Leptospirosis in horses: Sentinels for a neglected zoonosis? A systematic review

Eduardo A. Díaz¹, Gabriela Arroyo¹, Carolina Sáenz², Luis Mena³, and Verónica Barragán⁴

1. Escuela de Medicina Veterinaria, Colegio de Ciencias de la Salud, Universidad San Francisco de Quito, Diego Robles, Quito, 170157, Ecuador; 2. Hospital de Fauna Silvestre Tueri, Instituto iBIOTROP, Universidad San Francisco de Quito, Diego Robles, Quito, 170157, Ecuador; 3. Carrera de Medicina Veterinaria, Facultad de Ciencias Pecuarias, Escuela Superior Politécnica de Chimborazo ESPOCH, Riobamba, 060155, Ecuador; 4. Colegio de Ciencias Biológicas y Ambientales, Instituto de Microbiología, Universidad San Francisco de Quito, Diego Robles, Quito, 170157, Ecuador: **Corresponding author:** Eduardo A. Díaz, e-mail: eadiaz@usfq.edu.ec

Co-authors: GA: garroyo@usfq.edu.ec, CS: csaenz@usfq.edu.ec, LM: luismenamino@hotmail.com, VB: vbarragan@usfq.edu.ec

Received: 29-06-2023, Accepted: 14-09-2023, Published online: 14-10-2023

doi: www.doi.org/10.14202/vetworld.2023.2110-2119 **How to cite this article:** Díaz EA, Arroyo G, Sáenz C, Mena L, and Barragán V (2023) Leptospirosis in horses: Sentinels for a neglected zoonosis? A systematic review, *Veterinary World*, 16(10): 2110–2119.

Abstract

Background and Aim: Leptospirosis is considered a neglected tropical zoonosis in low-income countries due to surveillance system limitations and non-specificity of symptoms. Humans become infected through direct contact with carrier animals or indirectly through Leptospira-contaminated environments. Conventionally, equines have been considered an uncommon source of leptospirosis, but recent publications in Latin America suggest that their role in the maintenance and dispersion of the bacteria could be more relevant than expected, as horses are susceptible to a wide variety of zoonotic Leptospira spp. from domestic and wild animals with which they share the environment. A systematic review of the published literature was conducted to compile the available information on Leptospira spp. in Ecuador, with a special focus on equine leptospirosis, to better understand the epidemiology of the bacterium and identify possible knowledge gaps.

Materials and Methods: A systematic review of the published literature was conducted in PubMed, SciELO and Web of Science databases to compile the available information on Leptospira spp. in Ecuador, with a special focus on equine leptospirosis, to better understand the epidemiology of the bacterium. We used a combination of the terms (Leptospira OR Leptospirosis) AND Ecuador, without restrictions on language or publication date.

Results: Our literature review reveals that published scientific information is very scarce. Eighteen full-text original scientific articles related to Leptospira or leptospirosis cases in Ecuador were included in the systematic review. Most of the studies reported data obtained from one of the four regions (Coast), and specifically from only one of the 24 Provinces of Ecuador (Manabí), which evidence a large information bias at the geographical level. Furthermore, only the studies focused on humans included clinical signs of leptospirosis and there is only one study that analyzes the presence of Leptospira spp. in water or soil as a risk factor for pathogen transmission. Finally, only one study investigated Leptospira in horses.

Conclusion: Since sentinel species can provide useful data on infectious diseases when epidemiologic al information is lacking, and horses could be considered excellent sentinel species to reveal circulating serovars, we propose developing a nationwide surveillance system using horses. This cost-effective epidemiological survey method provides a baseline for implementing specific prevention and control programs in Ecuador and neighboring developing countries.

Keywords: Ecuador, equine leptospirosis, neglected tropical zoonosis, one-health, sentinel species, systematic review.

Introduction

Leptospirosis is a re-emerging zoonosis of public health concern due to morbidity and mortality in both humans and animals [1]. Humans are commonly infected through direct contact with the urine of *Leptospira* spp. carrier animals or indirectly by contaminated environments with this pathogen [2]. The disease is predominantly distributed among low-income populations in tropical developing countries [3]. It is estimated to cause more than 1 million severe human cases and approximately 60,000 deaths/year. However, the limitations of surveillance systems in impoverished regions probably contribute to an underestimation of the real impact of leptospirosis [4]. Furthermore, leptospirosis is usually underdiagnosed because of the difficulty in distinguishing its clinical signs from those of other endemic febrile diseases [5]. An overview of risk factors for leptospirosis suggests that epidemiological patterns are closely related to the bioclimatic context with heavy seasonal rains in tropical countries being one of the main risk factors [6]. Climate change and extreme weather events such as cyclones and floods are expected to occur with greater frequency and intensity, which could lead to increased leptospirosis outbreaks [7].

Latin America has one of the highest estimated incidences of leptospirosis in the world, including regions

Copyright: Díaz, *et al.* Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/ publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

where the burden of the disease is underappreciated [4, 8]. Usually, health authorities report cases that occur after floods in urban areas, but the cases in rural areas often go unreported [9]. In Ecuador, extreme weather conditions, such as "El Niño", and socio-economic factors, such as poor sewage infrastructure and inadequate hygienic conditions in rural areas, contribute to the fact that leptospirosis continues to be a neglected growing problem [10, 11]. Indeed, Chiriboga et al. [12] detected Leptospira-DNA in a high percentage of febrile patients from rural communities who had been tested for several endemic diseases, but not for leptospirosis. Barragan et al. [13] also found Leptospira-DNA in febrile people, cattle, and pigs from rural communities, suggesting that these animals may be the most important reservoir for human transmission. Nevertheless, recent bibliographic reviews have shown that equine leptospirosis may be more common than expected in Latin America [14, 15]. Leptospirosis in horses has traditionally been considered relatively uncommon, but new data suggest that the infection is widespread, with an incidence and infecting serovars varying considerably between different geographical regions [16]. Management factors such as the presence of other animal species, increasing age and herd size, free-ranging feeding practices, drinking untreated water, flooding, and poor sanitation are associated with increased risk of exposure to Leptospira spp. in horses [17–22]. Clinical signs include reproductive disorders, renal and hepatic dysfunctions, respiratory distress, and recurrent uveitis, but most infections remain asymptomatic [23]. Several worldwide studies have shown that, although the prevalence of infection may be higher than for other species, unrecognized subclinical infections commonly occur in apparently healthy horses [18, 24-28]. However, infected horses can become carriers and contribute to maintaining the bacterium in the environment by shedding Leptospira spp. in their urine [27, 29]. Seroprevalence and isolation studies indicate that the horse is susceptible to various incidental infections from different serovars [23, 30, 31]. Accordingly, since humans and other animals exposed to the bacteria share the environment with horses, these could be considered excellent sentinel species to reveal circulating serovars [20, 32, 33]. Serovars present in tropical regions with high wildlife richness are generally related to the wide range of mammalian reservoirs because wild mammals are more likely to be infected with *Leptospira* spp. [34, 35]. Ecuador is one of the most megadiverse countries on the planet, with hundreds of registered mammal species [36, 37]. This biodiversity, added to the climatic and socio-economic conditions of the region, makes it an ideal place to study the complex epidemiology of *Leptospira* spp. [13].

Equine population has increased considerably in recent years in Ecuador, both in peri-urban and rural areas for recreational activities and in more remote areas where they continue to be essential for agricultural labors [38]. Since the use of sentinel species may be a cost-effective study target to improve epidemiological surveillance in developing countries [39, 40], and horses have proven to be a suitable species for revealing circulating leptospiral serovars [20, 32, 33], we propose their use as sentinel species in the region.

A systematic review of the published literature was conducted to compile the available information on *Leptospira* spp. in Ecuador, with a special focus on equine leptospirosis, to better understand the epidemiology of the bacterium and identify possible knowledge gaps. Our results will improve prevention and control programs for both humans and animals.

Materials and Methods

Ethical approval

This study does not require ethical approval. Following Preferred Reporting Items for Systematic Reviews and Meta-analyses guidelines, we conducted a systematic review to collect current knowledge on *Leptospira* in Ecuador.

Study period and location

This study was conducted from January to May 2023 at the Universidad San Francisco de Quito, Diego Robles, Quito, 170157, Ecuador.

Search strategy

Published literature was searched in PubMed (http://www.ncbi.nlm.nih.gov/pubmed), SciELO (https://scielo.org) and Web of Science (http://apps. webofknowledge.com) databases on April 20, 2023, without restrictions on language or publication date. We used a combination of the terms (Leptospira OR Leptospirosis) AND Ecuador.

Article selection process

After retrieving the results from the databases, duplicate articles were identified and discarded by sorting the titles alphabetically using an Excel 2010 spreadsheet. At least two researchers read each article to confirm its relevance to the scope of the review. If a disagreement was observed, a third researcher was involved to further include or exclude the article. Article triage was performed in three stages, involving review of article's title, abstract, and full-text in accordance with inclusion and exclusion criteria.

Inclusion and exclusion criteria

Eligibility criteria included full-text original scientific articles related to *Leptospira* or leptospirosis in Ecuador; abstracts, letters to the editor, and articles without original data or non-scientific communications were excluded from the screening process.

Data extraction and synthesis

The following information was extracted from each study that met the inclusion criteria: (a) Article reference, (b) study area, (c) study species, (d) type sample, (e) positivity, (f) clinical signs, (g) *Leptospira* species, and (h) *Leptospira* serovars.

Results

A total of 72 articles were retrieved according to the search terms (6 from SciELO, and 31 from Web of

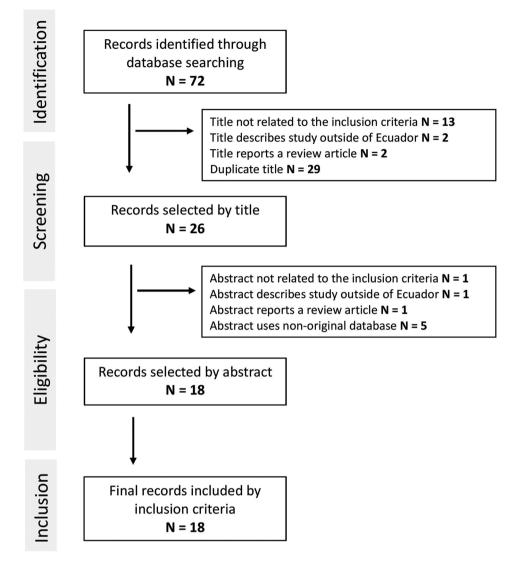


Figure-1: Flow diagram of the study selection process and exclusion identified.

Science, and 35 articles from PubMed). After removing 29 duplicate articles, 43 articles were screened by title. Of these, 13 were excluded because they were not related to the inclusion criteria, two described studies outside of Ecuador, and two reported review articles. By screening the remaining 26 abstracts, one was excluded as it was not related to the inclusion criteria, one described a study outside of Ecuador, one reported a review article, and five used a non-original database. Finally, 18 full-text original scientific articles related to *Leptospira* or leptospirosis cases in Ecuador were included in the systematic review. The flow diagram of the search strategy is summarized in Figure-1.

The number of published studies investigating *Leptospira* or leptospirosis in Ecuador increased over the years and more significantly in the last decade (before 2002, n = 0; between 2002 and 2011, n = 3; between 2012 and 2021, n = 15). Articles reported data obtained from three geographical regions (Coast n = 13; Amazon n = 2; Andes n = 1; and Galapagos n = 1) and six provinces (Manabí n = 10; Guayas

n = 2; Esmeraldas n = 1; Galapagos n = 1; Morona Santiago n = 1; Pastaza n = 1; and Pichincha n = 1); one study did not specify the geographical region or province. Included studies were mostly focused on domestic animals (n = 7) followed by those involving humans (n = 5), while fewer studies were centered on wildlife (n = 1) and *Leptospira* spp. in the environment (n = 1). Regarding the interrelationship between humans, domestic animals, wildlife, and the environment, the search retrieved studies that included information on humans, domestic and synanthropic species (n = 2), humans and domestic species (n = 1), and domestic and wild species (n = 1); no research provided joint data on all of them. The geographical location and studied populations are summarized in Figure-2.

In total, 1638 people and at least 15 different animal species were tested, including seven domestic species: 2192 cows (*Bos taurus*), 639 pigs (*Sus domesticus*), 34 dogs (*Canis familiaris*), three European rabbits (*Oryctolagus cuniculus*), three horses (*Equus caballus*), three sheep (*Ovis aries*), and one Guinea

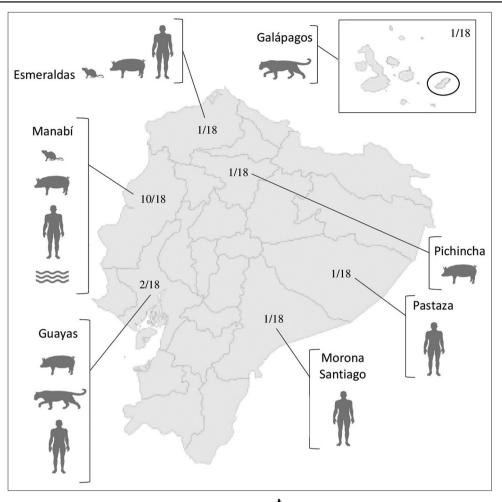


Figure-2: Provinces with published on *Leprosaria* in humans (n), synanthropic (**N**), domestic (**T**), wild (**T**) species and/or the environment (**S**). The first number shows the research from each province, the second number is the total of research identified in Ecuador (one study did not specify location). NO research was found in Azuay, Bolivar, Cañar, Carchi, Chimborazo, Cotopaxi, El Oro, Imbabura, Loja, Los Rios, Napo, Orellana, Santa Domingo, Sucumbíos, Tungurahua, and Zamora.

pig (*Cavia porcellus*); seven wild species: Seven Galapagos sea lions (*Zalophus wollebaeki*), three lions (*Panthera leo*), two ring-tailed coatis (*Nasua nasua*), one common woolly monkey (*Lagothrix lagotricha*), one Ecuadorian white-fronted capuchin (*Cebus aequatorialis*), one mountain coati (*Nasuella olivacea*), and one oncilla (*Leopardus tigrinus*); and one synanthropic genus: 107 rats (*Rattus spp.*). Regarding the presence of *Leptospira* in the environment, 133 soil samples and 136 river water samples were analyzed.

Most of the studies used serological techniques to detect exposure (n = 11), while only three studies identified *Leptospira* with molecular methods, allowing the identification of the infecting species. A total of 16 different serovars (Australis, Autumnalis, Ballum, Bataviae, Bratislava, Canicola, Cophenhageni, Cynopteri, Gripotyphosa, Hardjo, Icterohaemorrhagiae, Pomona, Pyrogenes, Sejroe, Tarassovi, and Wolffi) were identified by microscopic agglutination test (MAT). The most common serovars were Canicola and Icterohaemorrhagiae, which were detected in seven of the eight studies that provided

serovar information. A total of eight Leptospira species were identified (L. borgpetersenii, L. inadai, L. interrogans, L. kirschnerii, L. licerasiae, L. noguchii, L. santarosai, and L. wolffii). Reported positivity ranged from three to 100%, being above 50% in 10 of the 16 studies that provided this information. Interestingly, although there are only published data from three horses, all of them were seropositive, and it was the species that presented titers $\geq 1:200$ for more different serovars of all the species investigated. Finally, almost all human studies (n = 7/8) included clinical signs (mainly fever). In contrast, no animal studies included clinical signs and only two associated the presence of Leptospira antibodies with kidney lesions confirmed by necropsy. Information extracted from the studies that met the inclusion criteria is summarized in Table-1 [12, 13, 41–56].

Discussion

A recent epidemiological analysis of regional data showed that leptospirosis is a major public health problem in Latin America and that Ecuador is among the countries with the highest incidence rate per capita.

Table-1: Selected studies about Leptospira findings and leptospirosis cases in Ecuador.							
Region (province)	Species (N)	Sample (Assay*)	Positive (%)	Clinical signs	Leptospira species**	<i>Leptospira</i> serovars***	References
Coast	B. taurus (27)	Urine (PCR)	74	-	1, 2, 4, 8	-	[12]
(Esmeraldas)		Urine (PCR)	70	-	2	-	
	<i>H. sapiens</i> (464)	Blood (PCR)	40.7	-	1, 2, 8	-	
	<i>Rattus</i> spp. (6)	Kidney (PCR)	100	-	1, 2, 4, 8	-	
	S. domesticus (27)		66.6	-	1, 2	-	
Coast	<i>C. familiaris</i> (30)		70	-	2	-	[13]
(Manabí)	<i>H. sapiens</i> (464)	Blood (PCR)	40.7	Fever	1, 2, 8	-	
	<i>Rattus</i> spp. (6)	Kidney (PCR)	100	-	1, 2, 4, 8	-	
	S. domesticus (27)	· · ·	66.6	-	1, 2	-	
	S. domesticus	Urine (PCR)	21.1	-	1, 3, 5, 7, 8	-	
	(128)						
Galapagos	Z. wollebaeki (7)	Kidney/Placenta	71.4		Leptospira	-	[41]
(Galapagos)		(PCR)			spp.		
Coast	Water (136)	Water (PCR)	3.6	-	Leptospira spp.		[42]
(Guayas)	C. porcellus (1)	Blood (MAT)	100	-	-	4, 11	
	C. familiaris (4)	Blood (MAT)	100	-	-	6, 9, 10, 11	
	C. aequatorialis (1)	· · ·	100	-	-	4, 6, 7, 8, 10	
	E. caballus (3)	Blood (MAT)	100	-	-	1, 2, 4, 6, 7, 8, 9,	
			100			10, 11, 12, 14, 15	
	L. lagotrichia (1)	Blood (MAT)	100	-	-	4, 6, 8, 9, 10, 11, 16	
	L. tigrinus (1)	Blood (MAT)	100	-	-	2, 6, 7, 8, 10, 11,	
	M ====(2)	Dlaad (MAT)	100			12, 14, 16	
	N. nasua (2)	Blood (MAT)	100	-	-	1, 2, 4, 6, 7, 9, 10,	
	N aliveran (1)	Dlaad (MAT)	100			11, 12, 14, 16	
	N. olivacea (1)	Blood (MAT)	100	-	-	2, 6, 7, 10	
	O. cuniculus (3)	Blood (MAT)	100	-	-	4, 6, 9, 10, 11	
	O. aries (3)	Blood (MAT)	100	-	-	6, 9, 10, 11	
	P. leo (3) S. domesticus (3)	Blood (MAT)	100 100	-	-	6, 7, 8, 10, 12, 14	
Andes	S. domesticus (3)	· · ·	0	-	-	4, 6, 10	[43]
(Pichincha)	S. domesticus (1)	Sperm (PCR)	33	_	- 7	-	[43]
		Post-sperm (PCR)	100	-	7	-	
		Kidney (PCR)	100	-	7	-	
		Epididymis (PCR)	0	-	-	-	
		Testicles (PCR)	100	-	7	_	
Coast	<i>B. taurus</i> (2)	Urine (GS)	- 100	_	3	_	[44]
(Manabí)	H. sapiens (1)	Blood (GS)	_	Fever	7	_	[++]
Coast	B. taurus (320)	Blood (MAT)	50.9	kidney	-	5, 6, 7, 9, 10, 11,	[45]
(Manabí)	<i>D. taulus</i> (520)		50.5	lesions		12, 16	[43]
Coast	H. sapiens (1)	Blood (MAT)	100	Fever	_	3, 11	[46]
(Guayas)			100	I CVCI		5, 11	[10]
-	H. sapiens (2)	Blood (Culture)	100	Fever	Leptospira spp.	-	[47]
Coast	<i>B. taurus</i> (854)		57.4	-	-	5, 6, 7, 9, 10, 11,	[48]
(Manabí)						12, 16	[]
Amazon	H. sapiens (272)	Blood (ELISA)	14.7	Fever	-		[49]
(Pastaza)							
Coast	Soil (133)	Soil (PCR)	24	-	Leptospira spp.	-	[50]
(Manabí)	Water (136)	Water (PCR)	3.6	-	Leptospira spp.		
Coast	B. taurus (72)	Urine (PCR)	13.8	-		-	[51]
(Manabí)							
Àmazon	H. sapiens (216)	Blood (ELISA)	50	-	-	-	[52]
(Morona)							-
Coast	<i>B. taurus</i> (749)	Blood (MAT)	56.2	-	-	5, 6, 7, 9, 10, 11, 12,	[53]
(Manabí)	. ,					16	-
Coast	H. sapiens (2)	Blood (ELISA)	100	Fever	-	-	[54]
(Manabí)							
Coast	S. domesticus	Blood (MAT)	18.9	-	-	1, 4, 6, 10, 11, 14,	[55]
(Manabí)	(280)					15, 16	
Coast	S. domesticus	Blood (MAT)	16.5	Kidney	-	1, 5, 6, 9, 11, 12,	[56]
(Manabí)	(200)			lesions		13, 15	

Table-1: Selected studies about Leptospira findings and leptospirosis cases in Ecuador.

*Assay: ELISA=Enzyme linked immunosorbent assay, GS=Genome sequencing, HE=Hematoxylin-Eosin, MAT=Microscopic agglutination test, ME=Macroscopic examination, PCR=Polymerase chain reaction; ***Leptospira* species: 1=*L. borgpetersenii*, 2=*L. inadai*, 3=*L. interrogans*, 4=*L kirschnerii*, 5=*L. licerasiae*, 6=*L. noguchii*, 7=*L. santarosai*, and 8=*L. wolffii*; ***Leptospira serovars: 1=Australis, 2=Autumnalis, 3=Ballum, 4=Bataviae, 5=Bratislava, 6=Canicola, 7=Copenhageni, 8=Cynopteri, 9=Grippotyphosa, 10=Hardjo, 11=Icterohaemorragiae, 12=Pomona, 13=Pyrogenes, 14=Sejroe, 15=Tarassovi, and 16=Wolffi. *B. taurus=Bos taurus, C. familiaris=Canis familiaris, H. sapiens=Homo sapiens, S. domesticus=Sus domesticus, Z. wollebaeki=Zalophus wollebaeki, C. porcellus=Cavia porcellus, C. aequatorialis=Cebus aequatorialis, E. caballus=Equus caballus, L. lagotrichia=Lagothrix lagotricha, L. tigrinus=Lentinus tigrinus, N. nasua=Nasua nasua, N. olivacea=Nasuella olivacea, O. cuniculus=Oryctolagus cuniculus, O. aries=Ovis aries, P. leo=Panthera leo.* However, the authors also highlight that not all regions have robust surveillance systems, and that more animal studies are needed to better understand the current epidemiological situation [9]. In fact, the latest reviews on animal leptospirosis in Latin America show no records from Ecuador [35, 57]. Our systematic review is the first to compile and analyze data on the human, animal, and environmental epidemiology of *Leptospira*, focusing on horses as possible sentinel species.

We found that most of the studies reported data obtained from one of the four regions (Coast), and specifically from only one of the 24 Provinces of Ecuador (Manabí), which evidence a large information bias at the geographical level. Furthermore, only the studies focused on humans included clinical signs of leptospirosis and, although humans and animals may be exposed to the pathogen through contact with a contaminated environment, there is only one study that analyzed the presence of Leptospira spp. in water or soil as a risk factor for pathogen transmission. A one-health approach, which integrates the human-animal-environment interface, is the ideal framework to better understand and fight leptospirosis. Prevention and control measures should be approached from this perspective, but a major limiting factor has been the lack of communication and cooperation between the human and animal health-care communities [35, 58, 59]. In Ecuador, human leptospirosis is considered by the Epidemiology Department of the Ministry of Public Health a notifiable disease, with an annual incidence ranging between 0.27 and 2.45 cases/100,000 inhabitants, and a fatality rate of 3.06% [60]. However, to the best of our knowledge, there is no government data on morbidity and mortality patterns in domestic or wild animals.

Direct and indirect evidence suggest that leptospirosis is largely widespread among wildlife in Latin America [35, 57]. Surprisingly, our review shows that only two studies included samples from wild mammals. Denkinger et al. [41] found pathogenic Leptospira spp. in 71.4% of Galapagos sea lions, and Orlando et al. [42] found 100% seropositivity in common woolly monkeys, Ecuadorian white-fronted capuchin, Lion, Mountain coati, Oncilla, and Ringtailed coati. In addition, two studies [12, 13] included synanthropic rodents of the genus Rattus, showing a positivity of 3% and 100%, respectively. The small number of studies on wild species is alarming and requires special attention, given that the role of different animal hosts remains unclear. Something similar occurs with domestic species, since although ten studies that included samples of domestic animals, eight of these specifically studied cattle and/or pigs. In addition, there was a great variation in the number of individuals evaluated between species, with only three animals studied in the case of horses to more than 2000 cows. Therefore, our findings reflect the scarce information on leptospirosis in domestic animals in the region.

Leptospira's capacity to infect multiple hosts creates surveillance challenges but also provides opportunities to collect data from animal species that can be used to detect risks to human health. In developing countries, where data on the epidemiology of infectious agents is still lacking, the use of sentinel species can provide essential disease baseline data while enhancing cost-effective surveillance [39, 40]. Members of the order Carnivora are predators and/or scavengers that feed on a wide range of species, exposing themselves to various pathogens. Wild and domestic carnivores (especially dogs) have been proposed as good sentinels for epidemiological surveillance of leptospirosis [60-66]. In Ecuador, most wild carnivores are listed as threatened [67], which limits sampling opportunities due to regulatory and logistical constraints. Regarding domestic carnivores, there are vaccines against leptospirosis authorized for use in dogs, and the vaccination status may alter the serological results of the inoculated animals [68, 69]. In addition, there is no updated national registry of domestic dogs, which makes the epidemiological interpretation of results even more difficult. However, no vaccines against Leptospira are authorized for use in horses, and a national equine registry would allow longitudinal studies to be conducted to evaluate changes in the epidemiology of the disease. Serological surveys in many countries worldwide have shown that horses are exposed to a wide range of serovars, inferring complex epidemiology depending on climatic conditions and the presence of maintenance hosts. For example, 15 different serovars were detected in Switzerland [18], 16 in Ethiopia [20], 12 in Korea [24], 21 in South Africa [31], 20 in Australia [32], 16 in Brazil [70] and 17 in Poland [71]. Therefore, we agree with previous studies by Tsegay et al. [20], Wangdi et al. [32], and Dewes et al. [33] that horses could be a useful sentinel species to assess the occurrence of Leptospira regionally, allowing their use in Ecuador to avoid sampling bias and possible errors in the interpretation of results.

In our review, only Orlando et al. [42] investigated the seroprevalence of *Leptospira* in three horses and detected anti-Leptospira antibodies in all the specimens tested. The panel of antigens used for the MAT contained 21 serovars representing six Leptospira species (L. borgpetersenii serovars: Castellonis, interrogans Javanica. Tarassovi; L. serovars: Australis, Autumnalis, Bataviae, Bratislava, Canicola, Copenhageni, Hardjo, Hebdomadis, Pomona, Pyrogenes, Icterohaemorrhagiae, and Wolfii, Sejroe; L. kirschneri serovars: Cynopteri, Grippotyphosa; L. noguchii serovar: Panama; L. santarosai serovars: Shermani; and L. biflexa serovar: Patoc). Of the 21 serovars used, 12 were detected in hoses. The major serovars, in order of their decreasing seroprevalence, were Bataviae (100%), Gripotyphosa (100%), Icterohaemorrhagiae (100%), Autumnalis (67%), Canicola (67%), Cynopteri (67%), Hardjo (67%), Australis (33%), Cophenhageni (33%), Pomona (33%), Sejroe (33%), and Tarassovi (33%). All three horses presented titers ≥1:200 for more than one serovar. This study also included sera from seven domestic and 6 wild species. Although the low number of horses tested does not allow for conclusive considerations, the species presented reactivity to more different serovars. Serologic reactions to multiple serovars are common in horses; the reasons why a serum reacts with several serovars may be a cross-reaction or a coinfection with more than one serovar [18, 24]. The serovar providing but the highest antibody titer could be an infecting serovar. However, all serovars showing an antibody titer rising of at least four-fold should not be excluded from the list of suspected infecting serovars. On the other hand, low titers could indicate cross-reactivity between the serovars used in MAT [31, 72]. Accordingly, we cannot corroborate the susceptibility of horses to different local Leptospira serovars, since the authors detected anti-Leptospira antibodies by a single MAT, in addition to the fact that co-infections can only be confirmed by molecular typing tools [73]. Although the manuscript was unpublished at the time of the review, Orlando et al. [74] found 100% positivity in 108 horses from the coast, but samples were also analyzed by a single MAT using a panel of six serovars (Bataviae, Bratislava, Canicola, Grippotyphosa, Sejroe, and Tarassovi).

The reviewed studies reported a total of 16 Leptospira serovars in Ecuador based on antibody detection by MAT. This is considered the reference assay for diagnosing leptospirosis, and the information obtained on the infecting serovars is valuable from an epidemiologic standpoint. However, some intrinsic limitations of the MAT could lead to misinterpretations of the results obtained. Laboratories must include panels of Leptospira spp. with all locally circulating serovars because an incomplete panel should be responsible for false negative results [72, 75]. In previously un-surveyed areas, there is no guarantee that the panel is complete, and antibodies to unknown serovars may be missed in serological studies. In contrast, the identification and inclusion of new local serovars in the antigen MAT panels have revealed an increase in the prevalence of leptospirosis [76]. In cases, where serological tests are expected to be of poor sensitivity, incorporating molecular methods may be more appropriate. Molecular techniques can be used for leptospirosis surveillance and source tracking [73, 77]. We found no research identifying the serovars in the region, and a limited number of studies relied on a molecular diagnosis. Our research group was the first to report a pathogenic Leptospira species from the reproductive system of an asymptomatic boar by PCR and amplicon sequencing [43]. However, the authors also highlight that the need for bacteriological isolation of local serovars is critical for increasing the accuracy of MATs. Therefore, due to the limitations described above, the results provided in this review could suggest that there may be local leptospiral cycles that are not yet fully understood, and the results should be interpreted with great caution due to possible bias.

Conclusion

The lack of studies linking the interrelationships between people, animals, and the environment, combined with the limitations of the serological techniques used, show that the epidemiology of *Leptospira* is not yet well established in Ecuador. Considering the country's socio-economic challenges, future research should explore the use of sentinel species such as the horse, which would allow the development of longitudinal studies, increasing cost-efficiency surveillance. These studies should include a combination of serological and molecular diagnostic techniques to characterize the diversity of leptospira present in different bioclimatic regions of the country.

Authors' Contributions

EAD: Conceptualization, methodology, writing original draft, and editing. GA, CS, and LM: Methodology and data compilation. VB: Methodology, data compilation, writing review, and supervision. All authors have read, reviewed, and approved the final manuscript.

Acknowledgments

We are grateful to Agencia de Regulación y Control Fito y Zoosanitario (AGROCALIDAD) for providing the necessary support. Publication of this article was funded by the Universidad San Francisco de Quito Research Publication Fund.

Competing Interests

The authors declare that they have no competing interests.

Publisher's Note

Veterinary World remains neutral with regard to jurisdictional claims in published institutional affiliation.

References

- 1. Karpagam, K.B. and Ganesh, B. (2020) Leptospirosis: A neglected tropical zoonotic infection of public health importance-an updated review. *Eur. J. Clin. Microbiol. Infect. Dis.*, 39(5): 835–846.
- 2. Adler, B. and de la Peña Moctezuma, A. (2010) *Leptospira* and leptospirosis. *Vet. Microbiol.*, 140(3–4): 287–296.
- 3. Picardeau, M. (2015) Leptospirosis: Updating the global picture of an emerging neglected disease. *PLoS Negl. Trop. Dis.*, 9(9): e0004039.
- Costa, F., Hagan, J.E., Calcagno, J., Kane, M., Torgerson, P., Martinez-Silveira, M.S., Stein, C., Abela-Ridder, B. and Ko, A.I. (2015) Global morbidity and mortality of leptospirosis: A systematic review. *PLoS Negl. Trop. Dis.*, 9(9): e0003898.
- 5. Abela-Ridder, B., Sikkema, R. and Hartskeerl, R.A. (2010) Estimating the burden of human leptospirosis. *Int. J. Antimicrob. Agents*, 36(Suppl 1): S5–S7.
- 6. Mwachui, M.A., Crump, L., Hartskeerl, R., Zinsstag, J. and

1952.

Hattendorf, J. (2015) Environmental and behavioural determinants of leptospirosis transmission: A systematic review. *PLoS Negl. Trop. Dis.*, 9(9): e0003843.

- Lau, C.L., Smythe, L.D., Craig, S.B. and Weinstein, P. (2010) Climate change, flooding, urbanisation and leptospirosis: Fuelling the fire? *Trans. R. Soc. Trop. Med. Hyg.*, 104(10): 631–638.
- Munoz-Zanzi, C., Groene, E., Morawski, B.M., Bonner, K., Costa, F., Bertherat, E. and Schneider, M.C. (2020) A systematic literature review of leptospirosis outbreaks worldwide, 1970–2012. *Rev. Panam. Salud Pública*, 44(2020): e78.
- Schneider, M.C., Leonel, D.G., Hamrick, P.N., de Caldas, E.P., Velásquez, R.T., Paez, F.A.M., Arrebato, J.C.G., Gerger, A., Pereira, M.M. and Aldihieri, S. (2017) Leptospirosis in Latin America: Exploring the first set of regional data. *Rev. Panam. Salud Pública*, 41(2017): e81.
- Gestal, M.C., Holban, A.M., Escalante, S. and Cevallos, M. (2015) Epidemiology of tropical neglected diseases in Ecuador in the last 20 years. *PLoS One*, 10(9): e0138311.
- Calvopiña, M., Romero-Alvarez, D., Vasconez, E., Valverde-Muñoz, G., Trueba, G., Garcia-Bereguiain, M.A. and Orlando, S.A. (2023) Leptospirosis in Ecuador: Current status and future prospects. *Trop. Med. Infect. Dis.*, 8(4): 202.
- Chiriboga, J., Barragan, V., Arroyo, G., Sosa, A., Birdsell, D.N., España, K., Mora, A., Espín, E., Mejía, M.E., Morales, M., Pinargote, C., Gonzalez, M., Hartskeerl, R., Keim, P., Bretas, G., Eisenberg, J.N.S. and Trueba, G. (2015) High prevalence of intermediate *Leptospira* spp. DNA in febrile humans from urban and rural Ecuador. *Emerg. Infect. Dis.*, 21(2): 2141–2147.
- Barragan, V., Chiriboga, J., Miller, E., Olivas, S., Birdsell, D., Hepp, C., Hornstra, H., Schupp, J.M., Morales, M., Gonzalez, M., Reyes, S., de la Cruz, C., Keim, P., Hartskeerl, R., Trueba, G. and Pearson, T. (2016) High *Leptospira* diversity in animals and humans complicates the search for common reservoirs of human disease in rural Ecuador. *PLoS Negl. Trop. Dis.*, 10(9): e0004990.
- 14. Pinto, P.S., Libonati, H. and Lilenbaum, W. (2017) A systematic review of leptospirosis on dogs, pigs, and horses in Latin America. *Trop. Anim. Health Prod.*, 49(2): 231–238.
- Browne, E.S., Callefe, J.L.R., De Jesus, E.R.S., Zeppelini, C.G., Cremonese, C. and Costa, F. (2022) A systematic review of the geographic distribution of pathogenic *Leptospira* serovars in the Americas, 1930–2017. *An. Acad. Bras. Cienc.*, 94(3): e20201026.
- Toriz-Suarez, O., Pérez-Rivero, J., Herrera-Barragán, A., Torres-Barranca, J. and Lombardero-Goldaracena, G. (2021) Frequency of *Leptospira* spp serovars reported in horses: A literature review. *Abanico Vet.*, 11(2021): 1–16.
- 17. Båverud, V., Gunnarsson, A., Engvall, E.O., Franzén, P. and Egenvall, A. (2009) *Leptospira* seroprevalence and associations between seropositivity, clinical disease and host factors in horses. *Acta Vet. Scand.*, 51(1): 15.
- Blatti, S., Overesch, G., Gerber, V., Frey, J. and Hüssy, D. (2011) Seroprevalence of *Leptospira* spp. in clinically healthy horses in Switzerland. *Schweiz. Arch. Tierheilkd.*, 153(10): 449–456.
- Sohail, M.L., Khan, M.S., Avais, M., Zahoor, M.Y., Ijaz, M., Ullah, A., Fatima, Z., Naseer, O., Khattak, I. and Ali, S. (2016) Seroprevalence of *Leptospira* spp. in horses of distinct climatic regions of Punjab, Pakistan. *J. Equine Vet. Sci.*, 44(2016): 82–89.
- 20. Tsegay, K., Potts, A.D., Aklilu, N., Lötter, C. and Gummow, B. (2016) Circulating serovars of *Leptospira* in cart horses of central and southern Ethiopia and associated risk factors. *Prev. Vet. Med.*, 125(2016): 106–115.
- Bolwell, C.F., Rogers, C.W., Benschop, J., Collins-Emerson, J.M., Adams, B., Scarfe, K.R. and Gee, E.K. (2020) Seroprevalence of *Leptospira* in racehorses and broodmares in New Zealand. *Animals (Basel)*, 10(11):

- Bastiani, M.P., Lovato, L.T., von Laer, A.E., Pötter, L., Rodrigues, R.O., de Souza, B.C., Hermann, G.P., Sangioni, L.A. and de Avila Botton, S. (2021) Occurrence of *Leptospira* spp. and factors associated with the infection in horses from a military contingent in the Rio Grande do Sul State, Brazil. *Braz. J. Vet. Res. Anim. Sci.*, 58(2021): e180884.
- 23. Verma, A., Stevenson, B. and Adler B. (2013) Leptospirosis in horses. *Vet. Microbiol.*, 167(1–2): 61–66.
- 24. Jung, B.Y., Lee, K.W. and Ha, T.Y. (2010) Seroprevalence of *Leptospira* spp. in clinically healthy racing horses in Korea. *J. Vet. Med. Sci.*, 72(2): 197–201.
- 25. Houwers, D.J., Goris, M.G.A., Abdoel, T., Kas, J.A., Knobbe, S.S., van Dongen, A.M., Westerduin, F.E., Klein, W.R. and Hartskeerl, R.A. (2011) Agglutinating antibodies against pathogenic *Leptospira* in healthy dogs and horses indicate common exposure and regular occurrence of subclinical infections. *Vet. Microbiol.*, 148(2–4): 449–451.
- Hamond, C., Martins, G. and Lilenbaum, W. (2012) Subclinical leptospirosis may impair athletic performance in racing horses. *Trop. Anim. Health Prod.*, 44(8): 1927–1930.
- Hamond, C., Martins, G., Lawson-Ferreira, R., Medeiros, M.A. and Lilenbaum, W. (2013) The role of horses in the transmission of leptospirosis in an urban tropical area. *Epidemiol. Infect.*, 141(1): 33–35.
- 28. Tadich, T.A., Tapia, C. and González, D. (2016) Seroprevalence of *Leptospira* spp. in working horses located in the central region of Chile. *J. Equine Vet. Sci.*, 38(2016): 14–18.
- Hamond, C., Martins, G., Bremont, S., Medeiros, M.A., Bourhy, P. and Lilenbaum, W. (2016) Molecular characterization and serology of *Leptospira kirschneri* (serogroup Grippotyphosa) isolated from urine of a mare postabortion in Brazil. *Zoonoses Public Health*, 63(3): 191–195.
- Ellis, W.A (2015) Animal leptospirosis. Leptospira and leptospirosis *Curr. Top. Microbiol. Immunol.*, 387(2015): 99–137.
- Simbizi, V., Saulez, M.N., Potts, A., Lötter, C. and Gummow, B. (2016) A study of leptospirosis in South African horses and associated risk factors. *Prev. Vet. Med.*, 134(2016): 6–15.
- Wangdi, C., Picard, J., Tan, R., Condon, F., Dowling, B. and Gummow, B. (2013) Equine leptospirosis in tropical Northern Queensland. *Aust. Vet. J.*, 91(5): 190–197.
- Dewes, C., Fortes, T.P., Machado, G.B., Pacheco, P.S., Silva, J.P.M., Neto, A.C.P.S., Félix, S.R. and da Silva, E.F. (2020) Prevalence and risk factors associated with equine leptospirosis in an endemic urban area in Southern Brazil. *Braz. J. Dev.*, 6(8): 58380–58390.
- 34. Bharti, A.R., Nally, J.E., Ricaldi, J.N., Matthias, M.A., Diaz, M.M., Lovett, M.A., Levett, P.N., Gilman, R.H., Willing, M.R., Gotuzzo, E. and Vinetz, J.M. (2003) Leptospirosis: A zoonotic disease of global importance. *Lancet Infect. Dis.*, 3(12): 757–771.
- 35. Petrakovsky, J., Bianchi, A., Fisun, H., Nájera-Aguilar, P. and Pereira, M.M. (2014) Animal leptospirosis in Latin America and the Caribbean countries: Reported outbreaks and literature review (2002–2014). *Int. J. Environ. Res. Public Health*, 11(10): 10770–10789.
- Kleemann, J., Koo, H., Hensen, I., Mendieta-Leiva, G., Kahnt, B., Kurze, C., Inclan, D.J., Cuenca, P., Noh, J.K., Hoffmann, M.H., Factos, A., Lehnert, M., Lozano, P. and Fürst, C. (2022) Priorities of action and research for the protection of biodiversity and ecosystem services in continental Ecuador. *Biol. Conserv.*, 265(2022): 109404.
- Tirira, D.G., Brito, J., Burneo, S.F. and Carrera-Estupiñán, J.P. (2022) Mamíferos del Ecuador: Lista Oficial Actualizada de Especies. Asociación Ecuatoriana de Mastozoología, Quito, Ecuador.
- 38. Programa Nacional Sanitario Equino. (2016) Agencia Ecuatoriana de Aseguramiento de la Calidad del Agro.

Available from: https://www.agrocalidad.gob.ec/wp-content/uploads/2020/05/daj-2016144-0201.0037-programaequino.pdf. Accessed 1 Oct, 2023.

- McCluskey, B.J. (2003) Use of sentinel herds in monitoring and surveillance systems. In: Salman, M.D., editor. Animal Disease Surveillance and Survey Systems: Methods and Applications. Blackwell, Iowa, p119–133.
- Halliday, J.E.B. (2010) Animal Sentinel Surveillance: Evaluating Domestic Dogs as Sentinels for Zoonotic Pathogen Surveillance. Doctoral Dissertation. University of Edinburgh, Scotland.
- 41. Denkinger, J., Guevara, N., Ayala, S., Murillo, J.C., Hirschfeld, M., Montero-Serra, I., Fietz, K., Goldstein, T., Ackerman, M., Barragán, V., Cabrera, F., Chavez, C., Dubovi, E.J., Martinez, J. and Trueba, G. (2017) Pup mortality and evidence for pathogen exposure in Galapagos sea lions (*Zalophus wollebaeki*) on San Cristobal Island, Galapagos, Ecuador. J. Wildl. Dis., 53(3): 491–498.
- 42. Orlando, S.A., Perez, A., Sanchez, E., de la Cruz, C., Rugel, O. and Garcia-Bereguiain, M.A. (2020) High seroprevalence of anti-*Leptospira* spp. antibodies in domestic and wild mammals from a mixed use rescue center in Ecuador: Lessons for "One Health" based conservation strategies. *One Health*, 10(2020): 100140.
- Diaz, E.A., Luna, L., Burgos-Mayorga, A., Donoso, G., Guzman, D.A., Baquero, M.I., Pearson, T. and Barragan, V.A. (2022) First detection of *Leptospira santarosai* in the reproductive track of a boar: A potential threat to swine production and public health. *PLoS One*, 17(9): e0274362.
- 44. Barragan, V., Sahl, J.W., Wiggins, K., Chiriboga, J., Salinas, A., Cantos, N.E., Loor, M.N., Intriago, B.I., Morales, M., Trueba, G. and Pearson, T. (2016) Draft genome sequence of the first pathogenic *Leptospira* isolates from Ecuador. *Genome Announc.*, 4(3):16.
- 45. Macías, D.I.B., Goicochea, C.A.B., Ruano, M.P., Ruales, A.P.R., Flores, M.A.F., Loor, L.V., Rodríguez, R.A.J. and Fonseca-Rodríguez, O. (2020) Association between the presence of antibodies against Leptospira and kidney lesions in cattle [Asociación entre la presencia de anticuerpos contra *Leptospira* y lesiones renales en bovinos]. *Rev. Invest. Vet. Peru*, 31(4): e19028.
- Chedraui, P.A. and San Miguel, G. (2003) A case of leptospirosis and pregnancy. *Arch. Gynecol. Obstet.*, 269(1): 53–54.
- 47. Gelman, S.S., Gundlapalli, A.V. and Hale, D. (2002) Spotting the spirochete: Rapid diagnosis of leptospirosis in two returned travelers. *J. Travel Med.*, 9(3): 165–167.
- Macías, D.I.B., Ruano, M.P., Goicochea, C.A.B., Aguayo, M.D.Z., Valencia, H.P.S., Flores, M.A.F., Loor, L.V., Ruales, A.P.R. and Rodriguez, O.F. (2019) Determination of the seroprevalence of *Leptospira* spp. and the main serovars circulating in cattle in the province of Manabí, Ecuador. *Rev. Sci. Tech.*, 38(3): 787–800.
- 49. Manock, S.R., Jacobsen, K.H., de Bravo, N.B., Russell, K.L., Negrete, M., Olson, J.G., Sanchez, J.L., Blair, P.J., Smalligan, R.D., Quist, B.K., Espín, J.F., Espinoza, W.R., MacCormick, F., Fleming, L.C. and Kochel, T. (2009) Etiology of acute undifferentiated febrile illness in the amazon basin of Ecuador. *Am. J. Trop. Med. Hyg.*, 81(1): 146–151.
- Miller, E., Barragan, V., Chiriboga, J., Weddell, C., Luna, L., Jiménez, D.J., Aleman, J., Mihaljevic, J.R., Olivas, S., Marks, J., Izurieta, R., Nieto, N., Keim, P., Trueba, G., Caporaso, J.G. and Pearson, T. (2021) *Leptospira* in river and soil in a highly endemic area of Ecuador. *BMC Microbiol.*, 21(1): 17.
- 51. Revelo, A., De La Torre, E., Martínez, G., Baquero, M. and Casart, Y. (2020) Evaluation of genomic DNA extraction methods for the identification of *Leptospira* spp. in bovine urine samples by PCR [Evaluación de métodos de extracción de ADN genómico para laidentificación de

*Leptospir*a spp. en muestras de orina bovina mediante por PCR]. *Rev. Invest. Vet. Per*u, 31(2): e15522.

- Romero-Sandoval, N., Cifuentes, L., León, G., Lecaro, P., Ortiz-Rico, C., Cooper, P. and Martín, M. (2019) High rates of exposures to waterborne pathogens in indigenous communities in the Amazon region of Ecuador. *Am. J. Trop. Med. Hyg.*, 101(1): 45–50.
- Ruano, M.P., Macías, D.I.B., Goicochea, C.A.B., Aguayo, M.D.Z., Valencia, H.P.S., Flores, M.A.F., Loor, L.A.V., Ruales, A.P.R. and Fonseca-Rodríguez, O. (2020) Seroprevalence and risk factors of bovine leptospirosis in the province of Manabí, Ecuador. *Comp. Immunol. Microbiol. Infect. Dis.*, 72(28): 101527.
- Valarezo-Sevilla, D. and Sarzosa-Terán, V. (2014) Leptospirosis: Case-series report in a prison of the coast in Ecuador. *Rev. Esp. Sanid. Penit.*, 16(1): 20–23.
- 55. Gavilanes, M.P.Z., Goycochea, C.B., Pérez, L.L., Duarte, R.F. and Pozo, J.L.C. (2021) Renal lesions associated to *Leptospira* spp. seroprevalence in pigs at Portoviejo Slaughterhouse [Lesiones renales asociadas a la seroprevalencia de *Leptospira* spp. en cerdos del matadero de Portoviejo]. *Rev. Prod. Ani*m., 33(3): 39–53.
- 56. Gavilanes, M.P.Z., Pérez, L.L., Santana, M.V.G., Moreir, T.I.V., Loor, L.E.V., Mejía, R.R.V., Duarte, R.F., Goycochea, C.B. and Cuenca, J.C.C. (2020) Seroprevalence of antibodies against *Leptospira* spp. in pigs raised in Portoviejo, Ecuador [Seroprevalencia de anticuerpos contra *Leptospira* spp. en cerdos criados en Portoviejo, Ecuador]. *Rev. Cubana Med. Tro*p., 72(3): 540.
- 57. Vieira, A.S., Pinto, P.S. and Lilenbaum, W. (2018) A systematic review of leptospirosis on wild animals in Latin America. *Trop. Anim. Health Prod.*, 50(2): 229–238.
- Chappel, R.J. and Smythe, L.D. (2012) Leptospirosisimportance of a one health approach. *Microbiol. Aust.*, 33(4): 154–156.
- Schneider, M.C., Jancloes, M., Buss, D.F., Aldighieri,S., Bertherat, E., Najera, P., Galan, D.I., Durski, K. and Espina, M.A. (2013) Leptospirosis: A silent epidemic disease. *Int. J. Environ. Res. Public Health*, 10(12): 7229–7234.
- Calvopiña, M., Vásconez, E., Coral-Almeida, M., Romero-Alvarez, D., Garcia-Bereguiain, M.A. and Orlando, A. (2022) Leptospirosis: Morbidity, mortality, and spatial distribution of hospitalized cases in Ecuador. A nationwide study 2000–2020. *PLoS Negl. Trop. Dis.*, 16(5): e0010430.
- Millán, J., Candela, M.G., López-Bao, J.V., Pereira, M., Jiménez, M.A. and León-Vizcaíno, L. (2009) Leptospirosis in wild and domestic carnivores in natural areas in Andalusia, Spain. *Vector Borne Zoonotic Dis.*, 9(5): 549–554.
- 62. Millán, J., García, E.J., Oleaga, A., López-Bao, J.V., Llaneza, L., Palacios, V., Candela, M.G., Cevidanes, A., Rodríguez, A. and León-Vizcaino, L. (2014) Using a top predator as a sentinel for environmental contamination with pathogenic bacteria: The Iberian wolf and leptospires. *Mem. Inst. Oswaldo Cruz*, 109(8): 1041–1044.
- 63. Sasmal, I., Gould, N.P., Schuler, K.L., Chang, Y.F., Thachil, A., Strules, J., Olfenbuttel, C., Datta, S. and DePerno, C.S. (2019) Leptospirosis in urban and suburban American black bears (*Ursus americanus*) in western North Carolina, USA. *J. Wildl. Dis.*, 55(1): 74–83.
- 64. Roquelo, C., Kodjo, A., Marié, J.L. and Davoust, B. (2021) Serological and molecular survey of *Leptospira* spp. infections in wild boars and red foxes from Southeastern France. *Vet. World*, 14(4): 825–828.
- 65. Balboni, A., Mazzotta, E., Boniotti, M.B., Bertasio, C., Bellinati, L., Lucchese, L., Battilani, M., Ceglie, L., Marchione, S., Esposito, G. and Natale, A. (2022) Outbreak of *Leptospira borgpetersenii* serogroup Sejroe infection in kennel: The role of dogs as sentinel in specific environments. *Int. J. Environ. Res. Public Health*, 19(7): 3906.
- Pilau, N.N., Lubar, A.A., Daneji, A.I., Mera, U.M., Magaji, A.A., Abiayi, E.A., Chaiboonma, K.L., Busayo, E.I., Vinetz, J.M. and Matthias, M.A. (2022) Serological and

molecular epidemiology of leptospirosis and the role of dogs as sentinel for human infection in Nigeria. *Heliyon*, 8(5): e09484.

- 67. Tirira, D.G. (2021) Lista Roja de los Mamíferos del Ecuador. 3rd ed. Asociación Ecuatoriana de Mastozoología, Fundación Mamíferos y Conservación, Pontificia Universidad Católica del Ecuador y Ministerio del Ambiente, Agua y Transición Ecológica del Ecuador, Quito, Ecuador.
- Miller, M.D., Annis, K.M., Lappin, M.R. and Lunn, K.F. (2011) Variability in results of the microscopic agglutination test in dogs with clinical leptospirosis and dogs vaccinated against leptospirosis. J. Vet. Intern. Med., 25(3): 426–432.
- Martin, L.E.R., Wiggans, K.T., Wennogle, S.A., Curtis, K., Chandrashekar, R. and Lappin, M.R. (2014) Vaccine-associated *Leptospira* antibodies in client-owned dogs. *J. Vet. Intern. Med.*, 28(3): 789–792.
- Hashimoto, V.Y., Gonçalves, D.D., da Silva, F.G., de Oliveira, R.C., Alves, L.A., Reichmann, P., Muller, E.E. and de Freitas, J.C. (2007) Occurrence of antibodies against *Leptospira* spp. in horses of the urban area of Londrina, Paraná, Brazil. *Rev. Inst. Med. Trop. São Paulo*, 49(5): 327–330.
- 71. Arent, Z.J. (2013) Seroprevalence study of leptospirosis in horses in Northern Poland. *Vet. Rec.*, 172(10): 269.

- Chirathaworn, C., Inwattana, R., Poovorawan, Y. and Suwancharoen, D. (2014) Interpretation of microscopic agglutination test for leptospirosis diagnosis and seroprevalence. *Asian Pac. J. Trop. Biomed.*, 4(Suppl 1): S162–S164.
- Musso, D. and La Scola, B. (2013) Laboratory diagnosis of leptospirosis: A challenge. J. Microbiol. Immunol. Infect., 46(4): 245–252.
- 74. Orlando, A., Paez-Martínez, K., Sanchez, E., de la Cruz, C., Arcos, F., Torres, P. and García-Bereguiain, M.A. Ultrahigh seroprevalence of anti-*Leptospir*a spp. antibodies in racing horses from a breeding farm in Ecuador: Need for one health-based management. Under review.
- 75. Goris, M.G.A. and Hartskeerl, R.A. (2014) Leptospirosis serodiagnosis by the microscopic agglutination test. *Curr. Protoc. Microbiol.*, 32(1): 12E.5.1–12E.5.18.
- 76. Mgode, G.F., Machang'u, R.S., Mhamphi, G.G., Katakweba, A., Mulungu, L.S., Durnez, L., Leris, H., Hartskeel, R.A. and Belmain, S.R. (2015) *Leptospira* serovars for diagnosis of leptospirosis in humans and animals in Africa: Common *Leptospira* isolates and reservoir hosts. *PLoS Negl. Trop. Dis.*, 9(12): e0004251.
- 77. Guernier, V., Allan, K.J. and Goarant, C. (2018) Advances and challenges in barcoding pathogenic and environmental *Leptospira*. *Parasitology*, 145(5): 595–607.
