# A review: Virulence factors of *Klebsiella pneumonia* as emerging infection on the food chain

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#### Abstract

Health problems can be caused by consuming foods that have been processed in unsanitary conditions; hence, the study of the impact of contamination on food and its prevention has become critical. The disease caused by *Klebsiella pneumoniae* in food is increasing significantly every year across the world. The main factors that are essential for the virulence of *K. pneumoniae* are lipopolysaccharide and polysaccharide capsules. Furthermore, *K. pneumoniae* is capable of forming biofilms. Capsule polysaccharides, fimbriae types 1 and 3, are crucial virulence factors contributing to biofilm formation in *K. pneumoniae*. The food contamination by *K. pneumoniae* may not directly pose a public health risk; however, the presence of *K. pneumoniae* refers to unhygienic practices in food handling. This article aims to demonstrate that *K. pneumoniae* should be considered as a potential pathogen that spreads through the food chain and that necessary precautions should be taken in the future.

Keywords: biofilm, food chain, Klebsiella pneumonia, public health, virulence.

#### Introduction

Klebsiella pneumoniae predominantly has caused infectious diseases in immunosuppressed individuals. However, its emergence and spread are spreading even to healthy and immunocompromised people. Furthermore, the K. pneumoniae strain is becoming increasingly antibiotic-resistant, making treatment of infection with the strain extremely difficult [1]. The pathogenicity of K. pneumoniae bacteria is associated with several virulence factors that allow it to evade the host's innate immune mechanisms. Klebsiella pneumoniae virulence factors include capsules, exopolysaccharides associated with mucoviscosity, lipopolysaccharides (LPSs), adhesins, and iron uptake systems. The factors that aggravate the infection caused by K. pneumoniae are multiple antibiotic resistance and its ability to cause nosocomial infections in humans. Furthermore, the previous case involved six Asian patients admitted to a US hospital with K. pneu*moniae* liver abscess; the gastrointestinal tract was suspected as the route of entry in one of the cases [2]. Moreover, K. pneumoniae causes infectious diseases

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such as pneumonia, meningitis, and blood and urinary tract infections [3]. Klebsiella pneumoniae is recognized by most physicians as the cause of community-acquired bacterial pneumonia. The opportunistic pathogen is the primary reason for the hospitalization of immunocompromised patients and individuals with severe diseases. Klebsiella pneumoniae is the main cause of nosocomial Klebsiella infection, a necrotic process that tends to attack the weak. In addition, K. pneumoniae can cause localized diseases and other related infections such as liver abscess, endophthalmitis, and meningitis in healthy individuals [4]. Apart from nosocomial infections, K. pneumoniae also spreads through contaminated food materials and is often considered an agent of foodborne illness. The pathogen can be found in seafood, frozen foods, and fresh meat [5]. In recent decades, foodborne outbreaks have highlighted the importance of developing and implementing preventive measures and programs to ensure food safety [6].

The observations of multidrug resistance (MDR) of *K. pneumoniae* from retail foods, and the presence of isolates associated with highly exposed sources of antibiotics, also make the possibility of antibiotic treatment difficult if the organism causes infection. The authors are concerned that this might be the case as our results are consistent with reports of antibiotic-ic-resistant *K. pneumoniae* detected in various poultry and meat products [5].

Food hygiene and safety are based on various food safety issues, such as the presence of potential

pathogens in food, toxins, resistance to antibiotics or sanitizers, and other virulence characteristics. The many benefits for consumers are in the globalization of trade because it produces a wider variety of high-quality foods that are easily available, affordable, safe, and meet the consumer's needs. However, poor local infrastructure, characteristics of product sales, and lack of supervision in terms of sanitation are factors in the trade that can raise concerns about the potential for food poisoning due to microbiological contamination [7]. Consequently, it is prudent to exercise awareness of common and unexpected contaminants in food, such as K. pneumoniae and other antibiotic-resistant bacteria. In recent years, specifically in non-Asian cohorts, gastrointestinal transport has been recognized as a risk factor for K. pneumoniae colonization. Furthermore, in Australia, a recent study on the probability of transmission of K. Pneumonia revealed that 48% (13/27) of patients in the intensive care unit exhibited intestinal colonization before infection [8]. This review aimed to contemplate the potential risks of the pathogen in retail food hygiene, food safety, and public health and to investigate the virulence factors of K. pneumoniae as an emerging infection on the food chain.

## *Klebsiella pneumoniae* Virulence Factors and Pathogenicity

In addition to the clinical terrain, K. pneumoniae is present in foods, including raw vegetables, pulverized child formula, meat, fish, and road food [9]. In recent years, several foodborne outbreaks caused by K. pneumoniae have been reported in various countries. Klebsiella pneumoniae can express a variety of acridity factors, including capsule, endotoxin, siderophore, iron scavenging system, and adhesins, which play a crucial part in its pathogenesis. The capsule is a significant acridity factor, involved in at least two pathogenic mechanisms, that is, the protection of bacteria from phagocytosis and direct inhibition of the vulnerable host response. Several capsule types (K), especially, K1, K2, K54, K57, K20, and K5, are constantly associated with the pattern of community-acquired invasive pyogenic liver abscess, septicemia, and pneumonia. Moreover, K1, K2, K20, K54, and K57 are predominantly detrimental to experimental infections in mice and are frequently associated with severe infections in humans [10].

The capsules correspond to polysaccharides called K antigens, which are classified into 78 serological types. The capsule synthesis in *K. pneumoniae* is encoded by a gene located on the chromosomal operon, capsule polysaccharides (CPS). The CPS gene cluster hosts several genes i.e., *wzi*, *wza*, *wzb*, *wzc*, *gnd*, *wca*, *cpsB*, *cpsG*, and *galF*, that enable the formation of the capsule [11]. Furthermore, pathogenic bacteria require iron for their replication. Siderophores (iron carriers) are composites buried by microorganisms (similar to bacteria and fungi) to transport iron in the cell membranes. They have an advanced iron magnet than the host transport protein (transferrin). *Klebsiella pneumoniae* produce siderophores to gain iron from host iron-chelating proteins or the terrain for survival and reduplication during mammalian infection. The product of more than one siderophore by *K. pneumoniae* is a means to optimize the successful colonization of different napkins and/or avoid the neutralization of one siderophore by the host. Enterobactin, yersiniabactin, salmochelin, and aerobactin are different types of siderophores expressed by *K. pneumoniae*. The defining factors for high virulence and toxicity include capsules, siderophores, LPSs, and pili [12].

Klebsiella pneumoniae infection is caused by extended-spectrum beta-lactamase (ESBL)-producing bacteria and is resistant to carbapenems. ESBL can be caused by several beta-lactamases, encoded by genes such as TEM, SHV, CTX-M, and OXA [13]. The ESBLs have a greater impact on enhanced morbidity and mortality rates than bacteria that are not resistant to the infection. Nevertheless, multidrug-resistant or not, the capsule-deficient K. pneumoniae strains rarely result in complaints or complications in healthy individuals (except for urinary tract infections). In general, strains of K. pneumoniae or classic strains cause serious infections such as pneumonia, bacteremia, or even meningitis, when infecting individuals with compromised vulnerable systems, including diabetics [14].

The acridity of the K. pneumoniae strain was unaffected by transportation or the expression of medicine resistance, but it made treatment more difficult. Furthermore, due to its ability to infect both healthy and weakened vulnerable systems, a comparison of the K. pneumoniae hypermucoviscous (HV) strain and the classic K. pneumoniae (cKP) strain revealed that enterobactin expression is nearly ubiquitous among both strains and is thus considered to be the primary iron uptake system employed by K. pneumoniae [15]. Furthermore, the irp gene encodes the proteins required for yersiniabactin conflation, the ybt and *fvu* genes encode siderophores, and the *vbtO* gene encodes their uptake receptors. However, K. pneumo*niae* has yet to fully characterize these. The strain is known as hypervirulent K. pneumoniae (hvKP), and it is more toxic, pathogenic, and causes a different cardiovascular disease than cKP [16]. Furthermore, in the absence of lipocalin-2, enterobactin promotes lung colonization and dispersion. In the presence of lipocalin-2; however, the strain of K. pneumoniae that produced only this siderophore was ruled out. Salmochelin is also the c-glucosylated form of enterobacterin. Iron mediates transport in iron-laden forms, and this modification prevents the list of salmochelin to lipocalin-2, preventing siderophore neutralization and lipocalin-2-dependent inflammation induction. Aerobactin is a siderophore composed of citrate and hydroxamate. Furthermore, the presence of aerobactin, which is occasionally expressed by clinical isolates of

classical nosocomial *K. pneumoniae*, is always associated with hypercapsulation. However, not all hypercapsulated strains have aerobactin [17]. Furthermore, the single microorganism *K. pneumoniae* caused the liver abscess and is a concern as a new invasive syndrome [18]. It has been reported that *K. pneumoniae* has a 180 kbp plasmid containing the gene encoding aerobactin and its receptors, as well as the administration of *rmpA*, a mucoid phenotype. The comprehensive hvKp definition includes hypermucosal viscous phenotype, genotype, and clinical signs of metastatic infection [19].

Adhesins are cell surface factors or bacterial accessories that facilitate adhesion or attachment to other cells or shells, typically on the host where they infect or live. Adhesins are also a type of acridity factor. Adherence is a critical step in the pathogenesis of bacteria or infection that is required for the bacteria or infection to colonize a new host. Bacterial adhesions and adhesions are implicit targets for bacterial infection prevention or treatment. Fimbriae bonds (Fimbriae or pili) are structural proteins that help bacteria target specific kerchief shells in the host. There are three types of fimbriae in K. pneumoniae: 1, 3, and K. pneumoniae carbapenemase (KPC). Type 1 fimbriae are thin, hard, hair-like projections on the bacterial cell's surface. Chaperones/intercellular pathways aggregate them, and the *fim* gene cluster decrypts them [20]. The *fimA* gene encodes the major structural subunit of *fimA*, while the *fimH* gene encodes the small-terminal adhesin subunit of *fimH*. Furthermore, the *fimK* gene is present in *K*. *pneumoniae* but not in Escherichia coli. The fimK gene is involved in the regulation of type 1 fimbriae, and it is important to note that its absence results in the failure of type 1 fimbriae expression [21]. In the urinary tract, the type 1 fimbriae gene is expressed, but not expressed in the gastrointestinal tract or lungs, so these fimbriae can foray into bladder cells and form a biofilm in the bladder. Furthermore, type 3 fimbria are spirally paraphrased by the mrkABCD gene cluster, which may be chromosomal or plasmid-deduced. The *fimH* and *mrkD* genes which render type 1 and type 3 fimbriae independently are responsible for attachment to host cells [22]. These factors are known to contribute to acridity and are responsible for colonization, irruption, and pathogenicity. Alarmingly, some studies reported that multi-drug-resistant, indeed carbapenem-resistant hvKP isolates have surfaced, which is a major public health concern [23, 24]. Furthermore, these are the main fimbriae that are important for biofilm conformation and for clinging to tissues and affinity in K. pneumoniae. The mrkA can bind to abiotic shells similar to medical bias before and after insertion into the patient's body, whereas mrkD can bind to the extracellular matrix [25]. Klebsiella pneumoniae, particularly when hypermucoid, can cause invasive symptoms in a variety of species and is a common cause of mastitis in dairy farms. It can also thrive in a wide

range of hosts and environmental niches, including water and soil. Several studies reported that despite having the *mrkD* and *fimH* genes encoding form 3 and type 1 fimbriae, respectively, hypermucoviscous isolates added serotype K1, demonstrating low upstream adhesion due to the presence of hypercapsule harboring these fimbriae [26].

Lipopolysaccharides and CPS are the two main factors responsible for the pathogenicity of microorganisms. Lipopolysaccharides contain antigens such as lipid A, core, and O-polysaccharide that are required for microorganisms to repel complement-mediated payoff. CPS is the pathogen's most distant subcaste, and it is primarily involved in resistance to phagocytosis by polymorphonuclear cells by acting as a physical hedge [27]. As a result, both factors are necessary for microorganisms to spread through the blood and cause sepsis. However, little is known about how these two factors interact with K. pneumoniae. Due to active immunization with pure CPS-defended mice against the experimentally convinced K. pneumoniae, the experimental confirmation suggests that CPS may be important for K. pneumoniae conformation [28]. Similarly, monoclonal antibodies against Klebsiella CPS were found to reduce the inflexibility and hematogenic spread of K. pneumoniae in a recent study. Furthermore, CPS and LPS may play an important role in the development of necrotic lesions, but their role has not been thoroughly investigated [29].

The polysaccharide capsule prevents phagocytosis while also inhibiting complement-mediated lysis and opsonization. Complete LPS will elicit a strong seditious response, aiding in the transfer of Clq to bacteria and igniting the complement pathway. Furthermore, certain Klebsiella strains can modify LPS so that it cannot be used by vulnerable cells, whereas others can use the capsule to help the toll-like receptor detect LPS (TLR4). Klebsiella pneumoniae also has fimbriae types 1 and 3, which help with adhesion to biotic and abiotic shells, as well as epithelial cell irruption and biofilm conformation. This process also synthesizes siderophores such as aerobactin, enterobactin, salmochelin, and yersiniabactin, to gain iron from the host. Furthermore, through the process of phagocytosis and the production of vulnerable mediators similar to cytokines and chemokines, macrophages play an important role in the ingrain susceptible response. Interleukin (IL)-23 plays an important role in this process by inducing the product of IL-17 and IL-8, which promotes neutrophil reclamation. IL-12 stimulates the expression of IL-17 through IFN-. Another type of cytokine is IL-1, which is produced by activating the seditious pathway for nucleotide oligomerization domain (NOD)-like receptors similar to the pyrin receptor (NLRP3), as well as other pro-inflammatory cytokines such as TNF- and IL-6 (Figure-1) [30].

The LPS is an important element of the external membrane of *Klebsiella*, which is also known



Figure-1: The scheme of host innate immune response and virulence of Klebsiella pneumonia [30].

as an endotoxin conforming to three corridors antigen O, core oligosaccharides, and lipid A. The genes needed for their conformation are located in the wb, waa, and lpx gene groups. Lipopolysaccharide has a vital part in the acridity of these bacteria, where K. pneumoniae can alter lipid A, leading to the inactivation of the seditious response [31]. Furthermore, lipid A blocks the bactericidal effect of antimicrobial peptides. Lipopolysaccharide is the main avenue of defense against complement, wherein strains with full-length O antigen or smooth LPS are resistant to complement-intermediated payoff. In contrast, strains with abbreviated or absent O-chain or crude LPS are sensitive to complement-intermediated payoff indeed in the present capsule [30]. Moreover, it is believed that the part of O antigen is in baffling the attachment of C1q to bacteria, which blocks consecutive stimulants from completing the common pathway by fixing C3b in addition to the bacterial external subcaste and, therefore, frustrated bacterial lysis by the complex attack complement membrane [1].

Lipopolysaccharides are formed from lipid A, oligosaccharide synopsis, O antigens and are known as endotoxins based on all Gram-negative pathogens, including cKP and hvKP. At present, it is not explicit whether the resulting LPS hvKP strain has an individual role in hypervirulence. The outermost subunit based on LPS, O antigen is the primary constituent faced by the innate immune system and protects bacteria against complement-mediated inflammation. In particular, the O antigens bind to the complementary constituent C3b, which involves pore arrangement before mediating the drilling of pathogenic tissue. The number of O serotypes is estimated to be eight, and O1 antigen is the most common among clinical strains of *K. pneumoniae* [32].

During infection, *K. pneumoniae* overcomes mechanical and chemical barriers as well as cellular

and humoral host defenses. Furthermore, once inside the host, the host organism attacks the vulnerable cell, which is linked to pattern recognition. Following recognition, the receptor activates the product of central vulnerable intercessors, and the ingrain vulnerable response participates in the monocyte/macrophage system. Furthermore, this system is capable of phagocytosis and regulates the vulnerable response to cytokines and product chemokines. Neutrophils are immune cells that are the first to respond to an infection. Interleukin-8 and IL-23, which both play a role in converting a granulopoietic response, are important cytokine proteins in this phase [1]. Interleukin-12 stimulates the expression of IL-17 by producing interferon-gamma. The product of IL-1 participates in the vulnerable response through activation of the pyrine-containing NOD receptor sphere (NLRP3) in the seditious pathway, as do the products of other pro-inflammatory cytokines such as TNF- and IL-6 [33].

Capsule polysaccharides, LPSs, fimbriae, and siderophores are well-studied virulence factors [1]. Type 1 and type 3 fimbriae of K. pneumoniae are equipped with adhesins and are useful for epithelial attachment and cell impunity, as well as abiotic shells. Seventy-eight different capsule serotypes (K1 to K78) have been linked to K. pneumoniae based on the structure of the CPs. The hvKp strains are represented by serotypes K1 and K2. The high acridity position of the hvKp strain is due to an excess of capsular material (hypermucoviscos phenotype). Although, the capsule is important in protecting K. pneumoniae from the host's vulnerable response. Furthermore, certain K. pneumoniae strains can convert LPS to situations that are not recognized by host cells, while others can use the capsule to conceal LPS from detection by TLR4 [23, 30]. The ability of K. pneumoniae to take iron from the host explains their growth and

replication, which has been explained by iron motes or siderophores. The hvKP strain can also produce significantly less iron accession and is a more biologically active strain than other non-virulent K. pneumoniae strains. Furthermore, no natural marker has yet been identified that can distinguish hvKp from other K. pneumoniae strains. Furthermore, MDR hvKP can be caused by two distinct mechanisms. The hvKP strain was able to accept antibacterial agent horizontal gene transfer resistance genes or plasmids, it temporarily became MDR hvKP type I [34]. Multidrug resistance hvKP strains can also be derived from pathogenic plasmids such as pLVPK. Multidrug resistance hvKP type II is the most common MDR strain of K. pneumoniae. A recent study in China revealed fatalities. Furthermore, KPC-producing ST11 strains induced the acquisition of pLVPK-like pathogenic plasmid [35]. Klebsiella pneumoniae can form microbial cells that irreversibly cleave to the cell's surface or within the cell to polymeric extracellular matrix substances. Fimbria type 1 and fimbria type 3 are polysaccharide capsules that contribute to the biofilm formation process and are acridity factors. The process starts with the formation of a biofilm by fimbria type 3, which is made up of mrkA protein subunits. Furthermore, mrkD is another element located at the tip of the fimbria that functions as a complement to give tenacious parcels and also determines the fimbria's listed capacity [36].

In addition, despite the inherent resistance to ampicillin, hvKp strains are generally susceptible to various antibiotics, including cephalosporins and carbapenems. However, MDR and highly toxic strain (MDRhv) have recently emerged, mainly due to plasmid-mediated horizontal transmission of pathogens or resistance [37, 38]. Further, for ambiguous variant potential pathogens, mice, but not *Galleria melonella* is employed. *Galleria melonella* mortality test combined with the string test yields an accurate phenotype, and the test proves to improve clinical identification [39, 40].

Klebsiella pneumoniae forms a biofilm and is multidrug-resistant. Fimbriae influence adhesion stability, while CP influences cell-to-cell communication and biofilm structure. The bedded cell must be capable of carrying out rapid-fire and expansive changes in gene expression for the production process of biofilm conformation and variability of stimulants from the terrain. Klebsiella pneumoniae cells in the biofilm are only partially protected against vulnerable defenses. The matrix prevents antibacterial antibodies and peptides from reaching the bacteria, as well as reducing or suppressing the effectiveness of complement and phagocytosis [41]. The formation of K. pneumoniae biofilms on solid shells was characterized by cell adherence, microcolony conformation, and eventually the dissolution of free-living cells. Type 3 fimbria and CP are important corridors in the process of forming bacterial face structures, which directly and

laboriously have the capability of suppressing seditious responses and changing the vulnerable system in habitual infections, to determine resistance, the most important factor being the growth status of bacteria. The core part of the biophilic structure to which bacteria adapt to starvation and low oxygen environments causes bacterial growth to slow, reducing the efficacy of antibiotics that directly target metabolically active and dividing cells. In *K. pneumoniae*, apparent quorum detectors and autoinducers have been described [20].

### The Emergence of Multidrug-resistant Food

The medium of exchange between antibiotic resistance and acridity in *K. pneumoniae* is unknown. *Klebsiella pneumoniae* is frequently associated with nosocomial and salutary infections; this case has also been reported as a possible vector of transmission [42]. *Klebsiella pneumoniae* is no longer found in raw meat, raw vegetables, fruit authorities, or ready-to-eat (RTE) foods. Numerous studies on *K. pneumoniae* in food have been conducted. Antibiotic resistance is of particular concern, with foodborne *K. pneumoniae* being resistant to three or more classes of antibiotics (MDR) [5]. Joint and inter-agency sweats are required to address the issue of antibiotic-resistant bacteria in the food supply chain for public health.

K. pneumoniae is found in the normal foliage of animals and humans, food impurity may not be a threat to public health. The discovery of K. pneumoniae in food can be related to the hygienic running of food, similar to undercooking or post-cooking impurity, especially for RTE foods. Furthermore, K. pneumoniae has also been described to be dominant in husbandry, as K. pneumoniae has been shown in previous studies to increase yields under agrarian conditions [43]. In addition, numerous factors, such as the presence of raw vegetables, contribute to the presence of K. pneumoniae in food; the addition of organic diseases contributes to the presence of K. pneumoniae in food. The discovery of K. pneumoniae in raw foods in original requests and supermarkets emphasizes the importance of fresh food safety measures. Moreover, the cross-contamination with bacteria from the raw yield, meat authorities, or other polluted products, or from food instructors with poor particular hygiene, can contaminate cooked RTE foods, even if cooked or packaged safely [44].

*Klebsiella pneumoniae* can be spread through person-to-person contact during the food medication process. As a result, the general public and food consumers should be concerned, because they play a role in the spread (or control) of complaint-causing bacteria. The factors of genetic origin and geographical conditions are often risk factors in this situation [45]. MDR distribution follows a similar geographic pattern and is mostly found in Asian countries. However, it can be found all over the world and has been documented [38, 46].

According to the World Health Organization, food is an implicit means of transmission of

Food	virulence Factor	Antibiotic resistance/Resistance gene variant	Reference
Cooked meat	-	bla-B-10	[54]
Raw meat	-	<i>bla</i> CTX-M-15, <i>bla</i> SHV28, <i>bla</i> Tem-1B	
Noodles	K54, <i>wca</i> G	-	
Beverage	K2	-	
Pork dishes	wcaG	-	
Chili's	K, <i>wca</i> G	AMP-TE-AK-C-SXT	
Lettuce	K1, wcaG	-	
Pork liver	K2	AMP-C-CIP-TE-SXT	
Porridge	wcaG	-	
Black tiger shrimp	-	blaCTX, blaSHV	
Cooked and raw meat	-	blaOKP-B-10	[55]
Uncooked vegetables	-	aac(3)-IIc	[56]

Table-1: Factors contributing to toxicity from different kinds of food.

antimicrobial-resistant bacteria to humans, and consumption of food containing antimicrobial-resistant bacteria has resulted in antimicrobial resistance. These are global antimicrobial resistance measures, resistant infections, acridity characteristics, and antibiotic resistance biographies required to assess the public health threat posed by foodborne K. pneumoniae. According to reports, the most concerning health issue is antimicrobial resistance in K. pneumoniae [47, 48]. Foodborne pandemics have occurred in recent decades, reminding us of the importance of enforcing high hygiene standards as well as preventive measures and forestalment programs aimed at food safety for the community, which applies to food products aseptic. Moreover, approaches have also been taken in agriculture and all food production sectors to improve hygiene and reduce natural hazards. The differences in food composition and food processing can all play a role in the emergence of foodborne pathogens. Many cases of foodborne complaints have been reported due to factors such as changes in beneficial habits, increased cross-country travel, changes in food product processing and distribution processes, pathogen adaptation to new environments, antimicrobial resistance by microorganisms, advances in pathogen discovery, sanitation, and vector control measures, poor public health services, and consumer information [6].

An infectious disease epidemic necessitates three critical components: The source, the pathway, and the vulnerable population. In terms of habitat relevance, K. pneumoniae has been found in humans, animals, sewage, contaminated water samples, and soil. The origin of hvKP, on the other hand, is unknown [49]. Foods similar to dairy products can be a source of transmission of Enterobacteriaceae microorganisms that exhibit MDR to antibiotics and other acridity factors similar to biofilm products, as well as the conflation of proteolytic and lipolytic enzymes that are responsible for corruption processes in food products. Good hygiene conditions during processing and manufacturing, as well as storage and distribution processes can reduce or eliminate the presence of these microorganisms [50]. The emergence of antimicrobial resistance in K. pneumoniae is a major concern in life-saving drugs around the

world. *Klebsiella pneumoniae* multidrug-resistant strains have been identified and isolated from various samples [51].

Food is one of the factors that contribute to the occurrence of antibiotic-resistant bacteria and their genes in the mortal digestive tract. The prevalence of foodborne illnesses caused by K. pneumoniae has recently increased. There is currently little information available on the characteristics of K. pneumoniae that has been isolated from food. Certain food orders may influence the diversity of gut antibiotic resistance genes [52, 53]. Table-1 shows data on the factors that contribute to the toxicity of various types of food [54-56]. Furthermore, these bacteria can transfer the genes that determine antibiotic resistance to other types of pathogenic bacteria. As a result, surveillance and monitoring of antimicrobial-resistant bacteria in food are critical for enforcing targeted control strategies and selecting effective treatment options [57-59].

#### Conclusion

The emergence of MDR strains and the rise of *K. pneumoniae* have compelled scientists and researchers to seek out and define newer antibacterial treatments. There is a clear need to define and engage the public to increase surveillance of *K. pneumoniae* in food, as well as to improve our understanding of the epidemiological and public health implications of this foodborne pathogen. As a result, it is critical to recognize that *K. pneumoniae* is a major disease pathogen that can be transmitted through the food chain, and this must be done immediately.

#### **Authors' Contributions**

KHPR: Conceived the idea and drafted and revised the manuscript. MHE and FAR: Reviewed the manuscript. KHPR, AW, and ARK: Literature searches and edited and reviewed the manuscript. All authors have read and approved the final manuscript.

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#### **Competing Interests**

The authors declare that they have no competing interests.

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#### References

- 1. Paczosa, M.K. and Mecsas, J. (2016) *Klebsiella pneumoniae*: Going on the offense with a strong defense. *Microbiol. Mol. Biol. Rev.*, 80(3): 629–661.
- Oikonomou, K.G. and Aye, M. (2017) *Klebsiella pneumoniae* liver abscess: A case series of six Asian patients. *Am. J. Case Rep.*, 18(1): 1028–1033.
- Ku, Y.H., Chuang, Y.C., Chen, C.C., Lee, M.F., Yang, Y.C., Tang, H.J. and Yu, W.L. (2017) *Klebsiella pneumoniae* Isolates from meningitis: Epidemiology, virulence and antibiotic resistance. *Sci. Rep.*, 7(1): 6634.
- 4. Russo, T.A. and Marr, C.M. (2019) Hypervirulent *Klebsiella* pneumoniae. Clin. Microbiol. Rev., 32(3): e00001–19.
- Guo, Z.H., Qin, L., Pang, Z., Qin, T. and Ren, H. (2016) Frequency, antimicrobial resistance and genetic diversity of *Klebsiella pneumoniae* in food samples. *PLoS One*, 11(4): e0153561.
- 6. Smith, J.L. and Fratamico, P.M. (2018) Emerging and re-emerging foodborne pathogens. *Foodborne Pathog. Dis.*, 15(12): 737–757.
- Hanashiro, A., Morita, M., Matte, G.R., Matte, M.H. and Torres, E.A.F. (2005) Microbiological quality of selected street foods from a restricted area of Sao Paulo city, Brazil. *Food Control*, 16(5): 439–444.
- Gorrie, C.L., Mirceta, M., Wick, R.R., Edwards, D.J., Thomson, N.R., Strugnell, R.A., Pratt, N.F., Garlick, J.S., Watson, K.M., Pilcher, D.V., McGloughlin, S.A., Spelman, D.W., Jenney, A.W.J. and Holt, K.E. (2017) Gastrointestinal carriage is a major reservoir of *Klebsiella pneumoniae* infection in intensive care patients. *Clin. Infect. Dis.*, 65(2): 208–215.
- Davis, G.S. and Price, L.B. (2016) Recent research examining links among *Klebsiella pneumoniae* from food, food animals, and human extraintestinal infections. *Curr. Environ. Health Rep.*, 3(2): 128–135.
- Wei, D.D., Chen, K.Q. and Wang, L.H. (2016) Clinical and molecular characteristics of high virulent *Klebsiella pneumonia* in infection in intensive care unit. *Chin. J. Nosocomiol.*, 26(1): 5056–5059.
- Pan, Y.J., Fang, H.C., Yang, H.C., Lin, T.L., Hsieh, P.F., Tsai, F.C., Keynan, Y. and Wang, J.T. (2008) Capsular polysaccharide synthesis regions in *Klebsiella pneumoniae* serotype K57 and a new capsular serotype. *J. Clin. Microbiol.*, 46(7): 2231–2240.
- 12. Parrott, A.M., Shi, J., Aaron, J., Green, D.A., Whittier, S. and Wu, F. (2021) Detection of multiple hypervirulent *Klebsiella pneumoniae* strains in a New York City hospital through screening of virulence genes. *Clin. Microbiol. Infect.*, 27(4): 583–589.
- Sugumar, M., Kumar, K.M., Manoharan, A., Anbarasu, A., and Ramaiah, S. (2014) Detection of OXA-1 β-Lactamase Gene of Klebsiella pneumoniae from Blood Stream Infections (BSI) by Conventional PCR and In-Silico Analysis to Understand the Mechanism of OXA Mediated Resistance. *PLoS One*, 9(3): e91800.
- Meatherall, B.L., Gregson, D., Ross, T., Pitout, J.D. and Laupland, K.B. (2009) Incidence, risk factors, and outcomes of *Klebsiella pneumoniae* bacteremia. *Am. J. Med.*, 122(9): 866–873.
- 15. El Fertas-Aissani, R., Messai, Y., Alouache, S. and

Bakour, R. (2013) Virulence profiles and antibiotic susceptibility patterns of *Klebsiella pneumonia* strains isolated from different clinical specimens. *Pathol. Biol. (Paris)*, 61(5): 209–216.

- Lan, P., Shi, Q., Zhang, P., Chen, Y., Yan, R., Hua, X., Jiang, Y., Zhou, J. and Yu, Y. (2020) Core genome allelic profiles of clinical *Klebsiella pneumoniae* strains using a random forest algorithm based on multilocus sequence typing scheme for hypervirulence analysis. *J. Infect. Dis.*, 221(Suppl 2): S263–S271.
- Bachman, M.A., Oyler, J.E., Burns, S.H., Caza, M., Lepine, F., Dozois, C.M. and Weiser, J.N. (2011), *Klebsiella pneumoniae* yersiniabactin promotes respiratory tract infection through evasion of lipocalin 2. *Infect. Immun.*, 79(8): 3309–3316.
- Siu, L.K., Yeh, K.M., Lin, J.C., Fung, C.P., and Chang, F.Y. (2012) *Klebsiella pneumoniae* liver abscess: A new invasive syndrome. *Lancet Infec. Dis.*, 12(11): 881-887.
- Catalán-Nájera, J.C., Garza-Ramos, U. and Barrios-Camacho, H. (2017) Hypervirulence and hypermucoviscosity: Two different but complementary *Klebsiella* spp. phenotypes? *Virulence*, 8(7): 1111–1123.
- 20. Clegg, S. and Murphy, C.N. (2016) Epidemiology and virulence of *Klebsiella pneumoniae*. *Microbiol. Spectr.*, 4(1): 1-12.
- Rosen, D.A., Hilliard, J.K., Tiemann, K.M., Todd, E.M., Morley, S.C. and Hunstad, D.A. (2015) *Klebsiella pneumoniae* FimK promotes virulence in murine pneumonia. *J. Infect. Dis.*, 213(4): 649–658.
- 22. Shah, R.K., Ni, Z.H., Sun, X.Y., Wang, G.Q. and Li, F. (2017) The determination and correlation of various virulence genes, ESBL, serum bactericidal effect and biofilm formation of clinical isolated classical *Klebsiella pneumoniae* and hypervirulent *Klebsiella pneumoniae* from respiratory tract infected patients. *Pol. J. Microbiol.*, 66(4): 501–508.
- Guo, Y., Wang, S., Zhan, L., Jin, Y., Duan, J., Hao, Z., Lv, J., Qi, X., Chen, L., Kreiswirth, B.N., Wang, L. and Yu, F. (2017) Microbiological and clinical characteristics of hypermucoviscous *Klebsiella pneumoniae* isolates associated with invasive infections in China. *Front Cell Infect. Microbiol.*, 7(1): 24.
- Zhang, R., Lin, D., Chan, E.W.C., Gu, D., Chen, G.X. and Chen, S. (2016) Emergence of carbapenem-resistant serotype K1 hypervirulent *Klebsiella pneumoniae* strains in China. *Antimicrob. Agents Chemother.*, 60(1): 709–711.
- Sugawara, E., Kojima, S. and Nikaido, H. (2016) *Klebsiella pneumoniae* major porins OmpK35 and OmpK36 allow more efficient diffusion of lactams than their *Escherichia coli* homologs OmpF and OmpC. *J. Bacteriol.*, 198(23): 3200–3208.
- Cubero, M., Marti, S., Domínguez, M., González-Díaz, A., Berbel, D. and Ardanuy, C. (2019) Hypervirulent *Klebsiella pneumoniae* serotype K1 clinical isolates form robust biofilms at the air-liquid interface. *PLoS One*, 14(9): e0222628.
- Williams, P. and Tomás, J.M. (1990) The pathogenicity of Klebsiella pneumoniae. Rev. Med. Microbiol., 1(1): 196–204.
- Cryz, S.J. Jr., Fürer, E. and Germanier, R. (1986) Immunization against fatal experimental *Klebsiella pneumoniae* pneumonia. *Infect. Immun.*, 54(2): 403–407.
- Straus, D.C., Atkisson, D.L. and Garner, C.W. (1985) Importance of lipopolysaccharide-containing extracellular toxic complex in infections produced by *Klebsiella pneumoniae*. Infect. Immun., 50(3): 787–795.
- Piperaki, E.T., George, A.S., Leonidas, S.T. and George, L.D. (2017) *Klebsiella pneumoniae*: Virulence, biofilm and antimicrobial resistance. *Pediatr: Infect. Dis. J.*, 36(10): 1002–1005.
- Llobet, E., Martinez-Moliner, V., Moranta, D., Dahlstrom, K. M., Regueiro, V., Tomas, A., Cano, V., Pérez-Gutiérrez, C., Frank, C.G., Fernández-Carrasco, H., Insua, J.L., Salminen, T.A., Garmendia, J. and Bengoechea, J.A. (2015) Deciphering tissue induced *Klebsiella pneumoniae* lipid A structure. *Proc. Natl. Acad. Sci. U. S. A.*, 112(46): E6369–E6378.
- 32. Follador, R., Heinz, E., Wyres, K.L., Ellington, M.J., Kowarik, M., Holt, K.E. and Thomson, N.R. (2016) The

diversity of *Klebsiella pneumoniae* surface polysaccharides. *Microb. Genom.*, 2(8): e000073.

- Hua, K.F., Yang, F.L., Chiu, H.W., Chou, J.C., Dong, W.C., Lin, C.N., Lin, C.Y., Wang, J.T., Li, L.H., Chiu, H.W., Chiu, Y.C. and Wu, S.H. (2015) Capsular polysaccharide is involved in NLRP3 inflammasome activation by *Klebsiella pneumoniae* serotype K1. *Infect. Immun.*, 83(9): 3396–3409.
- 34. Liu, Y., Long, D., Xiang, T.X., Du, F.L., Wei, D.D., Wan, L.G., Deng, Q., Cao, X.W. and Zhang, W. (2019) Whole genome assembly and functional portrait of hypervirulent extensively drug-resistant NDM-1 and KPC-2 co-producing *Klebsiella pneumoniae* of capsular serotype K2 and ST86. J. Antimicrob. Chemother., 74(5): 1233–1240.
- 35. Gu, D., Dong, N., Zheng, Z., Lin, D., Huang, M., Wang, L., Chan, E.W.C., Shu, L., Yu, J., Zhang, R. and Chen, S. (2018) A fatal outbreak of ST11 carbapenem-resistant hypervirulent *Klebsiella pneumoniae* in a Chinese hospital: A molecular epidemiological study. *Lancet Infect. Dis.*, 18(1): 37–46.
- 36. Murphy, C.N. and Clegg, S. (2012) *Klebsiella pneumoniae* and Type 3 fimbriae: Nosocomial infection, regulation and biofilm formation. *Future Microbiol.*, 7(8): 991–1002.
- 37. Liu, C., Du, P., Xiao, N., Ji, F., Russo, T.A. and Guo, J. (2020) Hypervirulent *Klebsiella pneumoniae* is emerging as an increasingly prevalent *K. pneumoniae* pathotype responsible for nosocomial and healthcare-associated infections in Beijing, China. *Virulence*, 11(1): 1215–1224.
- Tang, M., Kong, X., Hao, J. and Liu, J. (2020) Epidemiological characteristics and formation mechanisms of multidrug-resistant hypervirulent *Klebsiella pneumoniae. Front. Microbiol.*, 11(1): 581543.
- 39. Russo, T.A. and MacDonald, U. (2020) The *Galleria mellonella* infection model does not accurately differentiate between hypervirulent and classical *Klebsiella pneumoniae*. *mSphere*, 5(1): e00850–19.
- Li, G., Shi, J., Zhao, Y., Xie, Y., Tang, Y., Jiang, X. and Lu, Y. (2020) Identification of hypervirulent *Klebsiella pneumoniae* isolates using the string test in combination with *Galleria mellonella* infectivity. *Eur. J. Clin. Microbiol. Infect. Dis.*, 39(9): 1673–1679.
- Hughes, K.A., Sutherland, I.W. and Jones, M.V. (1998) Biofilm susceptibility to bacteriophage attack: The role of phage-borne polysaccharide depolymerase. *Microbiology* (*Reading*), 144(Pt 11): 3039–3047.
- Calbo, E., Freixas, N., Xercavins, M., Riera, M., Nicolás, C., Monistro, O., Solé, M.D.M., Sala, M.R., Vila, J. and Garau J. (2011) Foodborne nosocomial outbreak of SHV1 and CTX-M-15–producing *Klebsiella pneumoniae*: Epidemiology and control. *Clin. Infect. Dis.*, 52(6): 743–749.
- 43. Epstein, E. (2015) Disposal and Management of Solid Waste: Pathogens and Diseases. CRC Press, Boca Raton, FL.
- 44. US Department of Agriculture, Food Safety and Inspection Service. (2011) Foodborne Illness: What Consumers need to know. US Department of Agriculture, Food Safety and Inspection Service, United States.
- 45. Sellick, J.A. and Russo, T.A. (2018) Getting hypervirulent *Klebsiella pneumoniae* on the radar screen. *Curr. Opin. Infect. Dis.*, 31(4): 341–346.
- Li, B., Zhao, Y., Liu, C., Chen, Z. and Zhou, D. (2014) Molecular pathogenesis of *Klebsiella pneumoniae*. *Future Microbiol.*, 9(9): 1071–1081.

- Riwu, K.H.P., Effendi, M.H. and Rantam, F.A. (2020) A review of extended-spectrum β-lactamase (ESBL) producing *Klebsiella pneumoniae* and multidrug-resistant (MDR) on companion animals. *Syst. Rev. Pharm.*, 11(7): 270–277.
- 48. Permatasari, D.A., Witaningrum, A.M., Wibisono, F.J. and Effendi, M.H. (2020) Detection and prevalence of multidrug-resistant *Klebsiella pneumoniae* strains isolated from poultry farms in Blitar, Indonesia. *Biodiversitas*, 21(10): 4642–4647.
- 49. Liu, B.T., Zhang, X.Y., Wan, S.W., Hao, J.J., Jiang, R.D. and Song, F.J. (2018) Characteristics of carbapenem-resistant *Enterobacteriaceae* in ready-to-eat vegetables in China. *Front. Microbiol.*, 9(1): 1147.
- Amorim, A.M.B. and Janaína-dos Santos, N. (2017) A highlight for non-*Escherichia coli* and non-*Salmonella* spp. *Enterobacteriaceae* in dairy foods contamination. *Front. Microbiol.*, 8(1): 930.
- Effendi, M.H., Bintari, I.G., Aksoro, E.B. and Hermawan, I.P. (2018) Detection of blaTEM gene of *Klebsiella pneumoniae* Isolated from swab of food-producing animals in East Java. *Trop. Anim. Sci. J.*, 41(3): 174–178.
- 52. Milanovic, V., Osimani, A., Aquilanti, L., Tavoletti, S., Garofalo, C., Polverigiani, S., Litta-Mulondo, A., Cocolin, L., Ferrocino, I., Di Cagno, R., Turroni, S., Lazzi, C., (2017) Occurrence of antibiotic resistance genes in the faecal DNA of healthy omnivores, Ovo-Lacto vegetarians and vegans. *Mol. Nutr. Food Res.*, 61(1): 1601098.
- 53. Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Short communication: Pattern of antibiotic resistance on extended-spectrum beta-lactamases genes producing *Escherichia coli* on laying hens in Blitar, Indonesia. *Biodiversitas*, 21(10): 4631–4635.
- Hartantyo, S.H.P., Chau, M.L., Koh, T.H., Yap, M., Yi, T., Cao, D.Y.H., GutiÉrrez, R.A. and Ng, L.C. (2020) Foodborne *Klebsiella pneumoniae*: Virulence potential, antibiotic resistance, and risks to food safety. *J. Food Prot.* 83(7): 1096–1103.
- Liu, Y., Cui, Y., Peng, W., Huang, B., Ma, L., Zheng, M., Ding, S. and Zhu, K. (2020) Prevalence of pathogens harbouring mobile antimicrobial resistance genes and virulence factors in retail beef and mutton. *FEMS Microbiol. Lett.*, 367(12): fnaa089.
- Boehme, S., Werner, G., Klare, I., Reissbrodt, R. and Witte, W. (2004) Occurrence of antibiotic-resistant enterobacteria in agricultural foodstuffs. *Mol. Nutr. Food Res.*, 48(7): 522–531.
- 57. Effendi, M.H., Tyasningsih, W., Yurianti, Y.A., Rahmahani, J., Harijani, N. and Plumeriastuti, H. (2021) Presence of multidrug resistance (MDR) and extended-spectrum beta-lactamase (ESBL) of *Escherichia coli* isolated from cloacal swabs of broilers in several wet markets in Surabaya, Indonesia. *Biodiversitas*, 22(1): 304–310.
- Wibisono, F.M., Wibisono, F.J., Effendi, M.H., Plumeriastuti, H., Hidayatullah, A.R., Hartadi, E.B. and Sofiana, E.D. (2020) A review of salmonellosis on poultry farms: Public health importance. *Sys. Rev. Pharm.*, 11(9): 481–486.
- 59. Rahmahani, J., Salamah, S., Mufasirin, M., Tyasningsih, W. and Effendi, M.H. (2020) Antimicrobial resistance profile of *Escherichia coli* from cloacal swab of domestic chicken in Surabaya traditional market. *Biochem. Cell. Arch.*, 20(1): 2993–2997.

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