

The effects of protein dietary supplementation on fecal egg counts and hematological parameters in goat kids with subclinical nematodosis

Priyanka Konwar, S. P. Tiwari, M. Gohain and Kiran Kumari

Department of Animal Nutrition, College of Veterinary Science and Animal Husbandry,
Chhattisgarh Kamdhenu Vishwavidyalaya, Durg, Chhattisgarh, India.

Corresponding author: Priyanka Konwar, e-mail: priyankavet2009@gmail.com, SPT: drsptiwari@gmail.com,
MG: manujgohain02@gmail.com, KK: drkirankumari22@gmail.com

Received: 14-07-2015, **Revised:** 19-10-2015, **Accepted:** 23-10-2015, **Published online:** 24-11-2015

doi: 10.14202/vetworld.2015.1351-1355 **How to cite this article:** Konwar P, Tiwari SP, Gohain M, Kumari K (2015) The effects of protein dietary supplementation on fecal egg counts and hematological parameters in goat kids with subclinical nematodosis, *Veterinary World* 8(11): 1351-1355.

Abstract

Aim: The aim of the present study was to assess the effect of dietary supplementation with different levels of protein on fecal egg counts and hematological parameters in goat kids with subclinical nematodosis under semi-intensive condition.

Materials and Methods: 20 goat kids (3-5 months old with an average body weight of 8.90 kg) were randomly allocated to four groups: T1, served as a negative control, without receiving concentrate feed, and T2, T3, and T4 that received concentrate feed containing 16, 20, and 24% digestible crude protein, respectively. The experiment was carried out for 60 days.

Results: In this study, protein supplementation had a significant ($p < 0.05$) effect on fecal egg counts even after 15 days; hemoglobin (Hb) (g/dl) after 45 days; total leukocyte count ($10^3/\text{mm}^3$) and total erythrocyte count ($10^6/\text{mm}^3$) after 30 days; packed cell volume (%), lymphocyte (%), and eosinophil (%) after 15 days of supplementation, whereas monocyte (%) and neutrophil (%) values were not significantly influenced by protein supplementation effect during the entire experiment. The values of mean corpuscular volume (fl) were affected significantly ($p < 0.05$, $p < 0.01$) due to protein supplementation after 30 days, mean corpuscular Hb (MCH) (pg) after 45 days, but MCH concentration (g/dl) was not significantly different among the experimental groups during the entire experiment.

Conclusion: The dietary supplementation with different levels of protein significantly improved the hematological profiles and inhibited the nematodosis infection in the experimental goat kids.

Keywords: fecal egg count, goat kids, hematological parameter, nematodosis, protein dietary supplementation.

Introduction

Gastrointestinal parasitism is a major problem in a small ruminant production worldwide, due to its impact on animal health and productivity and the associated costs of control measures [1]. The usual strategy of gastrointestinal nematodes control based on the repeated use of anthelmintics and is nowadays under question because of the increasing development of resistance to these molecules [2].

The manipulation of host nutrition is the alternative method to anthelmintics and is intended to improve the host resistance and/or resilience against parasitic infections. Research has already shown that increased dietary intake of metabolizable protein and energy and combined with high-quality pasture can directly promote the host resistance and resilience against worm infection by maintaining tissue and/or blood homeostasis and production [3,4]. An improvement of the host diet contributes to maintain the tissue and/or blood homeostasis and the host production despite the presence of worms.

The present study was, therefore, carried out to study the effect of protein dietary supplementation on

Egg per gram (EPG of feces) and different hematological parameters in goat kids infected with natural subclinical nematodosis.

Materials and Methods

Ethical approval

Permission of the Institutional Animal Ethics Committee was taken prior to the start of the experiment.

Experimental animal, diet, design, and management

20 indigenous goat kids of 3-5 months old infected by natural subclinical nematodosis were selected for the experiment conducted in the Department of Animal Nutrition, College of Veterinary Science and Animal Husbandry, Anjora, Durg (Chhattisgarh) and randomly allocated to four groups of five animals each (T1 served as a negative control, without receiving concentrate feed, and T2, T3, and T4 that received concentrate feed containing 16%, 20%, and 24% digestible crude protein [DCP], respectively). They were ear tagged for identification and experiment was carried out for 60 days with the completely randomized design under semi-intensive condition. Animals were individually fed and were reared under same conditions of hygiene and management.

The animals grazed in a pasture containing mixed grass predominant with sola grass (*Aeschynomene indica*) for 4 h (8 am to 12 noon) and concentrate feed

Copyright: The authors. This article is an open access article licensed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>) which permits unrestricted use, distribution and reproduction in any medium, provided the work is properly cited.

(100 g/day/animal) was offered at 3:30 pm. Fecal egg count was determined to confirm the occurrence of nematodosis. Diets were formed by using crushed maize, soybean meal, de-oiled rice bran; in detail, T2, T3, and T4 diets contained 70% of total digestible nutrients and 16%, 20% and 24% of DCP, respectively. The diets were further supplemented with mineral mixture (2%) and salt (1%). The diets were formulated as per standard requirement [5].

Blood and fecal sample collection

The fresh fecal samples from all kids were individually collected using hand gloves at 0, 15, 30, 45, and 60 days. The samples were put in the plastic bags for egg counting as described by the modified McMaster techniques [6]. Blood samples were also collected from all kids at 0, 15, 30, 45 and 60 days. In detail, 2 ml blood was taken in a glass vial containing ethylene diamine tetra acetate appropriate for hematological analyses. Immediately after blood collection, the tubes were gently rotated between palms to mix it with anticoagulant. Hemoglobin (Hb), packed cell volume (PCV), total erythrocyte count (TEC), total lymphocyte count (TLC), differential leukocyte count - monocyte, lymphocyte, neutrophil and eosinophil, mean corpuscular volume (MCV), mean corpuscular Hb (MCH), and MCH concentration (MCHC) were performed as described by Jain [7].

Statistical analysis

The results obtained during this study were analyzed by as per Snedecor and Cochran [8] using software package SPSS version 16.0 [9].

Results and Discussion

EPG

The mean values of EPG (Table-1) significantly differ ($p < 0.05$) among the treatments. The highest values were obtained in T1 and lowest in T3 and T4, even from the first 15 days of protein dietary supplementation. The values were increased throughout the experiment possibly due to flock grazing. Especially, T3 have significantly ($p < 0.01$) lower values than T1 and T2 groups suggesting that the supplementation influenced the rate of larval development in the kids. It might also be an effect of resistance and/or resilience increase against nematodes due to higher protein supplementation level. Previous studies have also revealed that there is a significant reduction in the fecal egg count and

worm burden in goats after supplementation with dietary protein [10,11].

Percentile depiction of nematode genera

From the pooled coproculture, the infective larvae were harvested, and it was observed that predominantly *Haemonchus* sp. were present in all the treatments at a ratio of 9:1.

Hematological profiles

The Hb concentration significantly differed ($p < 0.01$) amongst the different groups at 45th and 60th day of the experiment (Table-2). The concentration at the 45th day was higher in protein supplemented groups and at the 60th day, values were the highest in T3 and T4. The concentration of Hb decreased as the levels of infection and incubation period increased. However as the level of protein supplementation increased, the Hb content also increased after the 30th day of the experiment, especially after the 45th day. In the present study, a reduction in Hb content was observed, possibly as a result of either blood loss due to blood-sucking by larvae and adults [12] or due to mucosa damage [13]. Al-Rekani [14] revealed that there is significant reduction in Hb in infected goats compared to control. With increase in parasitemia rates, Hb concentration significantly decreased in small ruminants naturally infected with parasites [15]. Parasitic anemia is related with low Hb levels, eosinophilia and a moderate lymphopenia in naturally infected sheep [16]. Abdel Hameed *et al.* [17] reported higher values of Hb in diets with a higher level of protein supplementation in Sudan desert lambs compared to other experimental diets.

The mean TLC values significantly differed among the experimental groups on day 30 ($p < 0.05$) and on days 45 and 60 ($p < 0.01$) of the experiment; in general, T3 and T4 had higher values compared to T1 and T2 group (Table-2). TLC value increased as the levels of infection and incubation period increased, possibly as a result of parasitic infection. Qamar [18] reported that TLC values were significantly affected by the extent of nematode parasitosis. In the case of parasitized animals that are supplemented with proteins, the number of eosinophils and other leukocytes, mast cells and proteases released from the mast cells increased in the mucosa of the gastrointestinal tract [19]. TLC showed a significantly increased value in small ruminants naturally infected with parasites [15].

Table-1: The effect of protein supplementation on fecal egg count in goats.

Parameters	Period (day)	Treatments				Significance
		T1	T2	T3	T4	
EPG	0	1520±73.48	1480±73.48	1520±37.42	1500±70.71	NS
	15	1860±50.99 ^b	1720±58.31 ^{ab}	1680±37.42 ^a	1620±73.48 ^a	*
	30	2240±67.82 ^b	1920±58.31 ^a	1820±37.42 ^a	1840±81.24 ^a	**
	45	2580±48.99 ^b	2080±58.31 ^a	1920±37.42 ^a	2020±96.95 ^a	**
	60	2920±58.31 ^c	2320±58.31 ^b	2040±50.99 ^a	2220±86.02 ^{ab}	**

^{abc}Means within a row with different superscripts are significantly different (* $p < 0.05$, ** $p < 0.01$). NS=Not significant

The mean values of TEC significantly differed ($p<0.05$, $p<0.01$) among the experimental groups during the experiment apart from the day 0 to 15 (Table-2). Lower values were shown in T1 than the other treatments. Decreased level of TEC as a result of increase in parasitemia were reported in small ruminants with natural parasitic infestation [14,15]. Abdel Hameed *et al.* [17] reported higher TEC values in diets supplemented with different level of protein in Sudan desert lambs compared to other experimental diets.

The mean PCV values significantly differed ($p<0.01$) among the experimental groups during the entire period apart from day 0 (Table-2). On day 30, 45, and 60, T3 showed the highest and T1 the lowest values compared to T2 and T4 group, possibly as a result of the improved nutrition of the parasitized animals. The higher dietary protein improved the erythropoiesis or reduced the establishment or development of adult nematodes. The kids on the higher protein diet even with heavy infection better able to resist the adverse effect of gastrointestinal nematodosis. Al-Rekani [14], Esmailnejad [15] also reported decreased values of PCV in infected compared to control animals.

The influence of protein supplementation level on monocyte and neutrophil values are also presented in Table-3. No significant differences were found among the treatments. The values of monocytes did not follow any specific trend and in the case of neutrophil; there was a decreasing but not significant trend in T2, T3, and T4. Esmailnejad *et al.* [15] reported that neutrophil counts were significantly increased with the increase of parasitemia rates in natural parasitic small ruminants, a finding that is in accordance with the present study.

The influence of protein supplementation on lymphocyte count in kids was significant ($p<0.01$)

from the 15th day (Table-3). The highest values were constantly showed by T1 and lowest by T3 group. The results of the present study might be due to the chronic infection of nematodosis resulting in the exhausted immune system. Esmailnejad *et al.* [15] reported that number of lymphocyte increased significantly in natural parasitic small ruminants which were in accordance with the present findings.

As presented in Table-3, eosinophil count was significantly influenced by protein supplementation apart from day 0. The eosinophil count was higher in T1 followed by T2, T4 and T3. It has already been suggested that eosinophil count may provide an index of protective immune response of animals to parasitism [20]. With increase in parasitemia rates, eosinophil count shows a significantly increased value in small ruminants with natural parasitic infection [15].

The average values of MCV, MCH, and MCHC of the experimental groups are presented in Table-4. The MCV values were higher in T3 and T4 compared to other groups on days 30, 45 and 60. The MCH values were higher in T3 and T4 compared to T1 group on days 45 and 60. No significant differences in MCHC values were found among the groups. However, the values were decreased as the time period and level of infection increased. The present findings of MCV and MCH were supported by previous studies in goats infected with parasites [15]. However, in contrast with the present study, MCV appeared to slightly increase with increase in worm load in cattle affected by fascioliasis [21]. Abdel Hameed *et al.* [17] also reported higher values in diets with different level of protein supplementation in Sudan desert lambs compared to other experimental diets which were in accordance with present findings. MCHC showed no reasonable

Table-2: The effect of protein supplementation on hematological parameters in goats.

Parameters	Period (days)	Treatments				Significance
		T1	T2	T3	T4	
Hb (g/dl)	0	12.58±0.63	11.28±0.31	11.32±0.27	11.44±0.27	NS
	15	10.56±0.70	10.38±0.33	10.46±0.20	10.64±0.27	NS
	30	8.82±0.49	9.36±0.26	9.80±0.19	9.88±0.26	NS
	45	7.08±0.39 ^a	8.52±0.38 ^b	9.24±0.17 ^b	9.10±0.23 ^b	**
	60	5.72±0.25 ^a	7.18±0.27 ^b	8.66±0.16 ^c	8.30±0.26 ^c	**
TLC (10 ³ /mm ³)	0	7.14±0.11	7.17±0.16	7.18±0.11	7.27±0.16	NS
	15	7.25±0.12	7.37±0.17	7.56±0.12	7.59±0.17	NS
	30	7.39±0.09 ^a	7.61±0.17 ^{ab}	7.98±0.12 ^b	7.92±0.16 ^b	*
	45	7.51±0.07 ^a	7.83±0.18 ^{ab}	8.39±0.12 ^c	8.22±0.16 ^{bc}	**
	60	7.65±0.06 ^a	8.03±0.19 ^a	8.80±0.12 ^b	8.51±0.15 ^b	**
TEC (10 ⁶ /mm ³)	0	12.34±0.14	12.48±0.18	12.36±0.15	12.42±0.19	NS
	15	11.71±0.15	12.04±0.18	12.09±0.14	12.06±0.20	NS
	30	11.04±0.16 ^a	11.58±0.18 ^b	11.74±0.19 ^b	11.66±0.19 ^b	*
	45	10.35±0.15 ^a	11.08±0.17 ^b	11.49±0.12 ^b	11.26±0.20 ^b	**
	60	9.65±0.16 ^a	10.56±0.19 ^b	11.19±0.14 ^c	10.85±0.20 ^{bc}	**
PCV (%)	0	33.26±0.35	32.74±0.30	32.92±0.40	32.66±0.38	NS
	15	29.38±0.66 ^a	29.86±0.21 ^a	31.90±0.35 ^b	30.42±0.47 ^{ab}	**
	30	25.52±0.71 ^a	26.68±0.26 ^{ab}	30.58±0.35 ^c	28.40±0.44 ^b	**
	45	21.48±0.74 ^a	23.58±0.23 ^b	29.04±0.31 ^d	26.70±0.44 ^c	**
	60	17.88±0.76 ^a	20.02±0.37 ^b	27.52±0.21 ^d	24.26±0.46 ^c	**

^{abc}Means within a row with different superscripts are significantly different (* $p<0.05$, ** $p<0.01$). Hb=Hemoglobin, PCV=Packed cell volume, TEC=Total erythrocyte count, TLC=Total leukocyte count, NS=Not significant

Table-3: The effect of protein supplementation on DLCs in goats.

Parameters	Period (days)	Treatments				Significance
		T1	T2	T3	T4	
Monocytes (%)	0	1.60±0.75	1.80±0.80	1.40±0.98	1.40±0.98	NS
	15	1.40±0.60	1.40±0.98	1.40±0.87	1.20±0.97	NS
	30	1.60±0.81	1.20±0.97	1.40±1.40	1.40±0.98	NS
	45	1.20±0.73	1.20±1.20	1.60±1.03	1.60±1.17	NS
	60	2.00±0.89	1.20±0.97	1.40±1.40	1.40±1.17	NS
Lymphocytes (%)	0	51.60±0.93	51.40±0.93	51.60±1.03	51.60±0.75	NS
	15	50.80±0.80 ^d	48.00±0.55 ^c	44.40±0.60 ^a	46.20±0.37 ^b	**
	30	48.80±0.37 ^d	46.20±0.37 ^c	42.00±0.45 ^a	44.20±0.37 ^b	**
	45	45.80±0.58 ^d	44.20±0.37 ^c	40.20±0.37 ^a	42.00±0.32 ^b	**
	60	43.20±0.58 ^c	42.20±0.37 ^c	38.20±0.37 ^a	39.80±0.37 ^b	**
Neutrophils (%)	0	41.80±0.80	42.00±0.55	41.80±0.80	41.60±0.75	NS
	15	42.00±0.84	41.00±0.89	41.00±1.05	41.00±0.89	NS
	30	42.00±0.84	40.00±0.71	40.40±0.75	40.20±0.66	NS
	45	42.20±1.02	39.00±0.89	39.80±1.07	39.40±0.93	NS
	60	42.20±1.02	38.40±0.93	39.40±0.93	38.60±0.81	NS
Eosinophils (%)	0	5.20±0.37	5.40±0.24	5.00±0.32	5.20±0.20	NS
	15	9.20±0.37 ^b	8.40±0.24 ^b	7.00±0.32 ^a	7.40±0.24 ^a	**
	30	13.20±0.37 ^d	11.40±0.24 ^c	8.20±0.37 ^a	9.40±0.24 ^b	**
	45	16.20±0.37 ^d	13.80±0.49 ^c	9.40±0.24 ^a	11.40±0.24 ^b	**
	60	19.20±0.37 ^d	16.40±0.24 ^c	10.80±0.20 ^a	13.60±0.24 ^b	**

^{abc}Means within a row with different superscripts are significantly different (**p<0.01). NS=Not significant, DLC=Differential leukocyte count

Table-4: The effect of protein supplementation on red blood cell indices in goats.

Parameters	Period (days)	Treatments				Significance
		T1	T2	T3	T4	
MCV (fl)	0	26.96±0.29	26.24±0.28	26.64±0.55	26.31±0.57	NS
	15	25.08±0.53	24.81±0.29	26.38±0.47	25.25±0.68	NS
	30	23.12±0.59 ^a	23.04±0.27 ^a	26.08±0.69 ^b	24.39±0.64 ^{ab}	**
	45	20.76±0.64 ^a	21.29±0.43 ^a	25.28±0.47 ^b	23.72±0.61 ^b	**
	60	18.51±0.67 ^a	18.98±0.56 ^a	24.61±0.41 ^c	22.38±0.62 ^b	**
MCH (pg)	0	10.21±0.57	9.05±0.30	9.17±0.32	9.23±0.27	NS
	15	9.03±0.63	8.63±0.31	8.65±0.23	8.83±0.28	NS
	30	8.00±0.46	8.09±0.25	8.35±0.24	8.48±0.28	NS
	45	6.85±0.39 ^a	7.70±0.38 ^{ab}	8.04±0.20 ^b	8.09±0.27 ^b	*
	60	5.94±0.35 ^a	6.81±0.32 ^{ab}	7.75±0.21 ^b	7.66±0.29 ^b	**
MCHC (g/dl)	0	37.83±1.89	34.47±1.04	34.40±0.86	35.03±0.78	NS
	15	35.93±2.16	34.76±1.07	32.80±0.65	34.98±0.70	NS
	30	34.55±1.58	35.09±1.01	32.06±0.75	34.79±0.84	NS
	45	33.00±1.60	36.09±1.33	31.84±0.75	34.12±1.04	NS
	60	32.30±2.34	35.82±0.75	31.48±0.76	34.27±1.33	NS

^{abc}Means within a row with different superscripts are significantly different (*p<0.05, **p<0.01). NS=Not significant, MCV=Mean corpuscular volume, MCH=Mean corpuscular hemoglobin, MCHC=Mean cell hemoglobin concentration

change with increase in worm load in cattle affected with fascioliasis [21]. Reduced level of MCHC with the increase in infection was also observed in small ruminants with natural parasitic infection [15]. At the same time, higher values of MCHC were recorded in diets with different level of protein supplementation in Sudan desert lambs compared to other experimental diets [17].

Conclusion

It may be concluded that dietary supplementation with different levels of protein in goats with subclinical nematodosis had significant beneficial effects on reducing the worm egg count and on improving hematological profile with respect to Hb, TLC, TEC, PCV, lymphocyte, eosinophil, MCV, and

MCH. Thus, protein-rich concentrate feed may be used as part of a sustainable strategy to control gastrointestinal nematodosis in goat kids under semi-intensive conditions.

Authors' Contributions

PK carried out the experiment. SPT - Designed and guided the experiment. MG and KK help in sample processing and data analysis. All authors read and approved the final manuscript.

Acknowledgments

The authors are thankful to the Dean, College of Veterinary Science and Animal Husbandry, Anjora, Durg for providing necessary fund and facilities required to accomplish the experiment.

Competing Interests

The authors declare that they have no competing interests.

References

1. Knox, M.R., Torres-Acosta, J.F.J. and Aguilar-Caballero, A.J., (2006) Exploiting the effect of dietary supplementation of small ruminants on resilience and resistance against gastrointestinal nematodes. *Vet. Parasitol.*, 139: 385-393.
2. Sanyal, P.K. (1998) Integrated parasite management in ruminants in India. A concept note. Biological control of gastrointestinal parasites in ruminants using predacious fungi. FAO Animal Production & Health Paper 141. FAO, Rome. p54-65.
3. Knox, M.R. (2002) Effectiveness of copper oxide wire particles for *Haemonchus contortus* control in sheep. *Aust. Vet. J.*, 80: 224-227.
4. Pathak, A.K. and Tiwari, S.P. (2013) Effect of high plane of nutrition on the performance of *Haemonchus contortus* infected kids. *Vet. World*, 6(1): 22-26.
5. National Research Council. (1981) Nutrient Requirements of Domestic Animals; Nutrient Requirement of Goats. No. 15 National Academy Press, Washington, DC.
6. Skerman, K.D. and Hillard, J.J. (1966) A Handbook of Studies of Helminth Parasites of Ruminants. Near East Animal Health Institute, Iran, FAO, Rome.
7. Jain, N.C. (1986) Haematological techniques. In: Schalm's Veterinary Haematology. 4th ed. Lea and Febinger, Philadelphia, PA. p20-86.
8. Snedecor, G.W. and Cochran, W.B. (1994) Statistical Methods. 8th ed. Iowa State University Press, Ames, Iowa.
9. SPSS. (2010) Statistical Packages for Social Sciences. Version 16, SPSS Inc., Illinois, USA.
10. Mhomga, L.I., Nnadi, P.A., Chiejina, S.N., Idika, I.K. and Ngongeh, L.A. (2012) Effect of protein supplementation on weight gain and dressing percentage of West African Dwarf goats experimentally infected with *Haemonchus contortus* and *Trichostrongylus colubriformis*. *Glob. Adv. Res. J. Agric. Sci.*, 1: 279-287.
11. Houdijk, J.G.M. (2012) Differential effects of protein and energy scarcity on resistance to nematode parasites. *Small Rumin. Res.*, 103: 41-49.
12. Rowe, J.B., Nolan, J.V., Dechaneet, G. and Telani, F. (1988) The effect of haemonchosis and blood loss into the abomasums on digestion in sheep. *Br. J. Nutr.*, 59: 125-139.
13. Abbott, E.M., Parkins, J.J. and Holmes, P.H. (1988) Influence of dietary protein on the pathophysiology of haemonchosis in lambs given continuous infections. *Res. Vet. Sci.*, 45: 41-49.
14. Al-Rekani, A.M.A. (2012) Effect of natural infection with gastrointestinal nematode on milk composition and blood parameters of lactating native goats. *Int. J. Sci. Res.*, 1(2): 14-17.
15. Esmailnejad, B., Tavassoli, M. and Asri-Rezaei, S. (2012) Investigation of hematological and biochemical parameters in small ruminants naturally infected with *Babesia ovis*. *Vet. Res. Forum*, 3: 31-36.
16. Yuksek, N., Altug, N., Denizhan, V., Ceylan, E. and Agaoglu, Z. (2013) Enhancement of the glucose metabolism and the reverse cholesterol transport by a peroxisome proliferator receptor α (PPAR α) agonist included in the fasciolosis treatment in naturally infested sheep. *Rev. Med. Vet. Toulouse*, 164: 163-172.
17. Abdel Hameed, A.A., Salih, A.M., Fadel Elseed, A.M. and Amasab, E.O. (2013) Effect of feeding untreated or urea treated groundnut hull supplemented with different protein sources on blood parameters of Sudan desert lambs. *Online J. Anim. Feed Res.*, 3: 40-46.
18. Qamar, M.F. (2009) Epidemiology, serodiagnosis, economic losses and control of haemonchosis in sheep and goat. Ph.D. Thesis. Department of Parasitology. University of Veterinary and Animal Sciences Lahore, Pakistan.
19. Basabe, J., Eiras, D.F. and Romero, J.R. (2009) Nutrition and gastrointestinal parasitism in ruminant production. *Arch. Zootec.*, 58: 131-144.
20. Finkelman, F.D., Pearce, E.J., Urban, J.F. and Sher, A. (1991) Regulation and biological function of helminth-induced cytokine responses. *Immunol. Today*, 12: A62-A66.
21. Egbu, F.M.I., Patience, O.U. and Ikem, C.O. (2013) Haematological changes due to bovine fascioliasis. *Afr. J. Biotechnol.*, 12: 1828-1835.
