analysis. Odds ratio was used to measure the association between exposure and outcome. Descriptive statistics were implemented to determine the prevalence of identified species between the animals' agro-ecological zones and age groups. Pearson Chi-square test was also adopted to assess the degree of association between each risk factor and GIN. In the analyses, the confidence level was held at 95%.

Results

The goats in the highlands and foothills were more susceptible to nematode infestation than those in the lowlands and Senqu River Valley (Table-1). The prevalence of nematodes in goats in the lowlands was significantly (p<0.05) lower, and the chances of nematode infestation from the lowlands to the foothills, highlands, and Senqu River Valley increased significantly (p<0.05).

As illustrated in Table-1, the prevalence of coccidia infestation showed that the infestation was significantly (p<0.05) higher in the foothills. The results also demonstrated that the likelihood of increased coccidia infestation from the lowlands to the foothills, highlands, and Senqu River Valley was significant (p<0.05).

Goats in the lowlands were significantly (p<0.05) different from those in the highlands in the prevalence of cestodes. As depicted in Table-1, these results also displayed that the odds of having cestodes infestation from lowlands to foothills and Senqu River Valley increased significantly (p<0.05). However, the like-lihood of having a cestode infestation from the low-lands to highlands increased insignificantly (p>0.05).

The prevalence of nematodes was high (p<0.05) in immature goats, followed by adults, and the lowest prevalence was observed in kids (Table-2). The odds of having a nematode infestation from adult goats to immature goats increased insignificantly (p>0.05), while from adults to kids, odds decreased significantly (p<0.05).

In the case of coccidian (Table-2), the infestation was found significantly (p<0.05) lower in adult goats than in immature and young goats. The likelihood of coccidia infestation increased significantly (p<0.05) from adult goats to immature goats. Furthermore, from adults to kids, the probability of coccidia infestation also increased significantly (p<0.05).

In terms of cestodes, a high prevalence was observed in kids, followed by immature goats, and was lowest in adult goats. There was also a significant difference (p<0.05) discovered between the different age groups. The likelihood of goats exhibiting cestodes infestation from adults to immature and kids increased significantly (p<0.05). The low prevalence of cestode infestation observed in adults might be due to body resistance as they might have developed immunity due to repeated natural infestations.

Haemonchus contortus was the only species of GI nematodes that were discovered in this study. Of 172 samples examined, 130 were positive for *H. contortus*, and the overall prevalence in goats in this study was 75.6%. The cranial extremity of the infective larvae was observed to have a bullet-shaped head (Figure-2). According to van Wyk and Mayhew [17], the length of STE is an essential criterion for identification; therefore, the caudal extremity, which comprises a sheath tail filament, ranged following the report of van Wyk *et al.* [18], which was 2.2-2.5 µm and 10-15% for STE and filament, respectively. van Wyk *et al.* [18] van Wyk *et al.* [17] further emphasized

that *H. contortus* had an STE of $>2.0 \,\mu\text{m}$, with 10-15% filament, head bullet-shape, and temper fast to a point (Figure-3).

The higher (p<0.05) prevalence of *H. contortus* observed in the lowlands (Table-3) compared with other agro-ecological zones suggests that the environmental conditions were favorable for the development and survivability of infective larvae (*H. contortus*) and lack of shrubs for goats to browse; hence, forcing the goats to graze closer to the ground, thereby, contacting the infective larvae. A higher (p<0.05) *H. contortus* infestation was observed in kids than in adult and immature goats.

Agro-ecological zones	No. examined	Prevalence (%)	SE	Exp. (B)	Sig. Level
Nematodes					
Senqu river valley	1 251	64.7ª	0.014	0.771	0.000
Highlands	1 533	70.0 ^b	0.012	0.607	0.000
Foothills	1 724	67.1 ^{ab}	0.011	0.696	0.000
Lowlands	1 986	58.6°	0.011	1	
Coccidia					
Senqu river valley	1 251	26.3ª	0.012	0.650	0.000
Highlands	1 533	25.4ª	0.011	0.680	0.000
Foothills	1 724	33.6 ^b	0.011	0.458	0.000
Lowlands	1 986	18.8°	0.009	1	
Cestodes					
Senqu river valley	1 251	5.3ª	0.006	0.588	0.003
Highlands	1 533	4.2 ^{ab}	0.005	0.752	0.115
Foothills	1 724	4.6ª	0.005	0.682	0.028
Lowlands	1 986	3.2 ^b	0.004	1	

Table-1: Prevalence of gastrointestinal parasites by agro-ecological zones.

^{abc}Means in the same column with different superscripts differ significantly (p<0.05). Exp. (B)=Exponential Beta, No.=Number, Sig.=Significant level

Table-2: Prevalen	ce of gastrointestina	al parasites by age.
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Age	Samples examined	Prevalence (%)	SE	Exp. (B)	Significance level
Nematodes					
Kids	2 182	58.2ª	0.01	1.47	0.00
Immature	2 101	69.1 ^b	0.01	0.91	0.16
Adults	2 211	67.1 ^b	0.01	1	
Coccidia					
Kids	2 182	33.0ª	0.01	0.51	0.00
Immature	2 101	24.2 ^b	0.01	0.79	0.01
Adults	2 211	20.1°	0.01	1	
Cestodes					
Kids	2 182	6.6ª	0.005	0.332	0.00
Immature	2 101	3.6 ^b	0.004	0.629	0.01
Adults	2 211	2.3°	0.003	1	

 abc Means in the same column with different superscripts differ significantly (p<0.05). Exp. (B)=Exponential beta

Table-3: Prevalence of Haemonchus contortus in goats by different agro-ecological zones and age.

Category	No. examined	Positive cases	Prevalence (%)	χ^2 – value	p-value
Agro-ecological zones					
Senqu River Valley	41	34	82.9	31.745	0.000
Highlands	40	26	65.0		
Foothills	40	20	50.0		
Lowlands	51	50	98.0		
Age of goats					
Kids	64	55	85.9	5.939	0.051
Immature	58	40	69.0		
Adults	50	35	70.0		
			70.0		

 χ^2 – Pearson Chi-square value, P – Significance level

A higher (p<0.05) intensity of nematode FEC mean was observed in goats located in the foothills compared with other agro-ecological zones (Table-4). The likelihood of coccidia infestation increased significantly (p<0.05) from the lowlands to the foothills and from the lowlands to Senqu River Valley but decreased significantly (p<0.05) from the lowlands

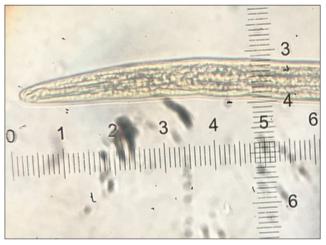


Figure-2: Head shape.

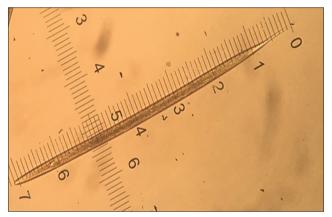


Figure-3: Typical Haemonchus spp.

Table-4: Means of gastrointestinal FEC in different agroecological zones.

Agro-ecology	EMM	SE	Exp (B)	Exp (B) in %
Nematodes				
Senqu river valley	440.13 ^{ab}	36.18	1.419	41.9
Highlands	453.36ª	21.99	1.461	46.1
Foothills	542.52ª	85.92	1.749	74.9
Lowlands	310.22 ^b	22.74	1	1
Coccidia				
Senqu river valley	927.18ª	463.47	5.756	475.6
Highlands	133.68 ^b	17.83	0.830	17.0
Foothills	329.41°	57.35	2.045	104.5
Lowlands	161.08 ^d	33.52	1	1
Cestodes				
Sengu river valley	33.81ª	10.27	0.813	18.7
Highlands	120.03 ^b	57.95	2.886	188.6
Foothills	56.32ª	27.81	1.354	35.4
Lowlands	41.59ª	13.70	1	1

^{abc}Means in the same column with different superscripts differ significantly (p<0.05). EMM=Estimated marginal means, SE=Standard error, Exp.(B)=Exponential Beta to the highlands. Our findings also displayed that the cestodes egg infestation from the lowlands to the foothills increased significantly (p<0.05) while from the lowlands to Senqu River Valley significantly (p<0.05) decreased.

The nematode egg load was higher in immature goats, followed by adult goats, and lowest in kids (Table-5). The adopted GEE revealed that nematode FEC increased significantly (p<0.05) from adults to immature goats, whereas from adults to kids, the like-lihood of infestation decreased significantly (p<0.05).

This study showed that adult goats were significantly lower (p<0.05) than other age groups in terms of coccidia fecal egg load. The study further demonstrated that coccidia egg count was significantly (p<0.05) higher in kids, followed by immature goats, and lowest in adult goats.

The kids had significantly (p<0.05) higher cestode egg load than immature and adult goats. The likelihood of infestation increased significantly (p<0.05) from adult to immature goats, and from adults to kids, the chances of infestation increased significantly (p<0.05).

Discussion

This study's results agree with the findings obtained by Koinari et al. [19], who reported a high prevalence of Strongvloides (nematodes) in goats in the highlands. This high prevalence of nematodes, coccidian, and cestodes observed in goats in the highlands, foothills, and Senqu River Valley, respectively, suggests the existence of favorable environmental conditions for survival and development of parasitic larvae to the infective stage. Nabi et al. [4] illustrated that the prevalence of GIT parasites varies in diverse geographical conditions and is influenced by climate, management, vegetation, and livestock density. The low GIP prevalence in goats reared in the lowlands agrees with the observation of Mpofu et al. [2], who illustrated that the low prevalence of nematodes, coccidia, and cestodes in arid zones (lowlands) might be attributed to the fact that these areas are extremely hot

Table-5: Means of gastrointestinal FEC in different agegroups of goats.

Age	ЕММ	SE	Exp. (B)	Exp. (B) in %
Nematodes				
Kids	323.19ª	16.01	0.712	28.8
Immature	517.61 ^b	67.82	1.140	14.0
Adults	454.23°	34.00	1	
Coccidia				
Kids	723.69ª	262.41	8.944	794.4
Immature	235.36 ^b	70.45	2.909	190.9
Adults	80.91 ^b	16.41	1	
Cestodes				
Kids	165.95ª	47.28	34.944	3 394.4
Immature	15.90 ^b	9.71	3.347	234.7
Adults	4.75°	1.38	1	

^{abc}Means in the same column with different superscripts differ significantly (p<0.05). EMM=Estimated marginal means, SE=Standard error, Exp.(B)=Exponential bet and receive scarce, erratic rainfall, which is unfavorable for GIP development, survival, and transmission.

However, in a study by Moiloa et al. [3], nematode infestation was higher in the lowlands of Maseru district and the foothills of Quthing district, while coccidia infestation was highest in the mountains of both Maseru and Quthing districts. Berhe and Aragaw [20] further emphasized that the higher parasitic infestation suggests that goats, which are typically browsers, may have been forced to graze, perhaps due to lack of adequate browsing material in the study area. Emiru et al. [8] emphasized that the high prevalence could be ascribed to overstocking, poor nutrition (starvation), poor management practice of the animals (lack of sanitation), and frequent exposure to the communal grazing lands contaminated, which agrees with the findings of this study since pastures are communally grazed. According to a study conducted by Dagnachew et al. [21], there was a statistically significant (p<0.05) difference in the species prevalence of helminthiasis, with the highest being in Chillga (lowland) for strongyle infestation (55.45%) and the lowest was in Dabat (highland) (22.52%). In addition, Khodakaram-Tafti and Hashemnia [22] emphasized that the variation in the prevalence and distribution of coccidiosis may be attributed to, among other factors, agroecology. Moreover, Rahman et al. [23] also outlined that the lower prevalence of Moniezia spp. (cestodes) might be due to less dissemination of eggs in the feces from the gravid segment.

The results of this study agree with those reported by Sunandhadevi et al. [16], who showed that the prevalence of GI nematode was more in immature (84.44%) than in adult (64.22%) and young goats (64.10%). Islam et al. [24] Mpofu et al. [2] also added that infestation with GIPs was more prevalent in immature goats than in adult goats and kids. This high prevalence of GI nematodes obtained in immature goats can be attributed to the fact that goats were kept in a contaminated environment, overstocked, and had low immunity. However, despite the above inference drawn, Dagnachew et al., [21] Nabi et al., [4] noted the significant difference between young and adult goats, in which young goats showed higher nematode infestation than adults. Mpofu et al. [2] also argued that young goats showed a higher incidence of nematode infestation than adult goats. Nabi et al. [4] emphasized that with an increase in age, the prevalence of GIT nematodes decreased due to the development of immunity, and it has been reported that small ruminants develop partial immunity against GIT nematodes. Furthermore, Lone et al. [25] stated that the most infested age group was kids (97.77%). Alternatively, Ardo and Bitrus [26] argued that adult animals were more infested than the young goats, whereas Yusof and Isa [27] viewed that the highest prevalence of GINs infestation was observed in immature (87.8%), followed by the adults (86.4%) and kids (53.1%).

According to the present findings, Gupta et al. [28] reported that the overall prevalence of gastrointestinal parasites was 98.05% of which the highest percentage was *Eimeria* spp. (coccidia) (86.34%). This may be attributed to the fact that kids are underdeveloped and have lower immunity resistance toward coccidia infestation than immature and adult goats. Similarly, Jittapalapong et al. [29] also reported that a higher GI parasitic prevalence was observed among goats aged <1 year (94.9%) than goats aged 1-2 years (75.4%) or >2 years old (51.4%). Zvinorova et al. [30] added that Eimeria (coccidia) infestation was the most prevalent parasitic infestation, followed by Strogyles (nematodes). Nevertheless, in a study conducted by Yusof and Isa [27], adult goats' well-developed resistance appeared to be relative rather than absolute since adults continued to harbor *Eimeria* spp. oocysts in feces, which contributed as a source of infestation for younger goats. Moreover, Mpofu et al. [2] added that adult animals may acquire immunity to the parasite through frequent challenges and may expel the ingested parasite before an infestation is established, thereby contaminating the environment.

These results also agree with the study conducted by Yusof and Isa [27], who noted that the prevalence of tapeworm (cestodes) was higher in young and juvenile age groups than adult animals. Furthermore, Raza [31] highlighted that the host animal's age was an essential factor influencing the prevalence of GI parasites.

The results of this study agree with the findings of a previous study in Papua New Guinea [19], who discovered that the mean EPG counts for goats infested with *Strongyloides* (nematodes) in the highlands (Tambul) were significantly higher than those in the lowlands (Labu). Koinari *et al.* [19] also added that the mean EPG for *Eimeria* was higher in the highlands than in the lowlands. However, Regassa *et al.* [32] argued that higher values were recorded for the lowland areas, followed by mid-altitude areas with the lowest values in the highland areas.

Sharma *et al.* [33] also observed the highest (1 690.1 EPG) number of oocysts in the immature age group, whereas a minimum (22.73 EPG) oocysts load was found in kids that were still suckling. This high nematode infestation of the immature age group than in kids and adult goats might be influenced by the weaker immunological response of immature goats, thereby becoming susceptible to GINs.

Following the results of this study, Kahan and Greiner [34] also added that the EPG of kids was significantly higher than in adult goats (p<0.01). These findings illustrated that the number of oocysts shed decreased as the animal matured. Kahan and Greiner [34] further added that this decline is most likely due to the development of an immunological resistance related to host's age and exposure to the parasites. In contrast with the current study, Kheirandish *et al.* [35] reported that adult goats had higher oocyst counts than kids and immature goats.

The findings in this study agree with the report of Yusof and Isa [27], who outlined that cestode infestation was prevalent in kids, and because of the low pathogenicity in adults, tapeworm (cestodes) infestation was considered minor.

Conclusion

It can be concluded that nematode and coccidia infestation was higher in the highlands and foothills, while nematodes and coccidia fecal egg loads were higher in foothills and Senqu River Valley. *H. contortus* was identified as the prevalent GIN species found in goats. Nematodes and coccidia highly infested immature and kids.

Authors' Contributions

LGM conceived and designed the research under the guidance of SM. LGM, MoP, and MaP conducted the sample collection. MoP and LGM carried out the morphological identification analysis. SM, MaP, and LGM carried out the data analyses. LGM wrote the manuscript. SM reviewed the manuscript. All authors read and approved the final manuscript.

Acknowledgments

Wool and Mohair Promotion Project, Lesotho, is acknowledged for funding this study under grant no. 2000000817 and project no. 2000000053. Farmers are also appreciated for providing the animals used in this study.

Competing Interests

The authors declare that they have no competing interests.

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References

- 1. Dappawar, M.K., Khillare, B.S., Narladkar, B.W. and Bhangale, G.N. (2018) Prevalence of gastrointestinal parasites in small ruminants in Udgir area of Marathwada. *J. Entomol. Zool. Stud.*, 6(4): 672-676.
- 2. Mpofu, T., Nephawe, K. and Mtileni, B. (2020) Prevalence of gastrointestinal parasites in communal goats from different agro-ecological zones of South Africa. *Vet. World*, 13(1): 26-32.
- Moiloa, M.J. (2017) Gastrointestinal parasites of Angora goats in Lesotho: Prevalence and control methods. *Afr. J. Rural Dev.*, 4(4): 471-478.
- Nabi, H., Saeed, K., Shah, S., Rashid, M., Akbar, H. and Shehzad, W. (2014) Epidemiological study of gastrointestinal nematodes of goats in district Swat, Khyber Pakhtunkhwa, Pakistan. *Sci. Int.*, 26(1): 283-286.
- Ng'ambi, J.W., Norris, D., Matebesi, P.A. and Khitsane, L. (2006) Productivity of Angora goats in the communal rangelands of Molimo-Nthuse area in Lesotho. *Trop. Anim. Health Prod.*, 38(7-8): 597-603.
- 6. Bath, G.F., van Wyk, J.A. and Pettey, K.P. (2005) Control measures for some important and unusual goat diseases in Southern Africa. *Small Rumin. Res.*, 60(1): 127-140.
- 7. Hoste, H., Torres-Acosta, J.F., Paolini, V. and

Aguilar-Caballero, A. (2005) Interactions between nutrition and gastrointestinal infections with parasitic nematodes in goats. *Small Rumin. Res.*, 60(1-2): 141-151.

- Emiru, B., Amede, Y., Tigre, W., Feyera, T. and Deressa B. (2013) Epidemiology of gastrointestinal parasites of small ruminants in Gechi district, Southwest Ethiopia. *Adv. Biol. Res.*, 7(5): 169-174.
- 9. Ademola, I., Fagbemi, B.O. and Idowu, S.O. (2007) Anthelmintic efficacy of *Nauclea latifolia* extract against gastrointestinal nematodes of sheep: *In vitro* and *in vivo* studies. *Afr. J. Tradit. Complement. Altern. Med.*, 4(2): 148-156.
- Zanzani, S.A., Gazzonis, A.L., Cerbo, A.D., Varady, M. and Manfredi, M.T. (2014) Gastrointestinal nematodes of dairy goats, anthelmintic resistance and practices of parasite control in Northern Italy. *BMC Vet. Res.*, 10(1): 1-10.
- 11. Mafongoya, P.L. and Ajayi, O.C. (2017) Indigenous Knowledge Systems and Climate Change Management in Africa. CTA, Wageningen, Netherlands, p316.
- 12. Moeletsi, M. and Walker, S. (2013) Agroclimatological suitability mapping for dryland maize production in Lesotho. *Theor. Appl. Climatol.*, 114(1-2): 227-236.
- 13. Admasu, P. and Nurlign, L. (2015) prevalence of gastrointestinal parasites of small ruminants in Kuarit district, North West Ethiopia. *Afr. J. Basic Appl. Sci.*, 6(5): 125-130.
- Dilgasa, L., Asrade, B. and Kasaye, S. (2015) Prevalence of gastrointestinal nematodes of small ruminants in and around Arsi Negele town, Ethiopia. *Am. Eurasian J. Sci. Res.*, 10(3): 121-125.
- 15. Glennon, H. (2013) Modified McMaster's Fecal Egg Counting Technique, p7-9.
- Sunandhadevi, S., Rao, M.L.V., Pooja, D., Das, G., Shukla, P. and Dixit, A.K. (2017) Prevalence of gastrointestinal nematodes and risk factors in goats in Jabalpur. *Environ. Ecol.*, 35(2A): 920-922.
- 17. van Wyk, J.A. and Mayhew, E. (2013) Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: A practical lab guide. *Onderstepoort J. Vet. Res.*, 80(1): 1-14.
- van Wyk, J.A., Cabaret, J. and Michael, L.M. (2004) Morphological identification of nematode larvae of small ruminants and cattle simplified. *Vet. Parasitol.*, 119(4): 277-306.
- Koinari, M., Karl, S., Ryan, U. and Lymbery, A.J. (2013) Infection levels of gastrointestinal parasites in sheep and goats in Papua New Guinea. *J. Helminthol.*, 87(4): 409-415.
- 20. Berhe, K. and Aragaw, K. (2019) Abomasal nematode parasites in goats slaughtered in Mekelle town, Northern Ethiopia. *Ethiop. Vet. J.*, 23(2): 90-100.
- 21. Dagnachew, S., Amamute, A. and Temesgen, W. (2011) Epidemiology of gastrointestinal helminthiasis of small ruminants in selected sites of North Gondar zone, Northwest Ethiopia. *Ethiop. Vet. J.*, 15(2): 57-68.
- 22. Khodakaram-Tafti, A. and Hashemnia, M. (2017) An overview of intestinal coccidiosis in sheep and goats. *Rev. Med. Vet.*, 168(1-3): 9-20.
- Rahman, M., Labony, S., Dey, A. and Zahangir, A.M. (2017) An epidemiological investigation of gastrointestinal parasites of small ruminants in Tangail, Bangladesh. *J. Bangladesh Agric. Univ.*, 15(2): 255.
- Islam, S., Hossain, S., Dey, A.R., Alim, A., Akter, S. and Alam, Z. (2017) Epidemiology of gastrointestinal parasites of small ruminants in Mymensingh, Bangladesh. J. Adv. Vet. Anim. Res., 4(4): 356-362.
- 25. Lone, B.A., Chishti, M.Z., Ahmad, F. and Tak, H. (2012) A survey of gastrointestinal helminth parasites of slaughtered sheep and goats in Ganderbal, Kashmir. *Glob. Vet.*, 8(4): 338-341.
- Ardo, M.B. and Bitrus, I. (2015) Prevalence of parasitic gastrointestinal nematodes of small ruminants at Jalingo abattoir, Taraba State, Nigeria. *Bayero J. Pure Appl. Sci.*, 8(2): 29-33.

- Yusof, A. and Isa, M.L. (2016) Prevalence of gastrointestinal nematodiasis and coccidiosis in goats from three selected farms in Terengganu, Malaysia. *Asian Pac. J. Trop. Biomed.*, 6(9): 735-739.
- Gupta, M.K., Rao, M.L.V., Dixit P., Shukla P.C. and Dixit, A.K. (2016) Prevalence of gastrointestinal nematodes in goats in and around Jabalpur. *Environ. Ecol.*, 34(4D): 2495-2497.
- Jittapalapong, S., Saengow, S., Pinyopanuwat, N., Chimnoi, W., Khachaeram, W. and Stich, R.W. (2012) Gastrointestinal helminthic and protozoal infections of goats in Satun, Thailand. J. Trop. Med. Parasitol., 35(2): 48-54.
- Zvinorova, P.I., Halimani, T.E., Muchadeyi, F.C., Matika, O., Riggio, V. and Dzama K. (2016) Prevalence and risk factors of gastrointestinal parasitic infections in goats in low-input low-output farming systems in Zimbabwe. *Small Rumin. Res.*, 143(9): 75-83.
- 31. Raza, M.A. (2013) Prevalence of Intestinal Parasites in

Small Ruminants and their Sensitivity to Treatments with Ethnobotanical Remedies in Cholistan, Pakistan. University of Kassel, Germany.

- 32. Regassa, F., Sori, T., Dhuguma, R. and Kiros, Y. (2006) Epidemiology of gastrointestinal parasites of ruminants in Western Oromia, Ethiopia. *Int. J. Appl. Res. Vet. Med.*, 4(1): 51-57.
- Sharma, A.N., Mandal, A., Nigam, P. and Bhushan, S. (2010) Coccidia and gastrointestinal nematode infections in semi-intensively managed Jakhrana goats of semi- arid region of India. *Trop. Subtrop. Agroecosyst.*, 12(1): 139-143.
- Kahan, T.B. and Greiner, E.C. (2013) Coccidiosis of goats in Florida, USA. *Open J. Vet. Med.*, 3(3): 209-212.
- Kheirandish, R., Nourollahi-Fard, S.R. and Yadegari, Z. (2014) Prevalence and pathology of coccidiosis in goats in Southeastern Iran. J. Parasit. Dis., 38(1): 27-31.
