

Hematological and immunological effect on chicken exposed to aflatoxin

Alo Odunayo Samuel¹, Oyebanji Olubukola² and Abatan Oluwole Matthew³

Faculty of Agriculture
Department of Animal Production and Health Sciences, University of Ado. **Nigeria**

Abstract

Chickens were exposed to aflatoxin sourced from rice substrate. Enzyme immunosorbent assay was used to evaluate the immune response to Newcastle disease vaccinations vis a vis the hematological pictures. Results show a dose dependent reduction in immune titer to Newcastle disease. Hematological findings confirm leucopenia, thrombocytopenia, with increased PCV and hemoglobin. Also increased is the heterophils which was masked by other leucocytes reduction. This hematological picture and immunosuppression is being linked to increased glucocorticoid in the subjects.

Keywords: Aflatoxin, Dose, Immunity, Newcastle, Titre.

Introduction

Studies on mycotoxin date back to turkey poult deaths recorded in England due to consumption of contaminated groundnut meal imported from Brazil {Blout 1961}. Aflatoxin is acutely toxic, mutagenic, carcinogenic. {Oguz and Kortoglu 2000} in addition aflatoxin has been implicated in immunosuppression. {Bondy 2000, Omigen 2004}.

The West African countries have tropical climates with all year round high ambient temperature and relative humidity that provides optimal condition for the growth of toxigenic moulds. The sub region has poorly developed infrastructures such as processing and storage facilities, transportation and skilled human resources. {Bankole and Adebayo 2003}.

Aflatoxin has been fingered as a strong immunosuppressive agent. {Omnigent research update 2004}. The complexity of the immune system implies that there are several methods by which immune function may be assayed. Simultaneous studies on hematological and antibody profile of chicken subjected to aflatoxin exposure will reveal the interplay of cellular and humoral factors in aflatoxin immunosuppression.

Materials and Methods

Aspergillus flavus La32g38, a toxigenic strain was procured from International Institute for Tropical Agriculture [IITA, pathology department]. It was inoculated with rice substrate to produce crude

aflatoxin according to the method of Shotwell 1966. Quantification of aflatoxin was done with the aid of ROSA® Aflatoxin kit [quantitative]. The feed was supplemented with the rice substrate to give 50-100 ppb in the treatment diet.

Ninety day old chicks were used for the study. They were divided into three groups of thirty birds with three replicates, making ten birds per experimental unit.

They received experimental diets and water ad libitum for 42 days. The experimental groups include Control

2 basal diet plus 50 ppb aflatoxin, in feed.

3 basal diet plus 100 ppb aflatoxin in feed.

Hematology

Differential WBC counts were made on mono layer blood films and stained with Giemsa – Wright's stain. Total red blood cell [TRBC], Total white blood cell count were determined by manual method using hemacytometer [Campbell 1995]. Packed Cell Volume [PCV] was measured by a capillary tube technique using microhematocrit capillary tubes centrifuged at 2500 rpm for 5 min. Hemoglobin concentration was measured by cyanomethemoglobin method. [Ganti 2000] Erythrocyte indices, mean corpuscular volume [MCV], mean corpuscular hemoglobin [MCH] and mean corpuscular hemoglobin concentration [MCHC] were calculated from TRBC, MCV and Hb. [Ritchie et al 1944].

1. Corresponding author e mail; metrovetgroup@yahoo.com, Tel. 234-08038434432.

2. Faculty of Agriculture, Department of Animal Science, Obafemi Awolowo University, Ile Ife, Nigeria.

3. Faculty of Veterinary Medicine, Department Of Veterinary Physiology and Pharmacology, University of Ibadan, Nigeria.

Table-1.

Group	PCV %	cell x 10 ¹² /L RBC±SE	Cellx10 ⁶ /L plats ±SE	Cellx10 ⁶ /L TWBC±SE	Cellx10 ⁶ /L CellxLYMP ±SE	Cellx10 ⁶ /L mono±SE	Cellx10 ⁶ /L MCHC±SE	Cellx10 ⁶ /L Eosin±SE	Cellx10 ⁶ /L Hetero±SE	G/gI HB ±SE
Control	24.7±1.0	2.0±0.23	102857±1463	16964±971.4	11778±714	391.29±56.0	32.7±0.42	392.86±51.3	4401.9±328.9	8.0±0.3
50ppb	23.1±1.08	2.0±0.23	138000±1463	15943±971.4	10.327±714	373.86±56.0	32.14±0.42	296.86±51.3	4934.9±328.9	7.6±0.3
100ppb	23.4±1.08	2.6±0.23	135000±14633	15557±971	10.09±714	204.14±56.0	32.14±0.42	276.71±51.3	5157.4±328.9	7.5±0.5

Serology.

Flockcheck Newcastle disease antibody test kit [Idexx Animal health] was used to provide a sensitive and specific method of antibody quantification. Statistical analysis.

All data generated were subjected analysis of variance [steel and toorie 1980] and means were separated by Duncan's multiple ranjge test. [Duncan 1995].

Results

Table-1 shows a dose dependent decrease in wbc, lymphocytes, monocytes, eosinophils, only decrease in wbc and lymphocytes is statistically significant. (p<0.05).

Discussion

The blood picture of chicken subjected to low doses of aflatoxin 50ppb and 100ppb is suggestive of lymphopenia, monocytopenia, eosinopenia, and heterophilia.

The total white blood cells count suggested leucopenia. These observations were based on comparison with control which were statistically significant. [p<0.05]

The immunosuppressive effect of aflatoxin is well documented. This supports the reduced immune titre (newcastle disease) of chicken exposed to aflatoxin in a dose dependent pattern. It gives basis to the hypothesis that aflatoxins effect on immunity is both cell-mediated and humoral. Of significant scientific importance is the heterophila despite generalised lymphopenia. This work suggests that heterophils [neutrophils in other animals] are peculiarly sensitised to aflatoxin intoxication as distinct from other granulocytes and lymphocytes. Dannis et al(2003) reported increased total WBC after aflatoxin dosing based on his work on rats which has higher number of

neutrophils which thus probably masked reduced lymphocytes .

The increased heterophil in this work has been effectively masked by reduced lymphocytes with resultant generalised leucopenia. In his works on wild turkeys poults, Quist et al(2000) observed increased PCV, WBC and lymphocyte. He claimed the increases in these parameters are either a direct or indirect effects of aflatoxin.

The numerical increase though not statistically significant in PCV, RBC and thrombocytes suggest an interplay of factors sequel to aflatoxin dosing. It is hypothesised sequel to this work, that the hematological picture following aflatoxin intoxication may not be a primary lesion but a secondary sequela to the general 'stressor' effects of a noxious chemical on the physiology of the animal. This becomes more evident, based on the fact that this hematology typifies glucocorticoid exposure. Glucocorticoids lowers the number of basophils in the circulation and increase the number of neutrophils, platelets and red blood cells. It also decreases the circulating lymphocyte count and the size of the lymph nodes and thymus by inhibiting lymphocyte mitotic activity. They also reduce secretion of cytokines by inhibiting the effect of NF-B on the nucleus. The reduced secretion of the cytokines K-2 leads to reduced proliferation of lymphocytes and these cells undergo apoptosis. (William 1990). The increased RBC, Hb and MCHC also correlate with stimulatory action of steroid on the renal erythropoietin secretion. (Bertram 1998).

It could be deduced from this study that hematological profile following aflatoxin intoxication may not be a primary lesion but a secondary sequel a to the general "stressor" effect of noxious chemical on the physiology of the animal. We hypothesize that such continuous stressor stimuli of aflatoxin resulting

Table-2. Effect of 50ppb and 100 ppb aflatoxin [AF] on newscastel disease antibody titre for cockrels on day 1 to 42 days of age.

Control	50	100	Titer at day old	Titre at day 42
+	-	-	599±74	531±51.2
-	+	-	599±74	437±29.2
-	-	+	599±74	403.7±22.0

in immunosuppression may form additional basis for aflatoxin carcinogenicity.

References

1. Bankole S.A. and Adebayo A. (2003): Mycotoxins in foods in West Africa: current situation and possibilities of controlling it. African journal of Biotechnology 2(9):254-263.
2. Bertram G.K. (1998): Basic and clinical pharmacology Applention and large Connecticut Bondy GS Pesca,(2000). Immunomodulation by fungal Toxin.J. Toxicol, Environ Health B, Crit.Rev 3: 109-43
3. Bondy GS Pesca,(2000): Immunomodulation by fungal Toxin.J. Toxicol, Environ Health B, Crit.Rev 3: 109-43
4. Dennis MH Micheal. J M Richard AR Sabine FC Rene ES Joseph S Alan W. And ming. WC (2003): Immunotoxicity of Aflatoxin B1 in Rats ; Effects on lymphocytes and the inflammatory response in a chronic intermitent Dosing study. Toxicological Sciences 73:362-377.
5. Duncan D.B. (1995): Multiple range and multiple F-Test Biometrics 42:1-42.
6. Duncan D.B. (1995): Multiple range and multiple F-Test Biometrics 42:1-42.
7. Ganti A.S. (2004): Veterinary clinical pathology CBS publishers and Distributors,New Delhi.
8. Nemi C.J.(1993): Essentials of Veterinary hematology Lea and Febiger.Philadelphia.
9. Oguz H.Hadimli HH,Kurtoglu V.and Erganis O.(2003): Evaluation of humoral immunity of Broilers during chronic Aflatoxin (50 and 100 ppb)and clinoptilolite exposure. Revue med vet 1547,483-486.
11. Oliveira,Marize S,Prado ,Guilherme and JuinQuira,Roberto G. (2000): Cienco. Tecnol. Aliment 2 (3): 369-374.
12. Shotwell O.L., hesseltine CV, Stable field R.D.and Sorenson W.G.(1996): Production of aflatoxin on rice.Appl Microbiol. 14: 425-429.
13. Steel RCD and torrie JH (1980): Principles and procedures of statistics. A biometrical Approach MC Graw Hill,New york.
14. Quist C F,Bounous D I,Kilburn S U, Nettle V F and Wyatts R D.(2000): Effect of dietary Aflatoxin on wild Turkey poults.Journal of wildlife disease. 36(3): 436-444.
