

## Neutrophils in tuberculosis: will code be unlocked ?

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### Abstract

Tuberculosis is a devastating disease throughout the world both in humans and animals. Its history is vast, which dates back to era of Robert Koch. There is a huge amount of immunological studies in the aspect of tuberculosis but there remain many unanswered questions. Neutrophils, cells of First line defence are being neglected in tuberculosis. Macrophages are considered as the key player in case of tuberculosis. Researches reveal that neutrophils play some interesting roles; it can be called as a bi-directional weapon. It plays instrumental role in killing mycobacterium, recruiting macrophages and also works hand in hand with macrophages in order to halt the spread of the organism. Neutrophils also activate innate immunity, secrete some substances like ectosomes, which favour in trapping the mycobacterial organisms. Whether neutrophils kills the mycobacterium or gets succumbed to it depends on the stage of infection. Neutrophils at times act like a suicidal bomber, by carrying the organisms to different organs and spreading the infections. In chronic cases they are also implicated in granuloma formation, the classic sign of TB.

Keywords: granuloma, *Mycobacterium tuberculosis*, neutrophil, trojan horse

### Introduction

*Mycobacterium tuberculosis* spares no one throughout the world and when it is unleashed it lashes more than 4000 people/day (WHO report 2010 – global tuberculosis control). Days are growing fast since the start of the research in Tuberculosis (TB) but the susceptibility to infection remains unanswered. The consequence of the TB ranges from early, asymptomatic clearance through latent infection to clinical disease [1]. Whenever there is talk about immunity in case of TB, macrophages are the first thing that comes to our mind and not neutrophils, though the latter is the first line of defence in the body.

### Neglected neutrophils

Neutrophils are the poorly ranked components of the host defence in case of TB. The reason behind this low profile chapter of neutrophils is because of the inherent difficulties in working with these cells. The key hurdles for neutrophils are:

- 1) they are short lived
- 2) easily activated
- 3) cannot be cryopreserved [1].

Hence *in-vitro* study is not feasible.

### Newer concepts of neutrophils in TB

- \* Neutrophils are the commonly affected phagocyte in human TB [2].
- \* Neutrophils significantly contribute to control of TB in human blood [3].
- \* Interferon in human whole blood which is neutrophil driven contributes to disease pathogenesis [4].

### Facts about neutrophils

Neutrophils kill organisms both by oxidative (phagocytic) and non oxidative (degranulation) [5]. There are 3 types of granules seen in neutrophils. They are:

- \* Azurophilic granules (or "primary granules") – contains proteins like myeloperoxidase, bactericidal/permeability-increasing protein (BPI), defensins, and the serine proteases neutrophil elastase.
- \* Specific granules (or "secondary granules") – contains lactoferrin and cathelicidin.
- \* Tertiary granules – contains cathepsin and gelatinase.

### Neutrophils in TB: two way traffic?

Neutrophils contribute to the early defence against mycobacteria. The scientific community is split, regarding the action of neutrophils in TB. LPS used to recruit neutrophils to the lungs of rat during airborne infection with 200 *M.tuberculosis*, decreases downstream colony forming units in lung [6]. Depleting granulocytes in mice before intra tracheal infection with  $10^5$  organisms, increases the number of colony forming units downstream in lung and spleen [7]. Science is not always constant and consistent. On the other side some works proved negative to the above mentioned action of neutrophils in TB. For example: no effect on bacterial load in mice with  $10^6$  colony forming unit of *M.tuberculosis*, *M.bovis*, BCG or *M. fortuitum* in case of granulocyte depletion [8]. Murine experiments with RB6-8C5 monoclonal antibodies to deplete granulocyte receptor positive cells, in addition to the target (Ly6G antigen) on neutrophils, it also targets Ly6C on plasmacytoid dendritic cells, mono-

cytes [9]. The stage of infection plays a crucial role: in established TB disease, higher neutrophil counts are associated with poor prognosis [10].

Begin with the basics

The preliminary steps taken by neutrophils after the entry of organism into the body are;

1. Recruitment
2. Recognition
3. Phagocytosis
4. Killing.

Neutrophil recruitment

Neutrophil recruitment occurs at the site mycobacterial infection. Recruitment occurs at the perivascular sites within an hour when intra-venous infection with *M.tuberculosis* [11]. There is infiltration of neutrophils in liver within 2 hours of systemic challenge with *M.avium* [12]. There is skin infiltration of neutrophils within 3 hours of BCG infection in rabbits [13]. In case of mice it takes 4 hours for skin infiltration with the same challenge study as that of rabbit [14].

Mechanism of recruitment: In sensitized animals there is a powerful immune response to mycobacterial challenge [11]. Interleukin 17 (IL 17) and IL 23 produced from T helper 17 (Th 17) cell are the key in recruitment of neutrophils [15]. IL 8 from macrophage also plays a key role [16]. In simpler terms we can say that the initial signal releases the inflammatory cytokine which activates local endothelium, coordinated with increase in adhesion molecules like Intra Cellular Adhesion Molecules (ICAM), E selectin, P selectin. These signals cause influx of neutrophils.

Phagocytosis of mycobacteria by neutrophils

Neutrophils engulf the entered mycobacteria [17]. The mechanisms which cause this interaction are:

1) Direct recognition 2) Opsonisation.

1) Direct recognition: Pattern Recognition Receptors makes this interaction possible [18]. Toll Like Receptor 2 (TLR2) also involved in this interaction. There is impaired control of *M.tuberculosis* and *M. avium* infection in TLR2 deficient mice [12]. Lipoarabinomannan or 19 kDa lipoprotein of mycobacteria is the ligand for TLR2. Blocking TLR4 reduces IL8 production, in response to infection [19]. Hence TLR4 may also be involved in this interaction between neutrophils and mycobacteria.

2) Opsonisation: Apart from direct recognition, opsonisation seems to play a key role in the phagocytosis of the mycobacteria by neutrophils. There is reduction in the ability of Ficoll isolated neutrophils to phagocytose mycobacteria after heat inactivation of serum [20].

Do neutrophils dissect mycobacteria?

This remains a controversial stuff. Theoretically speaking the neutrophils kill and stop the infection at an early stage. But in the absence of killing, the neutrophils traffics the infection to different organs of

the body. Thus playing the role of “granulocytic Trojan Horse” [21]. Mice treated with anti IL17 during *M.tuberculosis* infection shows 100 fold decrease in number of organism in spleen [22]. Treatment with IL17 causes reduction in neutrophil recruitment.

Mechanism of killing

A member of the  $\alpha$ -defensin family, Human Neutrophil peptides (HNP) plays a prime role in killing mycobacteria. They are cationic proteins seen in the azurophil granules which bind to anionic molecules [23]. Exceptions exist here too. *M.avium*, *M.kansasii*, *M.smegmatis* and *M.tuberculosis* fail to fuse to azurophil granules [24]. *lysX* gene of *M.tuberculosis* is same as *mprF* gene of *Staphylococcus aureus* that increase the lysine content of membrane lipids, hence negative charge is decreased and susceptibility to HNP also reduced [25].

Macrophages seem to take up free HNP and this increases their ability to kill mycobacteria [26]. Phagocytosis of apoptotic neutrophils by macrophages causes restriction of mycobacterial growth [27].

Neutrophil-macrophage co-operation

Clearing of cytotoxic, short lived neutrophils are carried out by macrophages. Macrophages are attracted by the chemokines derived from neutrophil [28]. Macrophage chemotaxis is also stimulated by mycobacterial lipoarabinomannan [29].

Process of apoptosis- pro inflammatory or anti inflammatory?

Controversy surrounds here too. Apoptosis is an anti inflammatory process resulting in induction of Tissue Growth Factor  $\beta$  (TGF  $\beta$ ), prostaglandin E2 (PGE2) and inhibits IL 6, IL 8 and Tissue Necrosis Factor (TNF) [30]. Sometimes the reverse may happen; ingestion of apoptotic cell with pathogen may result in pro inflammatory effect [30]. This may be due to expression of heat shock proteins [31] or activation of macrophages by neutrophil proteases.

Phagocytosis of apoptotic cell may produce anti/pro inflammatory effect based on the mycobacteria inside the neutrophil is alive or dead. If the organism is killed, there is anti-inflammatory effect. If the organism is living it causes pro-inflammatory effect.

Effect of neutrophils on acquired immunity

Neutrophils produce IL 12, macrophage inflammatory protein (MIP 1  $\alpha$ ) [32] which attract T lymphocytes and cause its maturity. Neutrophils can also produce IL 10 which might limit acquired immunity [33].

Neutrophil Extra cellular Traps (NETs)

Nuclear chromatin, mitochondrial DNA and granular anti microbial proteins form the NETs [34]. Pro-inflammatory stimuli like IL8, TNF forms NETs [35]. NETs can trap mycobacteria [36]. NETs formed against mycobacterial infection are unable to kill mycobacteria instead it kills *Listeria monocytogene*

which is found as a co-infection. Hence NETs can localize mycobacteria which may be the basis for granuloma formation.

Ectosomes (ECTs) from *M. tuberculosis* infected neutrophils

Vesicles released in response to pro-inflammatory stimuli from cell membrane are called as ectosomes [37]. ECTs are rich in cholesterol, express CD35 marker [38]. ECTs from neutrophil bind to endothelium selectively and macrophages but not to red blood cells, confirming its role in immune response.

#### Conclusion

Neutrophils are seen in the early stages of the mycobacterial infection. In chronic cases the same neutrophils may act in the pathology of granuloma formation. Thus neutrophil act as a “Double edged Sword”. The role of neutrophils in killing the mycobacteria is still doubtful. It may disseminate the organism to various organs by acting as a granulocytic trojan horse.

#### References

- Lowe, D.M et al. (2012). Neutrophils in tuberculosis: friend or foe? *Trends in immunology*. 33(1): 14-25.
- Eum, S.Y. et al. (2010). Neutrophils are the predominant infected phagocytic cells in the airways of patients with active pulmonary tuberculosis. *Chest* 137: 122–128.
- Martineau, A.R. et al. (2007). Neutrophil-mediated innate immune resistance to mycobacteria. *J. Clin. Invest.* 117: 1988–1994.
- Berry, M.P. et al. (2010). An interferon-inducible neutrophil-driven blood transcriptional signature in human tuberculosis. *Nature*. 466: 973–977.
- Kumar V & Sharma A. (2010). Neutrophils: Cinderella of innate immune system. *International Immunopharmacology*. 10(11): 1325-34, ISSN 1567-5769.
- Sugawara, I. et al. (2004). Rat neutrophils prevent the development of tuberculosis. *Infect. Immun.* 72: 1804–1806.
- Barrios-Payan, J. et al. (2006). Neutrophil participation in early control and immune activation during experimental pulmonary tuberculosis. *Gac. Med. Mex.* 142: 273–281.
- Seiler, P. et al. (2000). Rapid neutrophil response controls fast replicating intracellular bacteria but not slow-replicating *Mycobacterium tuberculosis*. *J. Infect. Dis.* 181: 671–680.
- Wojtasiak, M. et al. (2010). Gr-1+ cells, but not neutrophils, limit virus replication and lesion development following flank infection of mice with herpes simplex virus type-1. *Virology* 407: 143–151.
- Barnes, P.F. et al. (1988) Predictors of short-term prognosis in patients with pulmonary tuberculosis. *J. Infect. Dis.* 158: 366–371.
- Long, E.R. et al. (1931). Early cellular reaction to tubercle bacilli. *Arch. Pathol.* 12: 956–959.
- Feng, C.G. et al. (2003). Mice lacking myeloid differentiation factor 88 display profound defects in host resistance and immune responses to *Mycobacterium avium* infection not exhibited by Toll-like receptor 2 (TLR2)- and TLR4-deficient animals. *J. Immunol.* 171: 4758–4764.
- Shigenaga, T. et al. (2001). Immune responses in tuberculosis: antibodies and CD4-CD8 lymphocytes with vascular adhesion molecules and cytokines (chemokines) cause a rapid antigen-specific cell infiltration at sites of *Bacillus Calmette-Guérin* reinfection. *Immunology*. 102: 466–479.
- Abadie, V. et al. (2005). Neutrophils rapidly migrate via lymphatics after *Mycobacterium bovis* BCG intradermal vaccination and shuttle live bacilli to the draining lymph nodes. *Blood*. 106: 1843–1850.
- Cruz, A. et al. (2010). Pathological role of interleukin 17 in mice subjected to repeated BCG vaccination after infection with *Mycobacterium tuberculosis*. *J. Exp. Med.* 207: 1609–1616.
- Lyons, M.J. et al. (2002). *Mycobacterium bovis* BCG vaccination augments interleukin-8 mRNA expression and protein production in guinea pig alveolar macrophages infected with *Mycobacterium tuberculosis*. *Infect. Immun.* 70: 5471–5478.
- Wolf, A.J. et al. (2007). *Mycobacterium tuberculosis* infects dendritic cells with high frequency and impairs their function *in vivo*. *J. Immunol.* 179: 2509–2519.
- May, M.E. and Spagnuolo, P.J. (1987). Evidence for activation of a respiratory burst in the interaction of human neutrophils with *Mycobacterium tuberculosis*. *Infect. Immun.* 55: 2304–2307.
- Godaly, G. and Young, D.B. (2005). *Mycobacterium bovis* bacilli Calmette Guerin infection of human neutrophils induces CXCL8 secretion by MyD88-dependent TLR2 and TLR4 activation. *Cell Microbiol.* 7: 591–601.
- Majeed, M. et al. (1998). Roles of calcium and annexins in phagocytosis and elimination of an attenuated strain of *Mycobacterium tuberculosis* in human neutrophils. *Microb. Pathog.* 24: 309–320.
- Eruslanov, E.B. et al. (2005). Neutrophil responses to *Mycobacterium tuberculosis* infection in genetically susceptible and resistant mice. *Infect. Immun.* 73: 1744–1753.
- Redford, P.S. et al. (2010). Enhanced protection to *Mycobacterium tuberculosis* infection in IL-10-deficient mice is accompanied by early and enhanced Th1 responses in the lung. *Eur. J. Immunol.* 40: 2200–2210.
- Fu, L.M. (2003). The potential of human neutrophil peptides in tuberculosis therapy. *Int. J. Tuberc. Lung Dis.* 7, 1027–1032.
- Perskvist, N. et al. (2002). Rab5a GTPase regulates fusion between pathogen-containing phagosomes and cytoplasmic organelles in human neutrophils. *J. Cell Sci.* 115: 1321–1330.
- Maloney, E. et al. (2009). The two-domain LysX protein of *Mycobacterium tuberculosis* is required for production of lysinylated phosphatidylglycerol and resistance to cationic antimicrobial peptides. *PLoS Pathog.* 5: e1000534.
- Sharma, S. et al. (2000). Antibacterial activity of human neutrophil peptide-1 against *Mycobacterium tuberculosis* H37Rv: *in vitro* and *ex vivo* study. *Eur. Respir. J.* 16: 112–117.
- Tan, B.H. et al. (2006). Macrophages acquire neutrophil granules for antimicrobial activity against intracellular pathogens. *J. Immunol.* 177: 1864–1871.
- Mantovani, A. et al. (2011). Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat. Rev. Immunol.* 11: 519–531.
- Fietta, A. et al. (2000). *Mycobacterial* lipoarabinomannan affects human polymorphonuclear and mononuclear phagocyte functions differently. *Haematologica*. 85: 11–18.
- Krysko, D.V. et al. (2006). Clearance of apoptotic and necrotic cells and its immunological consequences. *Apoptosis*. 11: 1709–1726.
- Persson, Y.A. et al. (2008). *Mycobacterium tuberculosis*-induced apoptotic neutrophils trigger a pro-inflammatory response in macrophages through release of heat shock protein 72, acting in synergy with the bacteria. *Microbes Infect.* 10: 233–240.
- Seiler, P. et al. (2003). Early granuloma formation after aerosol *Mycobacterium tuberculosis* infection is regulated by neutrophils via CXCR3-signaling chemokines. *Eur. J. Immunol.* 33: 2676–2686.
- Dorhoi, A. et al. (2010). The adaptor molecule CARD9 is

- essential for tuberculosis control. *J. Exp. Med.* 207: 777–792.
34. Yousefi S, Mihalache C, Kozlowski E, Schmid I, Simon HU. (2009). Viable neutrophils release mitochondrial DNA to form neutrophil extracellular traps. *Cell Death and Differentiation*. 16(11): 1438–1444, ISSN 1476-5403.
35. Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss D, Weinrauch Y, Zychlinsky A. (2004). Neutrophil Extracellular Traps Kill bacteria. *Science*. 303(5663): 1532-1535.
36. Ramos-Kichik V, Mondragon-Flores R, Mondragon-Castelan M, Gonzalez-Pozos S, Muniz- Hernandez S, Rojas-Espinosa O, Chacón-Salinas R, Estrada-Parra S & Estrada-García I. (2009). Neutrophil extracellular traps are induced by *Mycobacterium tuberculosis*. *Tuberculosis (Edinb)*. 89 (1): 29–37.
37. Stein, J. M. & J. P. Luzio (1991) Ectocytosis caused by sublytic autologous complement attack on human neutrophils. The sorting of endogenous plasma-membrane proteins and lipids into shed vesicles. *Biochem J.* 274: 381-6.
38. Gasser, O., C. Hess, S. Miot, C. Deon, J. C. Sanchez & J. A. Schifferli (2003) Characterisation and properties of ectosomes released by human polymorphonuclear neutrophils. *Exp Cell Res.* 285: 243-57.

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